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# The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2023; 12(5): 257-264 © 2023 TPI www.thepharmajournal.com Received: 22-04-2023 Accepted: 25-05-2023

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### Influence of nematodes infesting mushrooms (Agaricus bisporus)

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#### Abstract

Mushroom nematodes are parasitic roundworms that can infect and damage the roots of mushroom crops. These nematodes are a major threat to mushroom growers and can cause significant economic losses. There are several species of mushroom nematodes, including *Aphelenchoides composticola* and *Aphelenchus avenae*, which are commonly found in mushroom compost and casing soil. Infected mushrooms may exhibit symptoms such as stunted growth, abnormal morphology, and reduced yields. Management strategies for mushroom nematodes include cultural, physical, chemical and biological methods. Early detection and effective management are essential to minimize the impact of mushroom nematodes on mushroom production. Furthermore, to manage nematode populations, the use of chemical pesticides can have negative environmental and health impacts. Researchers and growers are investigating different control strategies, such as the use of biocontrol agents including predatory nematodes, fungal organisms, and bacteria, to solve this issue. These agents can help reduce nematode populations in soil and protect mushroom crops without the negative impacts associated with chemical pesticides. Although the present generation of mushroom producers continues to face challenges from the nematodes, continued research and development activities offer hope for more long-lasting and efficient control methods in the future.

Keywords: Agaricus bisporus, nematodes, mushroom, mycophagous, saprophagous

#### Introduction

A mushroom is the fruiting body of a fungus that is fleshy, spore-bearing (Basidiomycota, Agaricomycetes), and edible that grows on dead and decaying organic matter (Manoharachary, 2022) <sup>[39]</sup>. Although numerous species have been discovered, they can be divided into three main categories: medicinal mushrooms, edible mushrooms and poisonous mushrooms. Mushroom contains high protein and low calorific value so adding them to our daily diet is beneficial (Nongthombam et al., 2021)<sup>[49]</sup>. It has a high vitamin B, copper, and selenium concentration, is low in salt, and contains over 90% water and relative to 1% fat. (Kratika 2018) <sup>[37]</sup>. In addition to their unique flavour, mushrooms are a meal that people adore because of the health benefits they provide. It is sold in various forms namely as fresh body, preserved/dried form, as pickles or even as canned foods which can be later made into soups, sauces, etc. The growing popularity of mushroom cultivation and processing is due in part to its high income and low-cost input, as well as its rich nutritional content and other positive health effects. (Gupta et al., 2018) [23]. Mushrooms contain bioactive substances that can have a positive impact on one's health, including phenolic compounds, steroids, terpenes, polysaccharides, and vitamins. Mushrooms also contain macronutrients, micronutrients, and a few bioactive substances that have medical use. Due to their immunological properties, the chemicals extracted from mushrooms are highly significant in pharmaceutical, cosmetic, neurological and therapeutic purposes in preventing and treating diseases (table 1) (Sen et al. 2021) <sup>[59]</sup>. However, mushroom cultivation is affected by many diseases (bacterial, viral and fungal), pests and nematodes (Okigbo and Anuagasi 2021)<sup>[50]</sup>. In this review paper, nematode infestation in button mushrooms is being highlighted.

Sl.no.	Properties and contents	References
1.	substances with bioactive properties such as polysaccharides, steroids, terpenes, and phenolic compounds	Kim et al. 2015 <sup>[35]</sup> ;
		Royse 2014 <sup>[57]</sup> ;
		Shang <i>et al.</i> 2015 <sup>[61]</sup> ;
		Sheu et al. 2007 [63];
		Mariga <i>et al</i> . 2014 <sup>[40]</sup>
2.	Vitamin B, vitamin C and vitamin D	Panjikkaran and Mathew 2013 <sup>[51]</sup>
3.	20-35% dry-weight protein and 9 essential amino acids	Kalac 2009 <sup>[27]</sup>
4.	Antigenotoxic	Wang et al. 2005 [71]
5.	Antioxidative	Roupas et al. 2012 [56]
6.	Antiproliferative	Zhou et al. 2013 [75]
7.	Tumorigenic	Kim et al. 2015 <sup>[35]</sup>
8.	Antihypertensive	Vaz et al. 2011 [69]
9.	Hypocholesterolaemic	Han et al. 2011 [24]
10.	Stress-reducing properties	Akata <i>et al.</i> 2012 <sup>[2]</sup>
11	Appetite suppressant	Kim et al. 2011 [34]

#### Taxonomy of Button Mushroom (Agaricus bisporus)

In India, two types of button mushrooms are cultivated i.e., white button mushroom (commonly grown) and brown button mushroom (Amin *et al.* 2021)<sup>[4]</sup>. *Agaricus bisporus*, the most commonly cultivated mushroom species, accounts for around 40% of global production. (Banik 2010)<sup>[6]</sup>. In contrast to seasonal growers who only grow them in the winter, commercial units grow button mushroom crops throughout the year. The most cultivated states are Maharashtra, Punjab, Haryana, Gujarat, Goa, Uttarakhand, and Himachal Pradesh. (Nihal 2022)<sup>[48]</sup>. *Agaricus bisporus* is a common edible mushroom that belongs to the kingdom Fungi, phylum Basidiomycota, class Agaricomycetes, order Agaricales, and family Agaricaceae (El-Sebaaly *et al.*,2021)<sup>[12]</sup> (Table 2)

Table 2: Taxonomic classification

Kingdom	Fungi	
Division	Basidiomycota	
Class	Agaricomyctes	
Order	Agaricales	
Family	Family Agaricaceae	
Genus Agaricus		
Species	bisporous	

#### **Mushroom-Nematodes interaction**

Mushroom cultivation, one of the employment-generating sources, is famous or popular mainly amongst marginal farmers, young entrepreneurs and housewives (Borah *et al.* 2019) <sup>[10]</sup>. Mushroom cultivation and processing require a hygienic place to carry out and large industries have pasteurization units to sanitize. But these marginal farmers, young entrepreneurs and housewives do not have this pasteurization unit. Numerous pests, such as nematodes, fungi, bacteria, insects, as well as biotic and abiotic factors, are attracted to this unsanitary situation, which lowers the quality and yield of the mushroom. As a result of this improper sanitization, nematode infestation is a popular biotic stress which affects mushroom cultivation and causes crop losses (Keshari, and Kranti 2020) <sup>[29]</sup>.

Mushroom nematodes are parasitic roundworms that attack mushroom crops. They are considered harmful to the mushroom crop and affect the quality of mushroom production (Zhang *et al.* 2020) <sup>[74]</sup>. They are mostly found in raw compost such as poultry manure, wheat straw, horse manure, etc. They developed mainly because of unhygienic, improper pasteurization and not properly prepared compost

and casing material (Keshari, and Kranti 2020) [29]. In mushroom cultivation, unhygienic conditions provide a favourable environment for many pests, diseases and nematode infestation. Nematodes carry the greatest threat of all because once a crop has been infested by nematodes, there is no coming back meaning that the crop has to be fully eradicated (Phani et al. 2021) [52]. So, nematode infestation causes the greatest yield loss or total crop failure in mushroom cultivation as the crop could not be saved. In this context, a study conducted by Seth in 1984 [76] at Solan revealed that there was extremely less harvest in the beginning, resulting in total crop failures due to 84.4% nematode infestation. The reason behind the fast infestation of nematodes is that they reproduce very quickly by adapting themselves to the biological requirements of the crops and thus causing crop losses of up to 100% (Nagesh and Reddy. 2000) <sup>[45]</sup>. Bhardwaj et al. (1972) <sup>[8]</sup> published the first report about nematode damage mushrooms in India from Himachal Pradesh. With the growth in the mushroom industry for the last two decades, there have been numerous reports of nematode-caused crop failure. (Thapa et al. 1977; Chhabra & Kaul 1982; Seth 1984) [68, 12, 76]

Nematodes are of 2 types:

1. Myceliophagus or mycophagous nematodes

2. Saprophagous nematodes.

#### Myceliophagus or mycophagous

Myceliophagus or mycophagous nematodes feed on the mycelium of the mushroom fungus. This type of nematode uses a bear-shaped stylet to penetrate the mycelium and drain out its contents, thus leaving the mycelium dead and nonfunctioning for further growth of the mushroom body (Reddy and Reddy 2021) <sup>[54]</sup>. In some cases, even if the mycelium grows into a mushroom body, these Mycophagous nematodes go up the pinhead or pilus of the mushroom and attack (Godfrey 2003) <sup>[20]</sup>. The mushroom body shows stunted growth, the colour changes and eventually it dies. Mycophagous nematode infestation is hazardous as once this nematode enters the mushroom unit, it is very hard to eradicate. Also, they serve as carriers of several bacterial (eg. Pseudomonas tolaasii), viral, and fungal diseases (Maurya et al. 2020) [42]. Mycophagous nematodes have short lifespans, high fecundity, are highly pathogenic and are a destructive pest of mushrooms but they are found rarely due to proper sanitization in modern cultivation (Sattar et al. 2021)<sup>[58]</sup>.

Table 3: Some	Myceliophagus	or mycophagous	nematodes
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Myceliophagus or mycophagous nematodes	References	
Aphelenchus avenae	Hesling 1979 <sup>[25]</sup>	
Aphelenchoides composticolaBajaj and Walia (1999) <sup>[5]</sup> , Khanna and Sharma (1988a, b) <sup>[30</sup> Gitanjali and Nandal (2001a, b) <sup>[18, 19]</sup>	Bajaj and Walia (1999) <sup>[5]</sup> , Khanna and Sharma (1988a, b) <sup>[30, 31]</sup> ,	
	Gitanjali and Nandal (2001a, b) <sup>[18, 19]</sup>	
Aphelenchoides agarici	Seth and Sharma (1986) <sup>[60]</sup> , Bajaj and Walia (1999) <sup>[5]</sup> , Seth (1984) <sup>[76]</sup>	
Aphelenchoides neocompositicola Seth and Sharma (1986) <sup>[60]</sup>		
Ditylenchus Myceliophagus Sharma et al. (1981) <sup>[62]</sup> , Thapa et al. (1981) <sup>[67]</sup>		

#### Saprophagous nematodes

Unlike mycophagous nematodes, Saprophagous nematodes have a tubular mouthpart with which they penetrate and withdraw the nutrient content of the mycelium (Marjanović 2022) <sup>[41]</sup>. While the mycophagous nematodes leave the mycelium to die, the saprophagous nematodes only injure the mycelium with their toxins and do not cause any direct damage (Keshari, and Kranti 2020)<sup>[29]</sup>. They usually affect the mushroom only after the infestation of the mycophagous nematodes. They reduce the quality of the compost beds making it unfavourable for the growth of mushrooms and also impacting the spawn run. They multiply very fast and are found in large numbers. Their population number depends on the quality of pasteurization of the compost beds (Sattar et al. 2021) <sup>[58]</sup>. Some symptoms usually seen are distortion, kidney-shaped mushrooms, gills colour changed into violet and sometimes sporophores turning brown in colour. These signs and symptoms are seen as a result of the nematode infestation in the mushroom beds. They carry the live bacteria in their gut and release them in the compost beds thereby inhibiting the growth of the fungal mycelium. (Keshari and Kranti 2020)<sup>[29]</sup>.

Some species of saprophagous nematodes are *Rhabditis spp.*, *Diplogaster spp.*, *Caenorhabditis elegans*, *Panagrolaimus spp. Like Panagrolaimus fuchsia* (Nagesh and Reddy 2000) [45].

#### Source of Nematode infestation

During the entire cultivation process, the button mushroom is extremely vulnerable to nematode attack. Damp wheat straw, poultry manures, casing soil, farm yard manure (FYM), garden soil, spent compost, platform soil, irrigation water, composting with contaminated water, and contaminated utensils are the major sources of nematode infestation (Bellettini et al. 2018) [7]. Nematodes can be moved across beds by flies, especially sciarids (Navarro et al.2021)<sup>[47]</sup>. Sometimes nematodes are contaminated through diseased compost and soil. It is also spread from infected to noninfected areas via equipment, clothing, and employees' hands. Once the roof is infested, the crop is affected by nematode larvae droppings or eggs in the water droplets condensing on the roof. Another main reason for the nematode infestation is the lack of awareness. As mushroom cultivation is practiced mainly by marginal farmers, housewives and young entrepreneurs, they are not fully aware of the need for sterilization and pasteurization and thus, the higher rate of nematode infestation. A study conducted by Fletcher and Gaze (2007) <sup>[14]</sup> states that nematodes feed on fungi and in this particular study, they found that the presence of a particular fungus species, Arthrobotrys spp. shows the high nematode contamination in the mushroom bed. The presence of this fungus can be seen in the brown fungal colonies in the compost bed or casing soil.

## Symptoms of nematodes infestation on *Agaricus bisporus* mushroom

Nematodes are microscopic worm-like organisms that can infect the mycelium of *Agaricus bisporus* mushrooms and cause damage to the crop (Zhang *et al.* 2020)<sup>[74]</sup>. If nematode infestation is suspected, it is important to take appropriate measures to prevent the spread of the infestation to other crops (Khanna *et al.* 2004)<sup>[33]</sup>. This may involve removing infected mushrooms and using nematode-resistant strains of *Agaricus bisporus* in future crops. Nematode infestations on *Agaricus bisporus* mushrooms can cause a variety of symptoms, including:

- 1. **Stunted growth**: Nematodes can cause the mycelium of the mushrooms to grow more slowly than usual, resulting in stunted growth and smaller mushroom caps.
- 2. **Reduced yields**: As a result of the slower growth, nematode-infected mushroom crops may produce lower yields than healthy crops and sometimes 100% crop failure.
- 3. **Poor quality**: Nematode-infected mushrooms may have a lower average weight, a higher proportion of malformed mushrooms, and other quality issues that make them less desirable for sale or consumption.
- 4. **Yellowing of mycelium**: In some cases, nematode infestations can cause the mycelium of the mushrooms to turn yellow or brown, indicating that the nematodes are feeding on the mycelium.
- 5. **Distorted mushroom caps**: Nematodes can also cause the caps of the mushrooms to become distorted or misshapen, which can affect their appearance and marketability.
- 6. **Reduced shelf life**: Nematode-infected mushrooms may have a shorter shelf life than healthy mushrooms, due to their lower quality and increased susceptibility to decay.

It is significant to remember that additional causes might potentially contribute to these symptoms, such as poor growing conditions or other pests and diseases, so growers should conduct a proper diagnosis to confirm nematode infestation. Growers can seek assistance from plant pathologists or nematologists to conduct tests and recommend appropriate control measures (Fletcher and Gaze 2007)<sup>[14]</sup>.

#### Negative interaction of nematodes on mushroom

Nematodes can have a negative interaction with mushrooms and are often considered a pest in mushroom cultivation (Zhang *et al.* 2020)<sup>[74]</sup>. Nematodes may attack and feed on the mushroom's mycelium, which may delay any development of the mycelium and lower the crop's overall yield (Rinker 2017)<sup>[55]</sup>. In some cases, nematodes can also feed on the developing mushroom caps, causing them to become distorted or misshapen. In addition to direct damage, nematodes can also transmit other pathogens to the mushroom culture, such as bacteria or fungi, which can further damage the mycelium and decrease the crop's quality and yield (Ahmad et al. 2021) <sup>[2]</sup>. Nematodes can create entry points for other pests and pathogens by causing wounds in the mycelium or weakening the overall health of the mushroom culture. They can enter mushroom cultures through contaminated substrates or other materials used in the cultivation process. They can also be introduced through contaminated water or equipment (Bellettini et al. 2018) [7]. Once they enter the culture, they can move through the soil or growing media, seeking out the mycelium of the mushrooms to feed on. To prevent nematode attacks on mushrooms, growers should take several measures, including using nematode-free substrates and materials, sterilizing growing media before use, maintaining good hygiene practices in the cultivation environment, and monitoring the growing environment for signs of nematode activity. In addition, some growers may choose to use nematicides or other control measures to treat nematode infestations (Tapia-Vázquez et al. 2022) [66].

#### Positive interaction of nematodes on mushroom

While nematodes are often considered a pest in mushroom cultivation, there are some instances where they can have a positive interaction with mushrooms. Mushroom nematodes are parasites that infect and damage mushroom crops, and as such, they are generally considered to have negative effects. However, there are many beneficial uses of mushrooms themselves, such as providing food, medicine, and ecological services. For example, some types of mushrooms are known to have immune-boosting, anti-inflammatory, and anti-cancer qualities. Some nematodes form mutually beneficial associations with mycorrhizal fungi, which are fungi that form symbiotic relationships with the roots of most plants, including mushrooms (Yuvaraj and Ramasamy 2020)<sup>[73]</sup>. The nematodes help the mycorrhizal fungi by breaking down organic matter and releasing nutrients that the fungi can use to grow and spread, while the fungi provide the nematodes with a source of food in the form of sugars and other nutrients. This mutualistic relationship benefits both the nematodes and the mushrooms, as it enhances nutrient cycling and can contribute to their overall health and growth (Carrasco and Preston 2020) [11]. Some species of nematodes can act as decomposers, breaking down organic matter in the soil or growing media and releasing nutrients that can be taken up by the mushrooms and used for growth (Adegbeye et al. 2020) <sup>[1]</sup>. This helps in recycling nutrients and making them available for mushroom growth, thereby positively influencing mushroom growth and productivity. This can improve the overall health and growth of the mushroom culture (Naik. et al. 2019) [46]. Additionally, there is potential for using certain species of nematodes as biocontrol agents against other pests and diseases that can affect mushroom cultivation, which may negatively impact mushroom growth. Some nematodes are known to feed on bacteria or other fungi that compete with mushrooms for resources, helping to reduce competition and promote the growth of mushrooms. Certain species of nematodes can act as biocontrol agents against pests that can attack mushrooms. Some nematodes are known to parasitize and kill insect pests, such as mushroom flies or mites, that can damage mushrooms (Rinker 2017) <sup>[55]</sup>. By reducing the population of pests, nematodes can help protect mushrooms from damage and promote healthy mushroom growth. Using these nematodes as a natural control measure

could reduce the need for chemical pesticides and improve the sustainability of mushroom cultivation (Soliman et al.2022). Furthermore, nematodes can also stimulate mycelial growth, which is the vegetative part of mushrooms, by feeding on certain substances or by physically breaking up mycelial mats (Kamalakannan et al.2020) [28]. This can result in increased mycelial growth and branching, which in turn can lead to enhanced mushroom fruiting. Overall, nematodes can have positive interactions with mushrooms by forming mutualistic associations, preying on competing organisms, contributing to nutrient recycling, acting as biocontrol agents, and stimulating mycelial growth, all of which can promote healthy mushroom growth and productivity (Zhang et al. 2020)<sup>[74]</sup>. However, it's important to note that the effects of nematodes on mushrooms can vary depending on the specific species of nematodes, mushrooms, and environmental conditions, and more research needed to better understand these is interactions (Koppenhöfer et al. 2020)<sup>[36]</sup>.

#### **Integrated Management of Mushroom Nematodes**

Nematodes can get controlled by using various methods like cultural, physical, biological and chemical methods (WA *et al.* 2021) <sup>[70]</sup>. Although nematicides are commonly used for the prevention of nematodes as they affect the environment and human health, scientists are researching more for alternatives to control nematodes that are not harmful (Gamalero and Glick 2020) <sup>[16]</sup>. Nematode entry into the beds should be prevented rather than controlled because of the crop's nature, the shortage of nematicides that are both safe and effective, residue issues, and other nematicide dangers (Reddy and Reddy 2021) <sup>[54]</sup>. There is currently no curative measure that can be used during the cropping stage. As a result, an integrated approach is used as the sole control measure.

#### 1. Cultural measures

In mushroom cultivation, cropping should be done in specially constructed mushroom chambers with enough ventilation, using properly pasteurized compost produced through the short method because sometimes in the long method of composting, the level of contamination is higher (Fletcher and Gaze 2007)<sup>[14]</sup>. Rigorous sanitary and sanitation measures must be implemented throughout the cropping season. Everything has to be sterilized with 2% formalin 24 hours before composting. Also, treatment with 4% formalin is required for all equipments, floors, walls, and galleries. (Kamalakannan et al. 2020)<sup>[28]</sup>. Disinfectants (5% formalin concentration) should be used by both employees and guests before entering and after leaving the mushroom chamber (Keshari and Kranti 2020) <sup>[29]</sup>. Ingredients for composting should always be kept in clean surroundings. Mushroom farms should be kept clean both inside and outside. Good sanitation practices such as removing and destroying infested mushroom substrates, cleaning and disinfecting equipment, and keeping the growing area clean can help to reduce nematode populations (Okigbo and Anuagasi 2021)<sup>[50]</sup>. The casing combination should be adequately pasteurized. Manures intended for composting should be properly decomposed and given adequate time to decompose. The growth rooms should have sufficient ventilation and doors and windows enclosed with wire mesh measuring 14 to 16 cm per layer to avoid the entry of insect pests. (Singh and Sharma 2016) <sup>[64]</sup> The mushroom growers can also practice crop

rotation, a cultural control measure that involves alternating mushroom cultivation with non-host crops. This helps to break the life cycle of the nematodes, thereby reducing their population (Wesemael *et al.* 2011) <sup>[72]</sup>. Another technique is Soil solarization which involves covering the mushroom bed with a clear plastic sheet and exposing it to sunlight for several weeks. This technique helps to kill nematodes, bacteria, and other soil-borne pathogens that can affect mushroom growth (Munshi *et al.* 2010) <sup>[44]</sup>. And finally, maintaining the Soil pH is as important as the other practices in the cultural control of mushroom nematodes. Mushroom nematodes thrive in acidic soils. Maintaining a neutral to slightly alkaline soil pH of 7-7.2 can help to reduce nematode populations (Keshari and Kranti 2020) <sup>[29]</sup>.

#### 2. Chemical control

Since mushrooms are grown indoors, the use of pesticides is limited (Reddy and Reddy 2021)<sup>[54]</sup>. Also, it is usually eaten fresh so the use of pesticides will be hazardous to human health. But when using a long composting technique, some of the chemicals that can be used can help control nematode

populations. (Keshari and Kranti 2020) [29]. Organochlorine insecticides were the first class of chemicals utilized to eradicate pests that affect mushrooms (Read 1968) [53]. Additionally utilized as nematicides are organophosphates. When mixed with the compost and applied as sprays during the spawn run, thionazin at 80 ppm (0.008%) effectively controlled D. Myceliophagus and A. composticola without leaving any hazardous residue. (Singh and Sharma 2016)<sup>[64]</sup>. The most effective method for controlling A. composticola and Rhabditis sp. was Dichlorvos at 0.04% administered for 3–4 days under polythene cover. (Keshari and Kranti 2020) <sup>[29]</sup> (table 4). Lack of awareness about the use of nematicides or pesticides in mushroom cultivation and processing is one of the main causes of decline in yield, insect resistance, selfharm to the mushroom growers, contaminated mushrooms and serious health issues (Reddy and Reddy 2021)<sup>[54]</sup>. In this modern world, where sustainable agriculture and natural farming are being encouraged, the most effective way to eliminate mushroom nematodes is farm hygiene and sanitization.

Table 4: Some nematicides/ pesticides used for mushroom nematodes

Sl. No.	Nematicides/ pesticides	Nematode species	References
1.	Thionazin at 80 ppm	D. myceliophagus and A. composticola	Singh and Sharma 2016) <sup>[64]</sup> .
2.	Dichlorvos (0.04%)	A. composticola and Rhabditis sp.	Keshari and Kranti 2020 <sup>[29]</sup>
3.	Phenamiphos (EC) at a dose of 20 mg/kg	Aphelenchoides composticola	Mcleod and Khair 2008 <sup>[43]</sup>
4.	diflubenzuron	Aphelenchoides spp.	Gahukar 2014 [15]
5.	benomyl and thiabendiazole	Aphelenchoides spp.	Mcleod and Khair 1978 <sup>[43]</sup>
6.	Carbofuran	Myceliophagus and saprophagous nematodes	Sharma <i>et al.</i> 1981 <sup>[62]</sup>

#### 3. Physical Control

Physical control measures for mushroom nematodes involve the use of physical barriers or traps to prevent or reduce the spread of nematodes (Georgis et al. 2006) [17]. Soil steaming is a physical control measure that involves using steam to heat the mushroom bed and soil to high temperatures. This helps to kill nematodes and other soil-borne pests (Fletcher and Gaze 2007) <sup>[14]</sup>. In order to control nematodes in the mushroom industry, heat is the most effective method. In order to eradicate compost nematodes, the pasteurization room's air and bed temperatures must be maintained for at least two hours at 60°C, and the incubation temperature must be maintained for five to six hours at 70°C or for thirty to sixty minutes at 80°C (Thapa et al. 1981) [68]. Disinfecting used equipments such as trays and utensils with 5% formalin or cresylic acid or by using boiling water for 1-2 minutes is ideal (Nagesh and Reddy 2000)<sup>[12]</sup>. Soil solarization, which was mentioned in the cultural control section, is also a physical control measure that involves covering the mushroom bed with a clear plastic sheet and exposing it to sunlight for several weeks. This technique helps to kill nematodes and other soil-borne pathogens that can affect mushroom growth (Munshi et al. 2010)<sup>[44]</sup>. Traps can also be used to reduce the population of mushroom nematodes. For example, sticky traps can be placed around the mushroom bed to trap and kill nematodes (Grewal 2000)<sup>[22]</sup>. Physical barriers such as mesh screens can be used to prevent nematodes from entering the mushroom bed or transfer nematodes from one bed to the other.

#### 4. Biological control

Natural enemies or other species, such as fungi and bacteria and their byproducts, are used in biological management to reduce mushroom nematodes as mushrooms are typically consumed fresh and the use of nematodes or pesticides is not advised (Keshari and Kranti 2020) [29]. Because of their extreme sensitivity to chemicals, fresh mushrooms shouldn't be consumed after being sprayed with a nematicide. Consequently, the importance of biocontrol agents has increased. Two nematophagous fungi, Arthrobotrys irregularis and Candelalretta musiformis, were shown to be very effective in suppressing nematode multiplication on mushroom mycelium, according to a study conducted in Solan (Himachal Pradesh). (Khanna and Sharma 1990; Grewal and Sohi 1988) <sup>[32, 21]</sup>. A wide range of plants has been found to have nematicidal properties. Another fungus known as Royal 300, Arthrobotrys robusta, is effective against mycophagous nematodes. It has been found that Candelalretta musiformis extracted from used compost is extremely good at preventing nematode multiplication (Keshari and Kranti 2020)<sup>[29]</sup>. Another fungus known as A. irregularis is incredibly successful at eliminating A. composticola, claim Grewal and Sohi (1988) [21] and Khanna and Sharma (1988a, b) <sup>[30, 31]</sup>. However, when growing mushrooms, one should only utilize biocontrol agents found in compost, such as predatory nematodes and nematophagous fungi, which are easy to grow and multiply since they are facultative parasites and they can consume both saprophytic bacteria and nematodes (Kumar et al. 2020)<sup>[28]</sup>. In order to manage plant infections, such as mushroom nematodes, biocontrol agents are increasingly being employed as a sustainable and environmentally friendly substitute to chemical pesticides (Soliman et al. 2022) [65]. Predatory nematodes, fungus, and bacteria are some of the biocontrol agents that have been identified to control mushroom nematodes. In general, the use of biocontrol agents in the

control of mushroom nematodes shows promise as a sustainable and effective approach to reducing nematode populations and protecting mushroom crops (Bilgrami 2008)<sup>[9]</sup>. Further study is required regarding integrated pest management strategies that combine biocontrol with other control techniques and to optimise the use of these agents.

#### Conclusion

In India, mushroom growing is a relatively young and expanding industry. The demand for producing mushrooms is increasing as people become more aware of their taste and high food value. Because mushrooms are a short-term crop, using insecticides to kill nematodes is not recommended owing to health hazards. There is a need to investigate the utilization of some plant materials that exhibit nematicidal qualities while still being safe for mushroom mycelium. Nematode issues in mushrooms are distinct in that the nematodes have fully adapted to the crop's ecological needs and also reproduce incredibly quickly, leading to crop losses of up to 100%. An integrated approach is used to control nematodes. In conclusion, mushroom nematodes pose a significant threat to mushroom growers, causing damage to mushroom crops and leading to economic losses. Chemical pesticides have traditionally been used to control nematode populations, but these can have negative environmental and health impacts. Alternative methods of managing mushroom nematodes, like the use of biocontrol agents, are more environmentally responsible and sustainable. Further study is required regarding integrated pest management strategies that combine biocontrol with other control techniques and to optimise the use of these agents. Overall, it is important for mushroom growers to remain vigilant in monitoring for nematode infestations and to implement effective control measures to minimize the impact of nematodes on mushroom production. By doing so, growers can protect their crops and ensure a more sustainable and profitable mushroom industry.

#### References

- 1. Adegbeye MJ, Reddy PRK, Obaisi AI, Elghandour MMMY, Oyebamiji KJ, Salem AZM, *et al.* Sustainable agriculture options for production, greenhouse gasses and pollution alleviation, and nutrient recycling in emerging and transitional nations-An overview. Journal of Cleaner Production. 2020;242:118319.
- Ahmad G, Khan A, Khan AA, Ali A, Mohhamad HI. Biological control: A novel strategy for the control of the plant parasitic nematodes. Antonie van Leeuwenhoek. 2021 Jul;114(7):885-912.
- Akata I, ERGÖNÜL B, Kalyoncu F. Chemical compositions and antioxidant activities of 16 wild edible mushroom species grown in Anatolia. International Journal of Pharmacology. 2012, 8(2).
- Amin Z, Wani FF, Gulzar H, Dar WA, Sheikh PA. Diseases of White Button mushroom (*Agaricus bisporus*)-A potential threat to mushroom industry. Int. J Curr Microbiol App Sci. 2021;10(02):2076-85.
- Bajaj HK, Walia KK. Three new species of Aphelenchoides Fisher, 1894 (Aphelenchina: Nematoda) from India along with a compendium of Indian species. Indian Journal of Nematology. 1999;29(2):154-70.
- 6. Banik S. Mushrooms: The magic store of health benefits. Everyman's Sci. 2010;64:360–365.
- 7. Bellettini MB, Bellettini S, Fiorda FA, Pedro AC, Bach

F, Fabela-Morón MF, *et al.* Diseases and pests noxious to Pleurotus spp. mushroom crops. Revista Argentina de microbiologia. 2018 Apr 1;50(2):216-226.

- 8. Bhardwaj AK, Sharma PL, Thakur JR. Mushroom nematodes. Indian Bot. Reptr. 1972;42:257–258.
- 9. Bilgrami AL. Biological control potentials of predatory nematodes. Integrated management and biocontrol of vegetable and grain crops nematodes, 2008, 3-28.
- Borah TR, Singh AR, Paul P, Talang H, Kumar B, Hazarika S. Spawn production and mushroom cultivation technology. ICAR research complex for NEH region, 2019, 46.
- 11. Carrasco J, Preston GM. Growing edible mushrooms: a conversation between bacteria and fungi. Environmental microbiology. 2020;22(3):858-872.
- 12. Chhabra HK, Kaul VK. Occurrence and control of *Aphelenchoides composticola* on mushrooms. Indian Journal of Nematology. 1982;12(1):191-2.
- 13. El-Sebaaly Z, Hammoud M, Sassine YN. History, health benefits, market, and production status of button mushroom. In *Mushrooms: Agaricus bisporus*. Wallingford UK: CABI; c2021, 1-65.
- 14. Fletcher JT, Gaze RH. Mushrooms: Pest and Disease Control: A Colour Handbook. Manson Publishing Ltd.: London, 2007, 192.
- 15. Gahukar RT. Mushroom pest and disease management using plant-derived products in the Tropics: A Review. International Journal of Vegetable Science. 2014 Jan 2;20(1):78-88.
- 16. Gamalero E, Glick BR. The use of plant growthpromoting bacteria to prevent nematode damage to plants. Biology. 2020;9(11):381.
- 17. Georgis R, Koppenhöfer AM, Lacey LA, Bélair G, Duncan LW, Grewal PS, *et al.* Successes and failures in the use of parasitic nematodes for pest control. Biological Control. 2006;38(1):103-123.
- 18. Gitanjali Nandal SN. Nematodes associated with white button mushroom (*Agaricus bisporus*) in Sonipat district of Haryana. Indian J Nematol. 2001a;31:79–98
- 19. Gitanjali Nandal SN. Effect of neem products and dazomet for the management of *Aphelenchoides composticola* on white button mushroom (*Agaricus bisporus*) under semicommercial conditions. Indian J Nematol. 2001b;31:52-57.
- 20. Godfrey SA. Molecular investigation of pseudomonads causative of *Agaricus bisporus* blotch disease in New Zealand mushroom farms; c2003.
- 21. Grewal PS, Sohi HS. A new and cheaper technique for rapid multiplication of Arthrobotrys conoides and its potential as a boi-nematicide in mushroom culture. Current science. 1988 Jan 5;57(1):44-6.
- 22. Grewal PS. Mushroom pests. Field Manual of Techniques in Invertebrate Pathology: Application and evaluation of pathogens for control of insects and other invertebrate pests, 2000, 497-503.
- 23. Gupta S, Summuna B, Gupta M, Annepu SK. Edible mushrooms: cultivation, bioactive molecules, and health benefits. Bioactive molecules in food. 2018;1:1-33.
- 24. Han EH, Hwang YP, Kim HG, Choi JH, Im JH, Yang JH, *et al.* Inhibitory effect of *Pleurotus eryngii* extracts on the activities of allergic mediators in antigen-stimulated mast cells. Food and Chemical Toxicology. 2011 Jun 1;49(6):1416-25.

The Pharma Innovation Journal

- 25. Hesling JJ. The role of eelworms in culture of mushrooms in UK. Mushroom J. 1979;83:423-429.
- 26. http://agridaksh.iasri.res.in/html\_file/mushroom/20Mush \_nematode\_pests.html
- Kalač P. Chemical composition and nutritional value of European species of wild growing mushrooms: A review. Food chemistry. 2009 Mar 1;113(1):9-16.
- 28. Kamalakannan A, Syamala M, Sankar PM, Shreedevasena MS, Ajay MB. Mushrooms–A hidden treasure; c2020.
- 29. Keshari N, Kranti KVVSK Integrated Management of Phytopathogenic Nematodes Infesting Mushroom. Springer Nature Singapore Pte Ltd. 2020;7:141-170.
- 30. Khanna AS, Sharma NK. Pathogenicity of various *Aphelenchoides* species on *Agaricus bisporus*. Indian Phytopathol. 1988a;41:472–473.
- Khanna AS, Sharma NK. Effect of population levels and time of infestation of *Aphelenchoides agarici* on mycelial growth of *Agaricus bisporus*. Nematol Mediterr. 1988b;16:125-127.
- 32. Khanna AS, Sharma NK. Efficacy of nematophagous fungi against Myceliophagus nematodes *in vitro*. In: Proceedings of national seminar on bio-agents in nematode management, New Delhi, IARI, 1990, 83-84.
- 33. Khanna PK, Sodhi HS, Kapoor S. Diseases of *Agaricus bisporus* and their management. Annual Review of Plant Pathology. 2004;2(2):163-205.
- 34. Kim K, Choi B, Lee I, Lee H, Kwon S, Oh K, Kim AY. Bioproduction of mushroom mycelium of *Agaricus bisporus* by commercial submerged fermentation for the production of meat analogue. Journal of the Science of Food and Agriculture. 2011 Jul;91(9):1561-8.
- 35. Kim SH, Jakhar R, Kang SC. Apoptotic properties of polysaccharide isolated from fruiting bodies of medicinal mushroom *Fomes fomentarius* in human lung carcinoma cell line. Saudi journal of biological sciences. 2015 Jul 1;22(4):484-90.
- Koppenhöfer AM, Shapiro-Ilan DI, Hiltpold I. Entomopathogenic nematodes in sustainable food production. Frontiers in Sustainable Food Systems. 2020 Aug 20;4:125.
- Kratika Sharma. Mushroom: Cultivation and Processing. International Journal of Food Processing Technology. 2018 Dec 20;5(2):9-12.
- Kumar R, Keshari N. Occurrence of Nematophagous fungi in the fresh and spent button mushroom compost in Bihar. Journal of Pharmacognosy and Phytochemistry. 2020;9(3):119-23.
- Manoharachary C. Edible and Medicinal Mushrooms: Some Aspects and Prospects. In Biology, Cultivation and Applications of Mushrooms. Singapore: Springer Singapore; 2022, 259-284.
- 40. Mariga AM, Yang WJ, Mugambi DK, Pei F, Zhao LY, Shao YN. Antiproliferative and immunostimulatory activity of a protein from *Pleurotus eryngii*. Journal of the Science of Food and Agriculture. 2014 Dec;94(15):3152-62.
- 41. Marjanović L. A. bisporus food web reconstruction and nematode analysis by stable isotope probing (Master's thesis); c2022.
- Maurya AK, Murmu R, John V, Kesherwani B, Singh M. Important Diseases and Pests of Mushroom. Agriculture & Food: e-Newsletter. December 2019-2020;1(2). e-

ISSN: 2581 – 8317

- 43. Mcleod RW, Khair GT. Control of *Aphelenchoides composticola* in mushroom compost with nematicides. Annals of Applied Biology. 1978 Jan;88(1):81-88.
- 44. Munshi NA, Dar GH, Ghani MY, Kauser S, Mughal N. Button mushroom cultivation. Communication and Publication Centre SKUAST, 2010, 1-28.
- 45. Nagesh M, Reddy P. Status of mushroom nematodes and their management in India, Integrated Pest Management Reviews. 2000;5:213–224.
- 46. Naik K, Mishra S, Srichandan H, Singh PK, Sarangi PK. Plant growth promoting microbes: Potential link to sustainable agriculture and environment. Biocatalysis and Agricultural Biotechnology. 2019 Sep 1;21:101326.
- Navarro MJ, Escudero-Colomar LA, Carrasco J, Gea FJ. Mushroom Phorid Flies—A Review. Agronomy. 2021 Oct;11(10):1958.
- 48. Nihal A. Mushroom Consumption as a Proactive Approach for Improving Human Health: A Review. Current Journal of Applied Science and Technology. 2022 Apr 2;41(5):17-25.
- Nongthombam J, Kumar A, Ladli BG, Madhushekhar M, Patidar S. A review on study of growth and cultivation of oyster mushroom. Plant Cell Biotechnology and Molecular Biology. 2021 Feb 11;22(5&6):55-65.
- Okigbo RN, Anuagasi CL. Diseases Affecting Mushrooms in Africa. Journal of Food Technology & Nutrition Sciences. SRC/JFTNS/143. DOI: doi. org/10.47363/JFTNS/2021. 2021;3(129):2-10.
- 51. Panjikkaran ST, Mathew D. An environmentally friendly and cost effective technique for the commercial cultivation of oyster mushroom [*Pleurotus florida* (Mont.) Singer]. Journal of the Science of Food and Agriculture. 2013 Mar 15;93(4):973-6.
- 52. Phani V, Khan MR, Dutta TK. Plant-parasitic nematodes as a potential threat to protected agriculture: Current status and management options. Crop Protection. 2021 Jun 1;144:105573.
- 53. Read WM. Pesticides for mushroom. MGA Bull. 1968;194:77-82.
- 54. Reddy PP, Reddy PP. Mushrooms. Nematode Diseases of Crops and their Management, 2021, 325-335.
- 55. Rinker DL. Insect, mite, and nematode pests of commercial mushroom production. Edible and medicinal mushrooms: Technology and applications, 2017, 221-237.
- 56. Roupas P, Keogh J, Noakes M, Margetts C, Taylor P. The role of edible mushrooms in health: Evaluation of the evidence. Journal of functional foods. 2012 Oct 1;4(4):687-709.
- 57. Royse DJ. A global perspective on the high five: Agaricus, Pleurotus, Lentinula, Auricularia & Flammulina. In Proceedings of the 8th International Conference on Mushroom Biology and Mushroom Products (ICMBMP8) 2014 Nov 19 (Vol. 1, pp. 1-6).
- 58. Sattar MN, Iftikhar S, Saeed F, Shaalan R, Hourani W, Bejjani R. Mushroom Farm Design, Management of Pests, and Control of Diseases. Mushrooms: Agaricus bisporus. 2021 Oct 6;37:297.
- 59. Sen P, Kosre A, Koreti D, Chandrawansi NK, Jadhav SK. Nutrients and bioactive compounds of Pleurotus ostreatus mushroom. NewBioWorld. 2021 Dec 31;3(2):8-12.
- 60. Seth A, Sharma NK. Five new species of genus

Aphelenchoides (Nematoda: Aphelenchida) infesting mushroom in northern India. Indian Journal of Nematology. 1986;16(2):205-15.

- 61. Shang HM, Song H, Xing YL, Niu SL, Ding GD, Jiang YY, *et al.* Effects of dietary fermentation concentrate of Hericium caput-medusae (Bull.: Fr.) Pers. on growth performance, digestibility, and intestinal microbiology and morphology in broiler chickens. Journal of the Science of Food and Agriculture. 2016 Jan 15;96(1):215-22.
- 62. Sharma NK, Thapa CD, Nath A. Pathogenicity and identity of myceliophagus nematode infesting *Agaricus bisporus* (Lange) Sing. in Himachal Pradesh (India). Indian Journal of Nematology. 1981;11(2):230-1.
- 63. Sheu F, Chien PJ, Wang HK, Chang HH, Shyu YT. New protein PCiP from edible golden oyster mushroom (*Pleurotus citrinopileatus*) activating murine macrophages and splenocytes. Journal of the Science of Food and Agriculture. 2007 Jun;87(8):1550-8.
- 64. Singh UA, Sharma K. Pests of mushroom. Advances in Crop Science and Technology, 2016, 4:1–6
- 65. Soliman G, Elkhateeb W, Wen TC, Daba G. Mushrooms as efficient biocontrol agents against the root-knot nematode, meloidogyne incognita. Egyptian Pharmaceutical Journal. 2022 Jan 1;21(1):68.
- 66. Tapia-Vázquez I, Montoya-Martínez AC, De los Santos-Villalobos S, Ek-Ramos MJ, Montesinos-Matías R, Martínez-Anaya C. Root-knot nematodes (*Meloidogyne spp.*) a threat to agriculture in Mexico: Biology, current control strategies, and perspectives. World Journal of Microbiology and Biotechnology. 2022 Feb;38(2):26.
- 67. Thapa CD, Sharma NK, Nath A. Mushroom nematodes and their sources of contamination in mushroom beds in India. Mushroom Sci. 1981;11:443–448
- 68. Thapa CD, Munjal RL, Seth PK. The mushroom nematodes and their control. Indian J. Mushroom. 1977;3:1-11.
- Vaz JA, Barros L, Martins A, Santos-Buelga C, Vasconcelos MH, Ferreira IC. Chemical composition of wild edible mushrooms and antioxidant properties of their water soluble polysaccharidic and ethanolic fractions. Food Chemistry. 2011 May 15;126(2):610-616.
- 70. WAE, Daba GM, Soliman GM. The anti-nemic potential of mushroom against plant-parasitic nematodes. J Microbiol Biotechnol. 2021;6(1).
- Wang JC, Hu SH, Liang ZC, Lee MY. Antigenotoxicity of extracts from *Pleurotus citrinopileatus*. Journal of the Science of Food and Agriculture. 2005 Apr 15;85(5):770-8.
- 72. Wesemael W, Viaene N, Moens M. Root-knot nematodes (*Meloidogyne spp.*) in Europe. Nematology. 2011 Jan 1;13(1):3-16.
- 73. Yuvaraj M, Ramasamy M. Role of fungi in agriculture. Biostimulants in Plant Science, 2020, 1-12.
- Zhang Y, Li S, Li H, Wang R, Zhang KQ, Xu J. Fungi– nematode interactions: Diversity, ecology, and biocontrol prospects in agriculture. Journal of Fungi. 2020 Oct 4;6(4):206.
- 75. Zhou J, Chen Y, Xin M, Luo Q, Gu J, Zhao M, *et al.* Structure analysis and antimutagenic activity of a novel salt-soluble polysaccharide from *Auricularia polytricha*. Journal of the Science of Food and Agriculture. 2013 Oct;93(13):3225-30.

76. Seth J, Kellett HA, Caldwell G, Sweeting VM, Beckett GJ, Gow SM. A sensitive immunoradiometric assay for serum thyroid stimulating hormone: A replacement for the thyrotrophin releasing hormone test?. Br Med J (Clin Res Ed). 1984 Nov 17;289(6455):1334-6.