



ISSN (E): 2277-7695

ISSN (P): 2349-8242

NAAS Rating: 5.23

TPI 2023; 12(11): 565-571

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www.thepharmajournal.com

Received: 02-10-2023

Accepted: 10-11-2023

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Influence of cane regulation and bio-regulators on berry quality of grape cv. Crimson seedless

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Abstract

A filed experiment on Studies on the influence of cane regulation and bio-regulators on growth, yield and quality parameters of grape cv. Crimson Seedless was conducted at Fruit Orchard, Division of Fruit Science, MHREC, University of Horticultural Sciences, Bagalkote, during 2021-22 and 2022-23. The combination of cane regulation at 25-33 canes per vine with 300-400 ppm of ethrel treatment at veraison and 30 days after veraison recorded maximum berry length (20.24 and 19.70 mm), berry diameter (17.48 and 16.86 mm), berry weight (3.07 and 2.61 g), berry firmness (21.88 and 21.22 lb/inch²) Juice percentage (36.63 and 35.77%), TSS (22.64 and 22.61), acidity (0.40 and 0.41%), TSS: acid ratio (55.69 and 55.16).

Keywords: Influence, bio-regulators, veraison, *Vitis vinifera* L.

Introduction

Crimson Seedless grape (*Vitis vinifera* L.) is a late season, attractive, red seedless grape with naturally small berries. Cultivar is the result of five generations of hybridization at the USDA Fresno location, starting in 1926. The source of seedlessness is from Thompson Seedless which was used as parent in the first generation of crossing. C33-199, a late-ripening white seedless grape (With all white grapes in its parentage) was used in final hybridization with emperor to produce 'Crimson seedless'. But inadequate colouration and a small size might detract from their quality, because these small berries do not meet the size requirements of the export market. 'Crimson Seedless' vines are vigorous and moderately productive when cane-pruned. Seven to eight canes may be necessary on mature vines to provide commercial production. Spur-pruned vines are unproductive (Ramming and Tarailo, 1995) [23].

Cane regulation is an essential form of canopy management practice in vineyard operation. This is mainly done to regulate the current season growth, yield and quality of grapes. Crop load adjustment should be considered as one of the technical cultural practices suitable to modify grapevine physiology and plant production towards a defined goal (Matti and Ferrini, 2005) [14]. Higher number of shoots per vine *i.e.*, increased shoot density impairs the productivity of shoots. Therefore, april pruning is done to develop shoots at this rate and their vigour may be curtailed by either pinching or thinning of shoots. While in october pruning, depending upon variety more number of canes are retained on vigorous vines, less is retained on less vigorous ones. Hence, cane thinning is considered as a technique which could lead to improvement in grape (Bravdo *et al.*, 1985) [4].

Plant bio-regulators are generally organic (either synthetic or natural) chemical compounds other than nutrients, performs crucial role in growth and development of any crop. Use of bio-regulators especially abscisic acid or ethylene have been shown to have effects on inducing the expression of the encoding genes in the anthocyanin biosynthesis pathway. These hormones have been used widely to increase the colour and quality of red grapes at commercial scale.

Materials and Methods

The present investigation "Studies on the influence of cane regulation & bi-regulators on growth, yield and quality parameters of Grape cv. 'Crimson Seedless' was carried out during 2021-22 and 2022-23 at grape orchard, MHREC, Sector no-70, UHS, Bagalkote. The experimental design adopted for the present investigation was split plot design with four main treatments, seven sub treatments and replicated thrice. In each vine five canes were selected randomly and tagged for detailed observation.

Results and Discussion

Berry length (mm), berry diameter (mm) and berry weight (g)

Significant differences were recorded among different levels of canopy management practices and bio-regulator treatments with respect to berry length, berry diameter and berry weight are presented in Table (1). Canopy regulation treatments significantly influenced the berry length, berry diameter and berry weight. The maximum berry length (18.80 mm), berry diameter (17.17 mm) and berry weight (3.19 g) was recorded in C₄. The present investigation revealed that 25 canes per vine is superior as it has recorded highest berry length, berry diameter and berry weight as compared to control. The data revealed that increase in berry parameters was found to be associated with reduction in cane per vine and also contributed to increase in bunch weight and yield per vine. This might be due to more availability of assimilates as there is less competition from source to sink. As the severity of pruning reduced, the berry weight decreased, which means they are inversely proportionate to each other (Palanichamy *et al.* (2004) [17]. Similar findings were reported in various studies in grape Porika (2013) [20] in grape cv. Red Globe, Khalil (2020) [9] in Flame Seedless grapes

The bio-regulator treatments were also significantly influenced the berry length, berry diameter and berry weight. The maximum berry length (18.19 mm), berry diameter (18.19 mm) and berry weight (2.87 g) was recorded in G₅. Application of bio-regulators results in increased berry length, berry diameter and berry weight might be due to positive action of bio-regulator helps in stimulating cell elongation process enhances the water absorption by roots and stimulating the biosynthesis of protein will lead to increased berry traits which supported by findings of Amiri *et al.* (2010) [1] in Beidaneh Ghermez grape and Roberto *et al.* (2012) [24].

The interaction effect between canopy regulation and bio-regulators also significantly influenced the berry attributing characters. The maximum berry length (20.24 mm), diameter (17.48 mm) and berry weight (3.39 g) was recorded in C₄G₅. Combined effect of cane regulation and bio-regulator treatments results in increased berry length, berry diameter and berry weight might be due to better mobilization of carbohydrates for better growth and development through cell expansion and cell division.

Berry firmness (lb/inch²)

The berry firmness was significantly influenced by canopy regulation and bio-regulator treatments were recorded in Table (2). The vine regulated with 25 canes per vine has recorded the better berry firmness (21.00 lb/inch²). This increase in berry firmness is attributed to increase in pulp and peel thickness of berries. The variation of berry firmness could be due to genetic nature of the cultivar. Similar reports made by Fawzi *et al.* (1984) [5] and Prculovski *et al.* (2021) [22] in cardinal variety of grape.

The bio-regulators showed significant differences with respect to berry firmness. The maximum berry firmness was recorded in G₅ (21.07 lb/inch²). Increase in berry firmness is strongly associated with turgor pressure of mesocarp cells. The similar results were noticed by Lurie *et al.* (2010) [12] in Crimson Seedless, Amiri *et al.* (2010) [1] in Beidaneh Ghermez grape, Marzouk and Kassem (2011) [15].

The interaction effect between canopy regulation and bio-regulators significantly influenced the berry firmness. In this

study the maximum berry firmness (21.88 lb/inch²) was recorded in C₄G₅. The combination treatment with cane regulation and bio-regulators increased the berry firmness.

Moisture content in berry (%)

Moisture content in berry as influenced by cane regulation and bio-regulators are presented in Table (2). The pooled data showed significant results with respect to moisture content in berry. Among the canopy regulation treatments, the maximum moisture content in berry was recorded in C₄ (16.90) which was followed by C₃ (16.86). The minimum moisture content in berry was recorded in C₁ (15.41). Increase in moisture content in berry is due to the effect of cane regulation helps in more accumulation of photosynthates results in bigger size berry with more moisture content.

The pooled data showed significant results with respect to moisture content in berry in bio-regulator treatments. The maximum moisture content in berry was recorded in G₁ (17.40) which was followed by G₅ (16.72), G₂ (16.58). The minimum moisture content in berry was recorded in G₄ (15.49). Decreased moisture content in berry mainly attributed to bio-regulators effect as a result of increasing membrane permeability which permits moisture stored in cell vacuoles to respire at faster rate.

The interaction effect between canopy regulation and bio-regulator treatments was significantly influenced the moisture content in berry. The new generation bio-regulators quickly reduces moisture content in berry by respiring at faster rate, hence the maximum moisture content was recorded in cane regulation with control treatment *viz.*, C₄G₁ (17.60) which was followed by C₂G₁ (17.59), C₃G₁ (17.50). The minimum moisture content in berry was recorded in C₁G₄ (14.05).

Juice content (%)

The data pertaining to juice content of berry as influenced by canopy management practice and bio-regulators is given in Table 2. The significant differences were recorded with respect to juice content of the berry, among the cane regulation treatments. The highest juice content (36.46%) was recorded in C₄. It is evident from the results that 25 canes per vine was significantly influenced the juice content of the berry. The reason for high juice content might be due to more pulp, more juiciness, less thickness of the skin and bigger size of the berry leads to more juice recovery. The similar results were reported by Bravdo *et al.* (1984) [3] in Carignane grapevines, Porika *et al.* (2015) [21] in Red Globe.

Bio-regulator treatment also significantly influenced the juice content of the berry. The maximum juice content in berry was recorded in G₁ (35.01)

Significant differences were recorded in the interaction effect between canopy regulation and bio-regulators. The maximum juice content (36.63) was recorded in C₄G₅. The higher juice content per gram of fruit in the control may be due to over load of the crop and thin skin of the berry. Similar reports were made by Bravdo *et al.* (1984) [3] in Carignane grapevines, Patil *et al.* (2012) [18] in Cabernet Sauvignon and Shiraz vines and Kaur *et al.* (2013) [30] in Flame seedless grape.

Total soluble solids (°Brix)

The data obtained in respect of total soluble solids (TSS) was influenced by various cane regulation and bio-regulator treatments are reported in Table (3). The data showed

significant results with respect to TSS of the berry maximum total soluble solids (21.94°Brix) was recorded in C₄ compared to control (21.34°Brix) as higher shoot number is positively correlated to the number of bunches but negative with girth of the cane which impacted on impairment of accumulation of sugars due to insufficient assimilates for the higher crop load. These results are strongly supported by the findings of Main and Morris (2000) [13] who mentioned that fruits exposed to sunlight are generally rich in total soluble solids and reduced titratable acidity, compared to non-exposed or canopy shaded. Kliewer and Antcliff (1970) [10] opined that increased TSS in shoot thinned vines might be either due to remobilization of stored carbohydrates or an increase in photosynthetic activity of leaves of remaining shoot and improvement in the light microclimate by reduced shoot number thus, an increase in sink strength. The results of present findings are in agreement with results of Kohale *et al.* (2013) [11] in cv. Sharad Seedless, Ashwini *et al.* (2016) [2] in wine grapes.

Bio-regulator treatments use of ABA and Ethrel significantly influenced the TSS content of the berry. The maximum total soluble solids of berry were recorded in G₇ (22.48°Brix). Application of ethrel and abscisic acid results in higher sugar accumulation leads to expression of genes encoding sucrose transporters resulting in higher sucrose translocation from biosynthesis sites to the berry vacuoles in which sucrose is enzymatically converted into glucose and fructose. The observations of increased TSS content by ethrel and abscisic acid treatments are in agreement with those reported earlier by Nanthakumar *et al.* 2021 [16] in Red Globe grape.

The interaction effect between canopy regulations and bio-regulator treatments found significant with respect to TSS of berry. Application of new generation bio-regulators combined with cane regulation improve the total soluble solid content in Crimson Seedless. The maximum total soluble solids (22.48°Brix) were recorded in C₄G₇. The combination treatment with cane regulation and bio-regulators increased the TSS. It is mainly attributed to mobilization of metabolites from source to sink. The increase in sugars might be ascribed to the conversion of starch and acids into sugars in addition to continuous mobilization of carbohydrates from leaves. Similar results were obtained by Amiri *et al.* (2010) [1] in Beidaneh Ghermez grape, Red Globe grape and Kaur *et al.* (2013) [30] in Flame seedless grape.

Titrateable acidity (%)

The percent of titrateable acidity differed significantly among the canopy management practice and bio-regulator treatments presented in Table (3). The percent titrateable acidity significantly increased with increase in number of shoots per vine. The cane regulation treatments significantly influenced the titrateable acidity of the berry. the lowest titrateable acidity (0.43%) of the berry was recorded in C₄, which was on par with C₂ (0.45%) and C₃ (0.45%). This clearly indicates that crop load has a negative effect on quality of bunches. The deterioration in quality might be due to increase in yield and consequent dilution of sugars in berries. The reason for low titrateable acidity in optimum thinned vines might be due to lesser competition for metabolites among the limited number

of bunches per vine, availability of more photosynthates consequent to better vigour and physiological activities induced in them. The predominant acids of grape *viz.*, malic and tartaric acids are synthesized in leaves. These acids are translocated from leaves to bunch. This higher quantum of acids might have deposited in bunch during development and this could have caused higher acidity in less intensive pruning treatments. Shikhamani *et al.* (2008) [26] reported that the higher number of canes per vine resulted in to denser canopy and decreased the interception of light hence hindered the reduction of acid at the time of berry maturity. These results are in line with Somkuwar and Ramteke, 2007 [2], Jogaiah *et al.* (2012) [8], Harikanth, 2013 [7] and Ashwini *et al.*, 2016 [2].

The pooled data with respect to titrateable acidity of the berry in bio-regulator treatments was found to be significant. The lowest acidity (0.41%) of the berry was recorded in G₇ which was at par with G₄ (0.42%). It clearly indicates the role of application of ethrel and abscisic acid results in reduction in acidity could be due to effect of these bio-regulators in increasing membrane permeability which permits acids stored in cell vacuoles to respire at faster rate. Results obtained are agreement with Peppi *et al.* (2006) [19].

The interaction effect between canopy regulation and bio-regulator treatments significantly influenced the titrateable acidity of the berry. The lowest titrateable acidity (0.40%) was recorded in C₄G₇ which was on par with C₄G₄ (0.41%). In the present study, cane regulation alone does not influence per cent titrateable acidity of berries. The combination treatment with cane regulation and bio-regulators reduce the acidity. Similar results were obtained by Kaur *et al.* (2013) [30] in Flame Seedless grape.

TSS to acid ratio

In the present study, TSS to acid ratio was significantly influenced by cane regulation and bio-regulators are presented in Table (3). Among the cane regulation treatments, the highest TSS: acid ratio was recorded in C₄ (51.09%), which was at par with C₃ (49.24%). The availability of more photosynthates consequent to better vigour and physiological activities. Similar findings were given by Senthil Kumar (2014) [25] in grapes cv. Italia, Porika *et al.* (2015) [21] in grape cv. Red Globe and Veena *et al.* (2015) [29].

Application of new generation bio-regulators *viz.*, abscisic acid and ethrel of showed significant differences with respect to TSS to acid ratio. The highest TSS: acid ratio (54.21%) was recorded in G₇ which was at par with G₅ (46.53%). It was obvious that the acidity reduced with an increase in the total soluble solids resulted in increased TSS: Acidity ratio. These findings are in conformity with results those reported earlier by and Singh *et al.* (1994) [27].

The interaction effect between canopy regulations and growth regulator treatments was found significant with respect to TSS: acid ratio of berry. The highest TSS: acid ratio (55.69%) was recorded in C₄G₇. The combination treatment with cane regulation and bio-regulators increased the TSS to Acid ratio. Similar results were obtained by Kaur *et al.* (2013) [30] in Flame Seedless grape'

Table 1: Berry length (mm), berry diameter (mm) and berry weight (g) after October pruning in grapes *cv.* Crimson Seedless as influenced by cane regulation and bio-regulators

Treatment	Berry length (mm)			Berry diameter (mm)			Berry weight (g)		
	2021	2022	Pooled	2021	2022	Pooled	2021	2022	Pooled
Main plot (Cane regulation)									
C ₁ (Control)	16.35	16.18	16.26	15.57	15.54	15.55	2.17	2.50	2.33
C ₂ (50 canes vine ⁻¹)	16.55	16.53	16.54	16.36	16.34	16.35	2.35	2.54	2.44
C ₃ (33 canes vine ⁻¹)	18.77	18.67	18.72	16.68	16.56	16.62	2.86	2.86	2.86
C ₄ (25 canes vine ⁻¹)	18.80	18.79	18.80	17.19	17.14	17.17	3.25	3.13	3.19
S.Em ±	0.17	0.08	0.08	0.16	0.20	0.10	0.13	0.07	0.07
CD at 5%	0.60	0.29	0.28	0.56	0.68	0.36	0.46	0.23	0.25
Sub plot (Bio- regulators)									
G ₁ (Control)	17.84	17.48	17.66	16.05	16.02	16.04	2.30	2.55	2.43
G ₂ (ABA 200 ppm)	18.01	17.94	17.98	16.73	16.67	16.70	2.73	2.74	2.73
G ₃ (ABA 300 ppm)	17.78	17.64	17.71	16.50	16.47	16.49	2.61	2.83	2.72
G ₄ (ABA 400 ppm)	17.40	17.22	17.31	16.09	16.05	16.07	2.63	2.65	2.64
G ₅ (Ethrel 300 ppm)	18.21	18.17	18.19	16.83	16.75	16.79	2.82	2.92	2.87
G ₆ (Ethrel 350 ppm)	17.33	17.26	17.29	16.61	16.54	16.58	2.84	2.84	2.84
G ₇ (Ethrel 400 ppm)	16.78	17.08	16.93	16.33	16.25	16.29	2.67	2.76	2.71
S.Em ±	0.27	0.26	0.16	0.19	0.18	0.14	0.11	0.07	0.06
CD at 5%	0.77	0.73	0.46	0.54	0.51	0.39	0.30	0.20	0.18
Interactions (Main plot × Sub plot)									
C ₁ G ₁	16.53	16.20	16.37	15.58	15.55	15.56	2.04	2.45	2.25
C ₁ G ₂	16.52	16.23	16.38	15.92	15.86	15.89	2.22	2.50	2.36
C ₁ G ₃	16.28	16.18	16.23	15.41	15.39	15.40	2.06	2.55	2.31
C ₁ G ₄	15.68	15.60	15.64	14.90	14.87	14.88	1.99	2.37	2.18
C ₁ G ₅	18.69	18.32	18.50	16.15	16.13	16.14	2.51	2.60	2.55
C ₁ G ₆	15.60	15.57	15.58	15.68	15.65	15.67	2.28	2.53	2.41
C ₁ G ₇	15.15	15.13	15.14	15.33	15.33	15.33	2.08	2.48	2.28
C ₂ G ₁	16.41	16.14	16.28	16.02	16.00	16.01	2.11	2.52	2.32
C ₂ G ₂	17.56	17.28	17.42	16.63	16.61	16.62	2.50	2.62	2.56
C ₂ G ₃	17.70	17.53	17.62	16.51	16.47	16.49	2.19	2.52	2.35
C ₂ G ₄	17.65	17.51	17.58	16.03	15.98	16.01	2.28	2.38	2.33
C ₂ G ₅	19.15	18.74	18.94	16.70	16.70	16.70	2.54	2.67	2.61
C ₂ G ₆	13.65	14.65	14.15	16.60	16.59	16.59	2.49	2.58	2.54
C ₂ G ₇	13.77	13.83	13.80	16.05	16.02	16.04	2.31	2.48	2.39
C ₃ G ₁	18.41	17.78	18.10	16.23	16.22	16.23	2.38	2.62	2.50
C ₃ G ₂	18.33	18.70	18.52	16.92	16.84	16.88	2.97	2.78	2.88
C ₃ G ₃	18.20	17.93	18.07	16.79	16.79	16.79	2.92	2.72	2.82
C ₃ G ₄	17.68	17.68	17.68	16.42	16.38	16.40	2.72	2.87	2.80
C ₃ G ₅	19.76	19.63	19.70	16.98	16.73	16.86	3.05	3.09	3.07
C ₃ G ₆	19.48	19.50	19.49	16.79	16.59	16.69	3.03	3.04	3.04
C ₃ G ₇	18.76	18.77	18.76	16.62	16.35	16.48	2.93	2.92	2.93
C ₄ G ₁	17.55	17.48	17.52	16.38	16.33	16.36	2.67	2.63	2.65
C ₄ G ₂	20.03	19.76	19.89	17.47	17.35	17.41	3.52	3.17	3.35
C ₄ G ₃	19.67	19.63	19.65	17.28	17.25	17.27	3.32	3.15	3.24
C ₄ G ₄	19.22	19.13	19.18	17.02	16.98	17.00	3.16	3.28	3.22
C ₄ G ₅	20.28	20.20	20.24	17.51	17.45	17.48	3.56	3.22	3.39
C ₄ G ₆	19.10	19.35	19.22	17.38	17.34	17.36	3.19	3.32	3.26
C ₄ G ₇	16.57	16.69	16.63	17.29	17.30	17.30	3.35	3.17	3.26
S.Em ±	0.54	0.52	0.33	0.38	0.36	0.28	0.21	0.14	0.12
CD at 5%	1.55	1.47	0.93	1.08	1.02	0.79	0.60	0.41	0.36

Table 2: Berry firmness, moisture percent and juice percentage in grapes *cv.* Crimson Seedless as influenced by cane regulation and bio-regulators

Treatment	Berry firmness (Newton)			Moisture (%)			Juice percentage (v/w)		
	2021	2022	Pooled	2021	2022	Pooled	2021	2022	Pooled
Main plot (Cane regulation)									
C ₁ (Control)	19.69	19.83	19.76	15.39	15.44	15.41	33.19	33.39	33.29
C ₂ (50 canes vine ⁻¹)	20.00	20.05	20.02	16.24	16.28	16.26	33.33	33.86	33.59
C ₃ (33 canes vine ⁻¹)	20.31	20.52	20.41	16.83	16.89	16.86	34.55	35.53	35.04
C ₄ (25 canes vine ⁻¹)	20.95	21.05	21.00	16.89	16.91	16.90	36.44	36.47	36.46
S.Em ±	0.17	0.16	0.13	0.21	0.17	0.16	0.15	0.20	0.10
CD at 5%	0.58	0.55	0.46	0.73	0.60	0.56	0.51	0.70	0.36
Sub plot (Bio- regulators)									
G ₁ (Control)	18.57	18.65	18.61	17.39	17.42	17.40	34.98	35.04	35.01

G ₂ (ABA 200 ppm)	20.85	20.86	20.86	16.56	16.60	16.58	34.61	35.06	34.84
G ₃ (ABA 300 ppm)	20.60	20.66	20.63	16.05	16.07	16.06	34.49	34.69	34.59
G ₄ (ABA 400 ppm)	20.04	20.22	20.13	15.47	15.51	15.49	34.22	34.28	34.25
G ₅ (Ethrel 300 ppm)	21.02	21.12	21.07	16.68	16.76	16.72	34.34	35.27	34.80
G ₆ (Ethrel 350 ppm)	20.68	20.69	20.69	16.40	16.42	16.41	34.21	34.51	34.36
G ₇ (Ethrel 400 ppm)	19.89	20.36	20.13	15.82	15.88	15.85	33.78	34.86	34.32
S.Em ±	0.21	0.27	0.18	0.21	0.17	0.15	0.25	0.21	0.17
CD at 5%	0.59	0.77	0.51	0.58	0.48	0.43	0.71	0.60	0.47
Interactions (Main plot × Sub plot)									
C ₁ G ₁	18.08	18.26	18.17	16.87	16.98	16.93	33.51	33.54	33.53
C ₁ G ₂	20.21	20.23	20.22	15.43	15.49	15.46	33.09	33.50	33.30
C ₁ G ₃	20.08	20.07	20.08	14.52	14.55	14.54	33.25	33.15	33.20
C ₁ G ₄	19.64	19.80	19.72	14.01	14.09	14.05	33.04	33.27	33.15
C ₁ G ₅	20.21	20.55	20.38	16.01	16.07	16.04	33.35	33.34	33.34
C ₁ G ₆	20.05	20.09	20.07	15.85	15.86	15.86	32.95	33.70	33.33
C ₁ G ₇	19.61	19.85	19.73	15.01	15.02	15.02	33.13	33.20	33.17
C ₂ G ₁	18.34	18.44	18.39	17.55	17.59	17.57	34.25	34.24	34.25
C ₂ G ₂	20.55	20.55	20.55	16.55	16.57	16.56	33.89	34.30	34.10
C ₂ G ₃	20.15	20.21	20.18	16.01	16.04	16.03	33.57	33.78	33.67
C ₂ G ₄	19.90	19.92	19.91	15.02	15.06	15.04	32.88	32.91	32.90
C ₂ G ₅	20.84	20.75	20.80	16.56	16.59	16.57	33.13	33.15	33.14
C ₂ G ₆	20.46	20.38	20.42	16.02	16.06	16.04	32.07	34.14	33.11
C ₂ G ₇	19.74	20.12	19.93	16.01	16.03	16.02	33.50	34.51	34.01
C ₃ G ₁	18.63	18.65	18.64	17.53	17.50	17.52	35.70	35.83	35.77
C ₃ G ₂	21.10	21.10	21.10	17.27	17.31	17.29	34.96	35.89	35.42
C ₃ G ₃	20.77	20.90	20.84	16.84	16.88	16.86	35.07	35.40	35.23
C ₃ G ₄	19.88	20.36	20.12	16.34	16.38	16.36	34.25	34.87	34.56
C ₃ G ₅	21.17	21.27	21.22	17.30	17.33	17.31	34.50	36.00	35.25
C ₃ G ₆	20.79	20.94	20.86	16.87	16.90	16.89	34.14	35.24	34.69
C ₃ G ₇	19.81	20.43	20.12	15.67	15.91	15.79	33.25	35.50	34.37
C ₄ G ₁	19.25	19.25	19.25	17.60	17.60	17.60	36.47	36.53	36.50
C ₄ G ₂	21.55	21.55	21.55	17.00	17.03	17.02	36.52	36.54	36.53
C ₄ G ₃	21.39	21.48	21.44	16.81	16.80	16.81	36.30	36.30	36.30
C ₄ G ₄	20.75	20.78	20.77	16.51	16.50	16.51	36.51	36.20	36.35
C ₄ G ₅	21.87	21.90	21.88	16.85	17.05	16.95	36.41	36.85	36.63
C ₄ G ₆	21.44	21.37	21.40	16.88	16.85	16.86	36.43	36.45	36.44
C ₄ G ₇	20.42	21.03	20.72	16.57	16.55	16.56	36.45	36.44	36.45
S.Em ±	0.42	0.54	0.36	0.41	0.34	0.30	0.50	0.42	0.33
CD at 5%	1.18	1.53	1.02	1.17	0.97	0.86	1.41	1.19	0.94

Table 3: TSS, titratable acidity (%) and TSS: acid (%) in grapes *cv.* Crimson Seedless as influenced by cane regulation and bio-regulators

Treatment	TSS (° Brix)			Titratable acidity (%)			TSS: acid (%)		
	2021	2022	Pooled	2021	2022	Pooled	2021	2022	Pooled
Main plot (Cane regulation)									
C ₁ (Control)	21.30	21.37	21.34	0.482	0.479	0.481	44.48	44.99	44.74
C ₂ (50 canes vine ⁻¹)	21.83	21.83	21.83	0.450	0.447	0.448	49.03	48.85	48.94
C ₃ (33 canes vine ⁻¹)	21.87	21.87	21.87	0.450	0.452	0.451	49.12	49.36	49.24
C ₄ (25 canes vine ⁻¹)	21.89	21.99	21.94	0.431	0.430	0.430	51.09	51.09	51.09
S.Em ±	0.06	0.06	0.05	0.007	0.009	0.004	0.33	0.20	0.16
CD at 5%	0.22	0.20	0.17	0.023	0.030	0.015	1.13	0.68	0.55
Sub plot (Bio- regulators)									
G ₁ (Control)	20.64	20.65	20.64	0.527	0.527	0.527	39.24	39.21	39.23
G ₂ (ABA 200 ppm)	21.37	21.44	21.40	0.476	0.469	0.473	45.04	45.68	45.36
G ₃ (ABA 300 ppm)	21.86	21.87	21.86	0.446	0.445	0.445	49.21	49.29	49.25
G ₄ (ABA 400 ppm)	22.42	22.43	22.42	0.418	0.418	0.418	53.73	53.74	53.74
G ₅ (Ethrel 300 ppm)	21.40	21.59	21.50	0.463	0.463	0.463	46.31	46.75	46.53
G ₆ (Ethrel 350 ppm)	21.91	21.92	21.91	0.428	0.430	0.429	51.36	51.03	51.20
G ₇ (Ethrel 400 ppm)	22.48	22.47	22.48	0.416	0.412	0.414	54.12	54.29	54.21
S.Em ±	0.13	0.15	0.11	0.012	0.010	0.008	2.68	2.59	2.03
CD at 5%	0.37	0.44	0.31	0.033	0.030	0.022	7.62	7.35	5.78
Interactions (Main plot × Sub plot)									
C ₁ G ₁	20.49	20.43	20.46	0.550	0.550	0.550	37.26	37.18	37.22
C ₁ G ₂	20.90	20.93	20.92	0.503	0.500	0.502	41.53	41.91	41.72
C ₁ G ₃	21.29	21.31	21.30	0.483	0.477	0.480	44.05	44.73	44.39
C ₁ G ₄	22.01	22.04	22.03	0.453	0.447	0.450	48.56	49.39	48.98
C ₁ G ₅	20.92	21.33	21.13	0.487	0.490	0.488	42.99	43.56	43.28

C ₁ G ₆	21.40	21.40	21.40	0.470	0.467	0.468	45.55	45.87	45.71
C ₁ G ₇	22.11	22.13	22.12	0.430	0.423	0.427	51.44	52.32	51.88
C ₂ G ₁	20.61	20.63	20.62	0.520	0.520	0.520	38.67	38.47	38.57
C ₂ G ₂	21.51	21.52	21.51	0.477	0.467	0.472	44.50	45.47	44.98
C ₂ G ₃	22.02	22.02	22.02	0.450	0.440	0.445	51.62	50.11	50.86
C ₂ G ₄	22.51	22.52	22.52	0.407	0.407	0.407	55.81	55.00	55.40
C ₂ G ₅	21.55	21.57	21.56	0.467	0.460	0.463	45.87	45.91	45.89
C ₂ G ₆	22.06	22.06	22.06	0.417	0.427	0.422	52.95	52.54	52.75
C ₂ G ₇	22.58	22.49	22.53	0.410	0.410	0.410	53.78	54.42	54.10
C ₃ G ₁	20.69	20.69	20.69	0.533	0.537	0.535	39.80	39.80	39.80
C ₃ G ₂	21.52	21.53	21.52	0.483	0.473	0.478	45.16	46.19	45.68
C ₃ G ₃	22.05	22.07	22.06	0.427	0.440	0.433	49.02	50.17	49.59
C ₃ G ₄	22.56	22.56	22.56	0.403	0.410	0.407	55.48	55.47	55.48
C ₃ G ₅	21.54	21.56	21.55	0.470	0.470	0.470	46.18	46.91	46.54
C ₃ G ₆	22.09	22.09	22.09	0.417	0.420	0.418	53.05	51.78	52.42
C ₃ G ₇	22.60	22.61	22.61	0.420	0.413	0.417	55.15	55.17	55.16
C ₄ G ₁	20.75	20.83	20.79	0.503	0.500	0.502	41.23	41.39	41.31
C ₄ G ₂	21.54	21.78	21.66	0.440	0.437	0.438	48.97	49.17	49.07
C ₄ G ₃	22.07	22.07	22.07	0.423	0.423	0.423	52.13	52.15	52.14
C ₄ G ₄	22.58	22.59	22.59	0.410	0.410	0.410	55.08	55.11	55.09
C ₄ G ₅	21.59	21.91	21.75	0.430	0.430	0.430	50.22	50.62	50.42
C ₄ G ₆	22.09	22.11	22.10	0.410	0.407	0.408	53.88	53.94	53.91
C ₄ G ₇	22.63	22.65	22.64	0.403	0.400	0.402	56.12	55.26	55.69
S.Em ±	0.26	0.31	0.22	0.023	0.021	0.015	0.77	0.75	0.59
CD at 5%	0.74	0.88	0.63	0.066	0.059	0.043	2.20	2.12	1.67

Conclusion

From this study, it can be clearly concluded that, the cane regulation is essential form of thinning in vineyard operation and considered as a technique which could lead to tremendous improvement in quality parameters of Crimson Seedless as it is very vigorous cultivar. Application bio-regulator is one of important operations to get export quality standards and has more demand in market. In case of bio- regulator treatments, application ethrel 300-400 ppm at veraison stage has positively influenced berry quality like, berry, length, berry diameter, berry weight TSS, acidity, juice percentage, TSS: Acid ratio and Total anthocyanins in skin and berry. Among the interaction effects, the vines regulated with 25-33 canes per vine with the application of ethrel 300-400 ppm at veraison stage followed by vines regulated with 25-33 canes per vine with the application of abscisic acid 200-400 ppm at veraison stage was found superior over the control and other interactions.

References

- Amiri ME, Fallahi E, Parseh. Application of Ethephon and ABA at 40% veraison advanced maturity and quality of 'Beidaneh Ghermez' grape. *Acta Hort.* 2010;884:371-378.
- Ashwini SG, Kulapati H, Sandhya GC, Shashidhar MS. Impact of canopy management on fruit quality of wine grapes under northern dry zone of Karnataka. *Int J Curr. Sci.* 2016;6(3):2762-2764.
- Bravdo B, Hepner Y, Loinger C, Kohen S, Tabacman H. Effect of crop level on growth, yield and wine quality of a high yielding Carignane vineyard. *American J Enol. Vitic.* 1984;35(4):247-252.
- Bravdo B, Hapner Y, Loinger RC, Kohen S, Tabacman H. Effect of crop level and crop load on growth, yield, must and wine composition and quality of Cabernet Sauvignon. *American J Enol. Vitic.* 1985;36(2):125-131.
- Fawzi FA, Bondok Z, Ghobrial GF. Effect of cane length on cropping and some mechanical and chemical properties of bunches in Thompson Seedless grape variety. *Annals Agric. Sci. Ain Shams. Univ. Cairo.* 1984;29(1):25-60.
- Han W, Han N, He X, Zhao X. Berry thinning to reduce bunch compactness improves fruit quality of Cabernet Sauvignon (*Vitis vinifera* L.). *Sci. Hortic.* 2019;246:589-596.
- Harikanth H. Studies on season and intensity of pruning on growth, yield and quality of grape (*Vitis vinifera* L.) cv. Red Globe, M.Sc., Thesis Tamil Nadu Agri. Univ., Coimbatore; c2013.
- Jogaiah S, Striegler K, Bergmeier R, Harris J. Influence of cluster exposure to sun on fruit composition of 'Norton' grapes (*Vitis aestivalis* M.) in Missouri. *Int. J Fruit. Sci.* 2012;12:410-426.
- Khalil AH. Improved yield, fruit quality, and shelf life in Flame Seedless grapevine with pre-harvest foliar applications of Forchlorfenuron, Gibberellic acid and Abscisic acid, *J Hort. Res.* 2020;28(1):77-86.
- Kliwer WM, Antcliff AJ. Influence of defoliation, leaf darkening and cluster shading on the growth and composition of Sultana grapes. *American J Enol. Vitic.* 1971;21:26-36.
- Kohale VS, Kulkarni SS, Ranpise SA, Garad BV. Effect of pruning on fruiting of Sharad Seedless grapes. *Bioinfolet.* 2013;10(1):300-302.
- Lurie S, Lichter A, Kaplunov T, Zutahy Y, Oren-Shamie M, Ovadia R. Improvement of 'Crimson Seedless grape colour by abscisic acid treatment. *Acta Hort.* 2010;880:183-189.
- Main GL, Morris JR. Leaf-removal effects on Cynthiana yield, juice composition, and wine composition. *American J Enol. Vitic.* 2000;55:147-152.
- Matti GB, Ferrini F. The effects of crop load on Sangiovese grapevine. *Acta Hort.* 2005;689:239-242.
- Marzouk HA, Kassem HA. Improving yield quality and shelf life of Thompson Seedless grape vine by pre-harvest foliar applications. *Scientia Hort.* 2011;130(2):425-430.
- Nanthakumar S, Manju V, Kumar AV. Effect of plant

- growth regulators to improve the colour and sugar content of grapes (*Vitis vinifera* L.). cv. Red Globe. Int. J Environ. Clim. 2021;11(11):216-221.
17. Palanichamy V, Jindal PC, Singh R. Studies on severity of pruning in grapes (*Vitis vinifera* L.) cv. Pusa Navrang-A teinturien Hybrid. Agric. Sci. Digest. 2004;24(2):145-147.
 18. Patil VS, Patil DR, Jamadar MM, Kanamadi VC, Swamy GSK. Influence of cane regulation on the fruit quality, pest and disease incidence in wine grapes (*Vitis vinifera* L.). Karnataka J Agric. Sci. 2012;25(3):367-369.
 19. Peppi MC, Fidelibus MW, Dokoozlian NK. Application timing and concentration of abscisic acid affect the quality of 'redglobe' grapes. J of Hort. Sci. Biotech. 2006;82(2):304-310.
 20. Porika H. Studies on season and intensity of pruning on growth, yield and quality of grape (*Vitis vinifera* L.) cv. Red Globe. M.Sc., Thesis, Tamil Nadu Agricultural University, Coimbatore; c2013.
 21. Porika H, Jagadeesha M, Suchitra M. Effect of pruning severity on quality of grapes cv. Red Globe for summer season. Adv. Crop Sci. Tech. 2015;10(5):29-36.
 22. Prculovski Z, Petkov M, Boskov K, Kryeziu S. Effect of bunch load on the quality of cardinal grape variety. Int. J agric. 2021;67(4):103-113.
 23. Ramming D, Tarailo RE. Crimson Seedless': A new late-maturing, Red Seedless grape. Hort. Sci. 1995;30(7):1473-1474.
 24. Roberto SR, De assis AM, Yamamoto LY, Miotto LV, Sato AJ, Koyama R, *et al.* Application timing and concentration of abscisic acid improve color of 'Benitaka' table grape 'Benitaka'. Sci. Hortic. 2012;142:44-48.
 25. Senthilkumar S. Studies on standardization of pruning and influence of pre-harvest practices on yield and quality of grape (*Vitis vinifera* L.) cv. Italia. *Ph.D., Thesis*, Tamil Nadu Agri. Uni. Coimbatore; c2014.
 26. Shikhamany SD, Somkuwar RG, Venugopalan R. Evaluation of canopy efficiency using leaf area index in Thompson Seedless vines. Acta Hort. 2008;785:389-391.
 27. Singh S, Singh IS, Singh DN. Effect of GA₃ on ripening and quality of grape (*Vitis vinifera* L.). Orissa. J Hort. 1994;22:66-70.
 28. Somkuwar RG, Ramteke SD. Effect of bunch retention, quality and yield in Sharad Seedless. Annual Report 2006-07, National Research Center for Grapes, Pune; c2007. p. 20.
 29. Veena J, Vinod K, Manoj D, Santosh P, Variath MT, Santosh K. Multivariate analysis of colored and white grape grown under semi-arid tropical conditions of Peninsular India. Int. J Agri. Crop Sci. 2015;8(3):350-365.
 30. Kaur M. Blended learning-its challenges and future. Procedia-social and behavioral sciences. 2013;93:612-617.