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## Biochemical changes in postpartum Bidri goats supplemented with maize alone or with safflower oil and estrus synchronised using CIDR based Protocol

**Prashantkumar, Bijurkar RG, Venkanagouda D, Tandle MK, Suranagi MD and Shrikant Kulkarni**

### Abstract

This study was conducted to find out the effect of supplementation of maize and safflower oil combined with CIDR estrus synchronization protocol on some biochemical parameters during major and minor breeding seasons. Seventy-five postpartum Bidri goats were selected from the day of kidding and divided into 5 groups (n=15). i. control, ii. maize, iii. maize and CIDR, iv. maize and safflower oil and v. maize, safflower oil and CIDR. The blood samples were collected on 0 (on the day of parturition), 15, 22, 30, estrus day and 30 days post mating. Samples were analysed for serum parameters viz., glucose, cholesterol, total protein and albumin. The serum glucose and cholesterol levels showed increasing trend following parturition and reaching high values on d-30 post mating and day of estrus, respectively. The supplementation of only maize or along with safflower oil with or without estrus induction using CIDR did not had any impact. Further, the total protein and albumin did not showed any specific trend or impact of any treatment or supplementation.

**Keywords:** Bidri goats, biochemical parameters, CIDR, estrus synchronization, postpartum period

### 1. Introduction

Goats, the “Poor man’s cow” are known for low cost of rearing and meagre management requirements. They have served the mankind especially the rural poor farmers since prehistoric times. The versatile properties of its various products like milk, meat, hair and skin makes it a reliable and potential source of income. Among the numerous breeds, Bidri goat is mostly reared in the Bidar and Kalaburagi districts of Northern Karnataka region of South India. It is one of the best dual-purpose breed with better milk yield and twinning ability making it as a breed of choice among the farmers. Reproduction is an important phenomenon that determines the income of the livestock farmers and is affected by various factors. Earlier studies have suggested that postpartum constituents of blood are important indicators of biochemical and physiological changes taking place during this critical period (Manat *et al.*, 2016)<sup>[8]</sup>. The biochemical changes are related to the stress condition in animals and are related to the productivity and management practice of the animals (Manat *et al.*, 2016; Soares *et al.*, 2018; Zamunar *et al.*, 2020)<sup>[8, 11, 17]</sup>.

Estrus synchronization is among the most important reproductive management strategy to augment fertility (Stevenson and Britt, 2017)<sup>[15]</sup>. The controlled internal drug release (CIDR) devices containing 0.3 gm of progesterone are currently the preferred method of progesterone administration in goats (Wheaton *et al.*, 1993)<sup>[15]</sup>. The Bidri goats are reared mostly under traditional method and their commercialization is the need of the time. The reproductive performance and estrus synchronisation studies on this breed are scanty. The present study therefore aimed to evaluate biochemical parameters in postpartum does to improve fertility by energy supplementation and estrus synchronization.

### 2. Material and Method

#### 2.1 Ethical approval

The study was conducted duly following approved guidelines from the Institutional Animal Ethics Committee.

#### 2.2 Location of the study

The study was conducted in and around Bidar city and lies between 17°35’ and 18°29’ North

latitude and 76°41' to 77°39' East longitude. The coldest months are December and January, the temperature begins to rise towards the end of February till May which is the hottest month of the year. The temperature varies in between 20°C and 42°C. The year may be divided into summer season from March to May, southwest monsoon season from June to September, post-monsoon season from October to November and winter season from December to February (CGWB, 2008). The complete study was carried out from October 2021 to November 2022. The phase one of the study was conducted between October 2021 to April 2022 and phase two was between May-2022 to November-2022.

### 2.3 Selection of animals

The experimental animals were selected from the flocks of Bidri goats maintained by local farmers (private farms), in and around Bidar district. For the study, seventy-five healthy, post-partum Bidri goats between 1-4 parity were selected and randomly divided into 5 groups of 15 goats each. The goats were reared under a semi-intensive housing system with free grazing for 4-6 hrs daily with *ad libitum* drinking water and routine deworming and vaccinations on regular basis.

### 2.4 Estrus synchronization protocols

The following treatment protocols were repeated for two sequential kidding's on the same goats.

#### 2.4.1 Group 1 (T<sub>0</sub>) - Control

This group was considered as control, the goats in this group were not provided with any additional feed other than grazing.

#### 2.4.2 Group 2 (T<sub>1</sub>) – Maize Feeding

Each animal in this group along with grazing, additionally fed with 250 g of maize grains daily (morning) for 30 days from the day of parturition.

#### 2.4.3 Group 3 (T<sub>2</sub>) – Maize + CIDR

Each animal in this group along with grazing, additionally fed with 250 g of maize grains daily (morning) for 30 days from the day of parturition. On 16<sup>th</sup> day postpartum, Controlled Internal Drug Release (CIDR= 0.3gm of progesterone) devices were inserted into the vagina aseptically with the help of an applicator and left in-situ for 14 days. Then, implants were withdrawn and an intramuscular injection of a synthetic prostaglandin, PGF<sub>2</sub>α (Cloprostenol Sodium) @ 125 µg/doe was administered.

#### 2.4.4 Group 4 (T<sub>3</sub>) – Maize + Safflower Oil Feeding

Each animal in this group were fed 250 g of maize grains daily (morning) along with 50 g of Safflower oil for 30 days from the day of parturition.

#### 2.4.5 Group 5 (T<sub>4</sub>) – Maize + Safflower Oil Feeding + CIDR

Each animal in this group were fed 250 g of maize grains daily (morning) along with 50 g of Safflower oil for 30 days from the day of parturition. On the 16<sup>th</sup> day Controlled Internal Drug Release (CIDR= 0.3gm of progesterone) devices were inserted into the vagina aseptically with the help of an applicator and left inside the vagina for 14 days. Then, implants were withdrawn and an intramuscular injection of a synthetic prostaglandin, PGF<sub>2</sub>α (Cloprostenol Sodium) @ 125 µg/doe was given.

### 2.5. Estrus detection and Mating

The does were observed for estrus signs twice daily at 12-hour interval. The does in estrus were allowed to mate with bucks for naturally.

### 2.6 Blood Collection

Blood samples, collected by jugular vein puncture in the morning hour before feeding, from all the groups on day 0 (day of kidding), 15, 22, 30, on the day of estrus and 30 days post mating. The serum samples were harvested by centrifugation at 3000 rpm for 10 minutes and samples were made into two aliquot and stored at -20°C until analysis.

### 2.7 Serum biochemical analysis

Serum biochemical analysis for glucose (mg/dL), cholesterol (mg/dL), total protein (g/dL) and albumin (g/dL) were performed as per assay procedures mentioned in the kits of Transasia Bio-Medicals Ltd. duly following the procedures as directed by the supplier with semi-automatic biochemical analyser (Microlab-300, Elite Tech Ltd.).

### 2.8 Statistical Analysis

The data obtained was analysed and compared using SAS 9.3 software.

## 3. Results and Discussion

The mean ± SE values of serum biochemical parameters for different groups at different reproductive stages in Bidri goats are presented in Tables 1 to 8.

### Serum glucose

In the first kidding (Phase-I), the serum glucose showed varying levels but was significantly high only on 30 days post-mating compared to parturition day in all the groups, but not in the second kidding. When compared between the groups in both the kidding's, except on the day of estrus and 30-days post-mating, the serum glucose was higher in all treatment groups either significantly (p<0.05, d-22 and d-30) or non-significantly (p>0.05, d-0 and d-15).

The glucose levels are reported to decrease up to day fourteen post-partum and then onwards an increase up to d-45 post-partum (Manat *et al.*, 2016) [8], which was similar to the present study except the decrease in glucose up to 14 day was noted here. Glucose is the primary source of energy for the regulation of multiple reproductive process and reduced blood levels of glucose can affect the follicular growth, ovulation, estrus response (Dupont *et al.* 2014; Sosa *et al.* 2009) [4, 12]. Previously, it has been reported that P4 treatment enhances the glucose level (Samad and Ford 1981) [10]; however, no variation was observed in the present study. All the glucose values were within physiologically normal range as reported for goats (Kaneko *et al.*, 2008) [2].

### Serum Cholesterol

In both the sequential kidding's (phase I & phase II), the serum cholesterol did not vary significantly (p>0.05) either between the groups or between the days except for between the days in phase II. In both the phases, the cholesterol levels showed sequential increase with advancement of post-partum period which was only significant (p<0.05) in all treatment groups on the day of estrus in phase II when compared to day 0. The cholesterol levels were within the normal range which concurs with the findings of 65-136 mg/dl (Mohammed *et al.*,

2016 and Daramola *et al.*, 2005) <sup>[9, 3]</sup> and an increasing trend up to the day of estrus was in agreement with Krajnicakova *et al.* (2003) <sup>[7]</sup> and Witte and Schafer (2007) <sup>[16]</sup> in the puerperal period of goats. In addition, Witte and Schafer (2007) <sup>[16]</sup> stated that cholesterol was precursor for steroid hormones, is needed for early resumption of postpartum ovarian activity and cyclicity. High level of cholesterol during estrus was supported by Ishwar and Pandey (1994) <sup>[6]</sup>, who stated that estrogen influences lipid metabolism through liponeogenesis which in turn causes increased production of cholesterol in endocrine gland tissue from acetate during estrus.

**Serum Total Protein**

In both the sequential kidding’s (phase I & phase II), the proteins levels did not follow a trend except for significantly high level on the day of estrus when compared day-0 in phase I.

The total protein concentration during postpartum period was reported to show slight increase from the day 7<sup>th</sup> up to 21<sup>st</sup> day postpartum but the differences were non-significant (Bhoite *et al.*, 2019) which was not observed in the present study. Few researchers reported that highest level of serum total protein on day 45<sup>th</sup> postpartum in Surti goat (Manat *et al.*, 2016) <sup>[8]</sup> and on day 40<sup>th</sup> postpartum in goat (Krajnicakova *et al.*, 2003) <sup>[7]</sup>, which are closer to our findings on 30 days post mating. Moreover, the physiological stage also influenced the total protein concentration in different species. The lactation period did not influence total protein in Sahel goat (Waziri *et al.*,

2010) <sup>[14]</sup>, which is in agreement with our study findings.

**Serum Albumin**

In both the sequential kidding’s (phase I & phase II), the serum albumin did not vary significantly (p>0.05) either between the groups or between the days except for treatment group 4 where it showed significantly high levels on day-22 when compared to day 0.

The albumin levels are reported to increase up to 14 days postpartum (Manat *et al.*, 2016) <sup>[8]</sup>, which is evident with present study findings and increase in albumin in 1<sup>st</sup> week followed by decrease in 2<sup>nd</sup> week and 3<sup>rd</sup> week after parturition (Iriadam, 2007) <sup>[5]</sup>, which is contrary to present study findings could be because of breed, season and managerial variations. All the albumin values were within the physiological normal range for goats (Kaneko *et al.*, 2008) <sup>[2]</sup>.

**4. Conclusion**

To conclude, the serum glucose and cholesterol levels have increasing trend following parturition and reaching high values on d-30 post mating and day of estrus, respectively. The supplementation of only maize or along with safflower oil with or without estrus induction using CDR did not had any impact. Further, the total protein and albumin also did not showed any specific trend or impact of any treatment or supplementation.

**Table 1:** Mean ± SE values of serum glucose (mg/dL) in different groups at different intervals (Phase 1)

Groups \ Days	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Control (T <sub>0</sub> )	47.40±2.02 <sup>aA</sup>	55.80±1.65 <sup>aA</sup>	53.60±1.97 <sup>aA</sup>	56.20±1.60 <sup>aA</sup>	52.27±1.47 <sup>aA</sup>	67.07±2.58 <sup>bA</sup>
Maize (T <sub>1</sub> )	53.60±1.60 <sup>acA</sup>	61.53±1.11 <sup>abA</sup>	57.13±0.70 <sup>adeAB</sup>	64.07±1.39 <sup>bdAB</sup>	49.53±1.60 <sup>ceA</sup>	64.53±2.32 <sup>bdA</sup>
Maize + CIDR (T <sub>2</sub> )	50.60±2.41 <sup>aA</sup>	60.53±0.90 <sup>bA</sup>	61.47±0.80 <sup>bAB</sup>	57.40±1.66 <sup>abA</sup>	50.53±1.70 <sup>aA</sup>	65.40±1.94 <sup>bA</sup>
Maize + SFO (T <sub>3</sub> )	52.00±1.96 <sup>aA</sup>	62.53±1.49 <sup>bA</sup>	61.80±0.80 <sup>bAB</sup>	66.47±0.98 <sup>bB</sup>	50.07±1.39 <sup>aA</sup>	64.87±2.64 <sup>bA</sup>
Maize + SFO + CIDR (T <sub>4</sub> )	51.40±2.37 <sup>aA</sup>	58.53±1.02 <sup>abA</sup>	65.73±1.48 <sup>bB</sup>	64.93±1.13 <sup>bAB</sup>	49.73±1.56 <sup>aA</sup>	66.13±2.44 <sup>bA</sup>

**Table 2:** Mean ± SE values of serum glucose (mg/dL) in different groups at different intervals (Phase 2)

Groups \ Days	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Control (T <sub>0</sub> )	52.67±1.74 <sup>aA</sup>	53.40±1.46 <sup>aA</sup>	52.47±1.96 <sup>aA</sup>	53.53±2.37 <sup>aA</sup>	49.73±0.79 <sup>aA</sup>	57.40±2.52 <sup>aA</sup>
Maize (T <sub>1</sub> )	51.93±2.83 <sup>aA</sup>	61.67±2.16 <sup>abA</sup>	68.07±2.55 <sup>bB</sup>	65.93±2.68 <sup>bCB</sup>	53.93±1.30 <sup>acDA</sup>	62.33±2.70 <sup>abA</sup>
Maize + CIDR (T <sub>2</sub> )	53.47±2.79 <sup>acA</sup>	62.27±2.00 <sup>abcA</sup>	71.33±2.32 <sup>bcdB</sup>	65.40±2.66 <sup>acAB</sup>	52.93±1.20 <sup>eA</sup>	62.47±2.27 <sup>adeA</sup>
Maize + SFO (T <sub>3</sub> )	52.60±2.82 <sup>aA</sup>	59.93±1.74 <sup>abA</sup>	66.07±2.20 <sup>bB</sup>	70.47±3.11 <sup>bB</sup>	53.07±1.16 <sup>aA</sup>	60.13±2.70 <sup>abA</sup>
Maize + SFO + CIDR (T <sub>4</sub> )	53.13±2.86 <sup>aA</sup>	60.13±2.25 <sup>aA</sup>	65.07±3.08 <sup>abB</sup>	68.67±2.38 <sup>bB</sup>	53.40±1.14 <sup>aA</sup>	61.53±2.42 <sup>abA</sup>

Means with different superscripts differ significantly at P<0.05

<sup>abcdefg</sup> Superscripts indicate the statistical difference in serum glucose values on different time interval within groups.

<sup>ABC</sup> Superscripts indicate the statistical difference in serum glucose values between groups on different time interval.

**Table 3:** Mean ± SE values of serum cholesterol (mg/dL) in different groups at different intervals (Phase 1)

Groups \ Days	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Group 1 (T <sub>0</sub> )	71.40±7.17 <sup>aA</sup>	72.87±8.34 <sup>aA</sup>	84.47±7.20 <sup>aA</sup>	73.73±7.90 <sup>aA</sup>	93.87±6.66 <sup>aA</sup>	104.27±4.48 <sup>aA</sup>
Group 2 (T <sub>1</sub> )	73.87±5.88 <sup>aA</sup>	74.13±6.89 <sup>aA</sup>	83.27±7.75 <sup>aA</sup>	76.33±6.91 <sup>aA</sup>	95.87±5.18 <sup>aA</sup>	103.40±4.44 <sup>aA</sup>
Group 3 (T <sub>2</sub> )	73.80±7.19 <sup>aA</sup>	72.20±6.16 <sup>aA</sup>	85.80±5.47 <sup>aA</sup>	94.80±15.07 <sup>aA</sup>	105.67±8.48 <sup>aA</sup>	108.93±5.24 <sup>aA</sup>
Group 4 (T <sub>3</sub> )	72.27±6.69 <sup>aA</sup>	74.53±5.40 <sup>aA</sup>	86.07±11.16 <sup>aA</sup>	81.53±10.28 <sup>aA</sup>	91.67±6.50 <sup>aA</sup>	106.93±4.63 <sup>aA</sup>
Group 5 (T <sub>4</sub> )	72.80±8.04 <sup>aA</sup>	73.53±5.77 <sup>aA</sup>	83.47±8.91 <sup>aA</sup>	83.60±7.32 <sup>aA</sup>	92.67±7.62 <sup>aA</sup>	108.33±4.82 <sup>aA</sup>

**Table 4:** Mean ± SE values of serum cholesterol (mg/dL) in different groups at different intervals (Phase 2)

Days Groups	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Group 1 (T <sub>0</sub> )	71.53±3.48 <sup>aA</sup>	73.33±4.58 <sup>aA</sup>	77.53±3.53 <sup>aA</sup>	80.53±4.21 <sup>aA</sup>	90.07±3.34 <sup>aA</sup>	86.27±3.89 <sup>aA</sup>
Group 2 (T <sub>1</sub> )	71.20±2.79 <sup>aA</sup>	75.13±4.14 <sup>abA</sup>	79.73±4.70 <sup>abA</sup>	81.27±3.64 <sup>abA</sup>	93.27±2.60 <sup>bA</sup>	87.93±2.50 <sup>abA</sup>
Group 3 (T <sub>2</sub> )	72.47±2.60 <sup>aA</sup>	75.40±4.56 <sup>aA</sup>	79.20±3.78 <sup>abA</sup>	82.00±4.35 <sup>abA</sup>	96.00±3.40 <sup>bA</sup>	90.93±2.89 <sup>abA</sup>
Group 4 (T <sub>3</sub> )	72.13±3.63 <sup>aA</sup>	78.80±3.99 <sup>aA</sup>	80.13±4.04 <sup>acA</sup>	83.13±4.16 <sup>acA</sup>	105.73±2.64 <sup>bA</sup>	98.73±2.08 <sup>bcA</sup>
Group 5 (T <sub>4</sub> )	72.87±3.93 <sup>aA</sup>	80.67±4.00 <sup>aA</sup>	83.40±3.70 <sup>aA</sup>	86.47±2.60 <sup>acA</sup>	118.53±2.71 <sup>bA</sup>	103.60±3.58 <sup>bcA</sup>

Means with different superscripts differ significantly at P<0.05

<sup>abcdefg</sup> Superscripts indicate the statistical difference in serum cholesterol values on different time interval within groups.

<sup>ABC</sup> Superscripts indicate the statistical difference in serum cholesterol values between groups on different time interval.

**Table 5:** Mean ± SE values of serum total protein (g/dL) in different groups at different intervals (Phase 1).

Days Groups	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Group 1 (T <sub>0</sub> )	7.01±0.44 <sup>aA</sup>	6.78±0.36 <sup>aA</sup>	6.69±0.35 <sup>aA</sup>	7.44±0.36 <sup>aA</sup>	10.29±0.78 <sup>bA</sup>	7.89±0.42 <sup>abA</sup>
Group 2 (T <sub>1</sub> )	6.17±0.43 <sup>aA</sup>	7.11±0.37 <sup>aA</sup>	6.68±0.39 <sup>aA</sup>	7.33±0.36 <sup>aA</sup>	10.49±0.73 <sup>bA</sup>	8.45±0.38 <sup>abA</sup>
Group 3 (T <sub>2</sub> )	7.45±0.30 <sup>abA</sup>	6.21±0.43 <sup>aA</sup>	5.95±0.39 <sup>aA</sup>	7.07±0.35 <sup>abA</sup>	8.98±0.64 <sup>bA</sup>	8.43±0.36 <sup>abA</sup>
Group 4 (T <sub>3</sub> )	6.59±0.56 <sup>aA</sup>	7.31±0.42 <sup>aA</sup>	6.13±0.40 <sup>aA</sup>	7.07±0.30 <sup>aA</sup>	10.07±0.77 <sup>bA</sup>	8.37±0.43 <sup>abA</sup>
Group 5 (T <sub>4</sub> )	7.85±0.36 <sup>aA</sup>	7.27±0.43 <sup>aA</sup>	6.46±0.52 <sup>aA</sup>	7.75±0.36 <sup>aA</sup>	10.59±0.66 <sup>bA</sup>	8.31±0.44 <sup>abA</sup>

**Table 6:** Mean ± SE values of serum total protein (g/dL) in different groups at different intervals (Phase 2).

Days Groups	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Group 1 (T <sub>0</sub> )	6.27±0.21 <sup>abA</sup>	5.76±0.20 <sup>aA</sup>	7.42±0.36 <sup>bA</sup>	5.67±0.20 <sup>aA</sup>	6.62±0.27 <sup>abA</sup>	6.88±0.15 <sup>abA</sup>
Group 2 (T <sub>1</sub> )	6.25±0.17 <sup>aA</sup>	5.83±0.17 <sup>aA</sup>	7.90±0.30 <sup>bA</sup>	5.93±0.23 <sup>aA</sup>	7.02±0.24 <sup>aA</sup>	6.93±0.18 <sup>abA</sup>
Group 3 (T <sub>2</sub> )	6.22±0.24 <sup>acA</sup>	5.72±0.26 <sup>aA</sup>	7.89±0.41 <sup>bcA</sup>	5.21±0.19 <sup>aA</sup>	7.32±0.25 <sup>ceA</sup>	7.08±0.19 <sup>bdeA</sup>
Group 4 (T <sub>3</sub> )	5.92±0.27 <sup>adeA</sup>	5.79±0.17 <sup>adA</sup>	7.78±0.23 <sup>bfeA</sup>	5.43±0.35 <sup>aA</sup>	6.85±0.23 <sup>cdefA</sup>	7.20±0.25 <sup>eA</sup>
Group 5 (T <sub>4</sub> )	6.58±0.22 <sup>acdA</sup>	5.61±0.27 <sup>aA</sup>	8.47±0.32 <sup>bcA</sup>	6.05±0.21 <sup>afA</sup>	7.43±0.19 <sup>cegA</sup>	7.11±0.25 <sup>dfgA</sup>

Means with different superscripts differ significantly at P<0.05

<sup>abcdefg</sup> Superscripts indicate the statistical difference in serum total protein values on different time interval within groups.

<sup>ABC</sup> Superscripts indicate the statistical difference in serum total protein between groups on different time interval.

**Table 7:** Mean ± SE values of serum albumin (g/dL) in different groups at different intervals (Phase 1).

Days Groups	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Group 1 (T <sub>0</sub> )	2.73±0.32 <sup>aA</sup>	3.05±0.22 <sup>aA</sup>	3.61±0.32 <sup>aA</sup>	3.69±0.44 <sup>aA</sup>	3.44±0.21 <sup>aA</sup>	2.65±0.42 <sup>aA</sup>
Group 2 (T <sub>1</sub> )	2.67±0.26 <sup>aA</sup>	2.87±0.15 <sup>aA</sup>	3.72±0.44 <sup>aA</sup>	3.45±0.41 <sup>aA</sup>	3.47±0.25 <sup>aA</sup>	2.71±0.41 <sup>aA</sup>
Group 3 (T <sub>2</sub> )	2.93±0.24 <sup>aA</sup>	2.88±0.14 <sup>aA</sup>	3.20±0.21 <sup>aA</sup>	3.37±0.45 <sup>aA</sup>	3.05±0.20 <sup>aA</sup>	2.58±0.20 <sup>aA</sup>
Group 4 (T <sub>3</sub> )	2.81±0.26 <sup>aA</sup>	3.35±0.17 <sup>aA</sup>	3.71±0.51 <sup>aA</sup>	3.38±0.46 <sup>aA</sup>	3.03±0.13 <sup>aA</sup>	2.58±0.42 <sup>aA</sup>
Group 5 (T <sub>4</sub> )	2.57±0.24 <sup>aA</sup>	2.76±0.11 <sup>aA</sup>	3.23±0.20 <sup>aA</sup>	3.48±0.17 <sup>aA</sup>	3.20±0.18 <sup>aA</sup>	2.43±0.18 <sup>aA</sup>

**Table 8:** Mean ± SE values of serum albumin (g/dL) in different groups at different intervals (Phase 2)

Days Groups	0 <sup>th</sup> Day	15 <sup>th</sup> Day	22 <sup>nd</sup> Day	30 <sup>th</sup> Day	Estrus Day	P.D. Day
Group 1 (T <sub>0</sub> )	2.41±0.11 <sup>aA</sup>	3.11±0.35 <sup>aA</sup>	2.75±0.11 <sup>aA</sup>	2.84±0.34 <sup>aA</sup>	2.77±0.14 <sup>aA</sup>	2.59±0.12 <sup>aA</sup>
Group 2 (T <sub>1</sub> )	2.47±0.11 <sup>aA</sup>	3.04±0.36 <sup>aA</sup>	3.01±0.16 <sup>aA</sup>	2.53±0.19 <sup>aA</sup>	2.94±0.18 <sup>aA</sup>	2.89±0.13 <sup>aA</sup>
Group 3 (T <sub>2</sub> )	2.43±0.12 <sup>aA</sup>	3.05±0.24 <sup>aA</sup>	2.78±0.30 <sup>aA</sup>	3.29±0.53 <sup>aA</sup>	3.06±0.18 <sup>aA</sup>	2.84±0.11 <sup>aA</sup>
Group 4 (T <sub>3</sub> )	2.42±0.11 <sup>aA</sup>	3.11±0.16 <sup>aA</sup>	3.11±0.39 <sup>aA</sup>	2.48±0.20 <sup>aA</sup>	2.62±0.17 <sup>aA</sup>	2.67±0.14 <sup>aA</sup>
Group 5 (T <sub>4</sub> )	2.43±0.08 <sup>aA</sup>	3.20±0.17 <sup>abA</sup>	3.89±0.57 <sup>bA</sup>	2.73±0.19 <sup>abA</sup>	3.13±0.20 <sup>abA</sup>	2.91±0.11 <sup>abA</sup>

Means with different superscripts differ significantly at P<0.05

<sup>abcdefg</sup> Superscripts indicate the statistical difference in serum albumin values on different time interval within groups.

<sup>ABC</sup> Superscripts indicate the statistical difference in serum albumin values between groups on different time interval.

**5. References**

1. Bhoite S, Khodke M, Dalvi S, Golher D. Protein profile during peripartum period in Berari goat. Chem Sci Rev Lett. 2019;8(29):48-52.
2. Kaneko J, Harvey J, Bruss M. Clinical Biochemistry of Domestic Animals, Edn 6, Elsevier Inc, 2008, 882-888.
3. Daramola J, Adeloye A, Fatoba T, Soladoye A. Haematological and biochemical parameters of West African Dwarf goats. Livest. Res. Rural Dev. 2005;17:7-8.
4. Dupont J, Scaramuzzi RJ, Reverchon M. The effect of nutrition and metabolic status on the development of follicles, oocytes and embryos in ruminants. Animal. 2014;8:1031-1044.

5. Iriadam M. Variation in certain hematological and biochemical parameters during the peri-partum period in Kilis does. *Small Rumin. Res.* 2007;73(1):54-57.
6. Ishwar AK, Pandey JN. Blood metabolite changes in Black Bengal goats following estrus synchronization and super-ovulation. *Small Ruminant Res.* 1994;13(3):251-256.
7. Krajnicakova M, Kovac G, Kostecky M, Valocy I, Maracek I, Sutiakova I, *et al.* Selected clinico-biochemical parameters in the puerperal period of goats. *Bull. Vet. Inst. Pulawy.* 2003;47:177-182.
8. Manat TD, Chaudhary SS, Singh VK, Patel SB, Puri G. Hemato-biochemical profile in Surti goats during post-partum period. *Vet. World.* 2016;9(1):19-24.
9. Mohammed SA, Razzaque MA, Omar AE, Albert S, Al-Gallaf W. Biochemical and hematological profile of different breeds of goat maintained under intensive production system. *African J. of Biotech.* 2016;15[24]:1253-1257.
10. Samad AR, Ford EJH. The effects of progesterone on glucose and lactate metabolism in ovariectomized sheep. *Quarterly Journal of Experimental Physiology* Banner. 1981;66:73–80.
11. Soares GSL, Souto RJC, Cajueiro JFP, Afonso JAB, Rego RO, Macêdo ATM, *et al.* Adaptive changes in blood biochemical profile of dairy goats during the period of transition. *Revue Médecine Vétérinaire.* 2018;169(1-3):65-75.
12. Sosa C, Abecia JA, Carriquiry M, Forcada F, Martin GB, Palacin I, *et al.* Early pregnancy alters the metabolic responses to restricted nutrition in sheep. *Domestic Animal Endocrinology.* 2009;36:13–23.
13. Stevenson JS, Britt JH. A 100-Year Review: practical female reproductive management. *J. Dairy Sci.* 2017;100(12):10292–10313.
14. Waziri MA, Ribadu AY, Sivachelvan N. Changes in the serum proteins, hematological and some serum biochemical profiles in the gestation period in the Sahel goats. *Vet. Arhiv.* 2010;80:215-224.
15. Wheaton JE, Carlson KM, Windels HF, Johnston LJ. CIDR: A new progesterone releasing intravaginal device for induction of estrus and cycle control in sheep and goats. *Anim. Reprod. Sci.* 1993;33:127–141.
16. Witte TS, Schafersomi S. Involvement of cholesterol, calcium and progesterone in the induction of capacitation and acrosome reaction of mammalian spermatozoa. *Anim. Reprod. Sci.* 2007;102(3-4):181-193.
17. Zamuner F, DiGiacomo K, Cameron Awn, Leury BJ. Endocrine and metabolic status of commercial dairy goats during the transition period. *Journal of Dairy Science.* 2020;103(6):5616-5628.