www.ThePharmaJournal.com

The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2022; SP-11(9): 610-616 © 2022 TPI www.thepharmajournal.com

Received: 27-06-2022 Accepted: 29-07-2022

Suryaprakash Pannu

Ph.D. Scholar, Animal Reproduction, Gynaecology and obstetrics, ICAR-National Dairy Research Institute (Deemed University), Karnal, Haryana, India

Tapendra Kumar

Ph.D. Scholar, Department of Veterinary Gynaecology & Obstetrics, College of Veterinary & Animal Science, RAJUVAS, Bikaner, Rajasthan, India

Pratyush Kumar

Ph.D. Scholar, Animal Reproduction, Gynaecology and obstetrics, ICAR-National Dairy Research Institute (Deemed University), Karnal, Haryana, India

Sandeep Dholpuria

Assistant Professor, Department of Veterinary Gynaecology & Obstetrics, College of Veterinary & Animal Science, Bikaner Rajasthan, India

Manisha Mehra

Assistant Professor, Department of Veterinary Pathology, College of Veterinary & Animal Science, Bikaner Rajasthan, India

Maya Mehara

M.V.Sc. Scholar, Department of Veterinary Public health, College of Veterinary & Animal Sciences, Bikaner Rajasthan, India

Corresponding Author:

Suryaprakash Pannu Ph.D. Scholar, Animal Reproduction, Gynaecology and obstetrics, ICAR-National Dairy Research Institute (Deemed University), Karnal, Haryana, India

Post-partum metabolic diseases in cattle and buffalo: A review

Suryaprakash Pannu, Tapendra Kumar, Pratyush Kumar, Sandeep Dholpuria, Manisha Mehra and Maya Mehara

Abstract

The periparturient period is when cows are most susceptible to infections and metabolic problems, therefore nutritional management throughout the dry season may have an impact. Increased body tissue mobilisation, decreased plasma glucose levels, and increased plasma concentrations of non-esterified fatty acids (NEFA) and ketone bodies are all results of the endocrine changes and regulation that occur during calving. Immune competence and abnormal body mobilisation serve as fundamental rationals for metabolic and immune state and support the creation of markers for metabolic imbalance and the onset of diseases early on. The highest incidence of metabolic illnesses such milk fever, postpartum haemoglobinuria, and ketosis is connected with the transitional period between three weeks prepartum and three weeks postpartum. Metabolic blood profiles are frequently used to detect and signal metabolic disturbances and low productivity. Reduced milk production, an increased chance of delayed estrous, and lower conception rates are all symptoms of metabolic disorders.

Keywords: Metabolic, milk fever, ketosis, cattle, buffalo

Introduction

In the peripartum period, there is a correlation between depression of feed intake and metabolic disorders. The prevalence of metabolic problems in buffaloes associated to production or parturition is limited. (Purohit et al., 2013) [58]. These changes are likely noticeable because buffaloes produce less milk, which leads to a less severe negative energy balance during parturition (Grasso et al., 2004) ^[27] diminished blood stream lipid metabolization in buffalo at their peak lactation (Monteiro et al., 2012)^[53] without any signs of fatty liver in the postpartum buffalo (Jalali et al., 2011)^[38] and better utilization of minerals by buffalo from coarser roughages (Sebastian et al., 1970)^[63]. A series of diseases known as metabolic disorders of cattle affect dairy cows right after parturition. For dairy cows, the transition period is roughly 3 weeks prior to delivery through 3 weeks following delivery. During this time of transition, disease incidence and severity are at their highest phases when the immune system is compromised. Around the time of calving, metabolic and microbial diseases involve a disordered inflammatory response. Dairy cows must be monitored for changes in feed intake during the periparturient period since inadequate intake of dry matter is associated with numerous of postpartum diseases, such as ketosis, metritis and mastitis (Bareille *et al.*, 2003; Huzzey *et al.*, 2007)^[5, 36].

Post parturient haemoglobinuria

In India, postpartum hemoglobinuria (PPH) represents a significant threat to milking buffaloes during late pregnancy and the first few months of lactation (Ghanem and El-Deeb 2010; Mahmood *et al.*, 2013)^[24, 47]. According to Durrani *et al.* (2010)^[20] the prevalence was highest at the fifth lactation and lowest at the first. Acute intravascular hemolysis and severe anaemic anoxia make it highly lethal. Haemoglobinuria is the common problem in buffaloes specially fed on leguminous fodder during summer season (Jain *et al.*, 2012)^[37] possibly due to metabolic deficits during high milk yields (Choudhary *et al.*, 2013)^[12]. This condition has phosphorus deficiency as a constant finding, and treatment with sodium acid phosphate is effective in clinical situations (Pandey and Misra, 1987). Hypophosphataemia is anticipated to lead to a reduction in red blood cells, glycolysis, and ATP generation. Subnormal concentration of ATP predispose red blood cells to altered function and structure, a loss of normal deformability and increase in fragility and hemolysis with resulting hemoglobinaemia and hemoglobinuria (Khan and Akhtar, 2007)^[42].

Due to decreased level of ATP, erythrocytes loose their deformability become rigid and ultimately leave the circulation due to decreased life span (Radostits et al., 2007; Akhtar et al., 2007b)^[59, 1]. According to Mata and Bhardwaj (1985) hypothesis, phosphorus deficit in PPH buffaloes reduces the rate at which erythrocytes use glucose and produce ATP, which lowers their ability to synthesise glutathione and makes them more susceptible to the harmful effects of oxidative stress. Intravascular hemolysis is eventually caused by the oxidative stress that accompanies the lipid peroxidation of the red cell membrane. Although the specific pathophysiology of this condition is not entirely understood, risk factors for it include eating cruciferous plants, berseem saponin not getting enough phosphorus in your diet, having low levels of serum copper and selenium and having high levels of molybdenum. (Neto et al., 2007; Brechbuhl et al., 2008) [55, 10]. While an excess of molybdenum also lowers phosphorous content by interfering with its absorption and increasing its elimination through urine, low phosphorous diets with high calcium content cause hypophosphataemia by decreasing phosphorous absorption from the gastro-intestinal tract due to a wider ratio of calcium and phosphorous (Khan and Akhtar, 2007; Dua, 2009)^[42, 15]. Additionally, hypermolybdenosis causes a decrease in copper, which decreases the activity of the copper-containing enzyme of superoxide dismutase, a component the erythrocytendefence mechanism against oxidative stress (Kahn and Line, 2005; Radostits et al., 2007)^[40, 59].

Clinical findings

Disease with haemoglobinemia as a primary clinical symptom anemia, hemoglobinuria and straining while urinating (Gupta *et al.*, 2010)^[29]. With the progression of anaemia, tachycardia, quick and shallow breathing, depression, and reduced milk production, mucous membranes becoming pale or icteric (Ghanem and El-Deeb, 2010)^[24]. Increased heart rate and pulse rate may be seen (Akhtar, 2006; Gupta *et al.*, 2010)^{[3, ^{29]}. Depending on the severity and length of the sickness, haemoglobinuric buffaloes produced urine that ranged in colour from red to dark red to coffee-colored (Akhtar, 2006) ^[3]. If the animal is left untreated, there is a gradual loss of body function, and recumbency is followed by death.}

Diagnosis

The history of the recently calved animal and clinical symptoms are used to make the diagnosis. A blood analysis demonstrates a sharp decline in haemoglobin the number of total erythrocytes, and packed cell volume. Urine examined under a microscope reveals that there are no erythrocytes and is positive for the benzidine test. Low levels of 0.5-3.0 mg/dl of serum inorganic phosphorous (normal range: 4.0-7.0 mg/dl) are present. Numerous authors reported that during hemolytic crises, serum inorganic phosphorus levels are extremely low (0.13-0.49 mmol/l) (Ellison *et al.*, 1986)^[21].

Line of treatment

Following intravenous administration of 60 g of sodium acid phosphate dissolved in 300 ml of 5 percent dextrose, the same amount was then given orally twice daily for three days (Singh *et al.*, 1989)^[69]. Ascorbic acid at 5g per animal helps iron maintain a reducing state, reduce intravascular hemolysis, and reduce oxidative stress on red blood cells (Benerjee, 1998)^[4]. It was successful to treat PPH in buffaloes by injecting botropase (10 ml/animal), a blood coagulant made from the venom of the snake *Bothrops jararaca*, which appears to have antifibrinolytic activity (Goel *et al.*, 1988) ^[25]. For 5-7 days, oral supplements of monosodium phosphate, dicalcium phosphate, or 100 gm of bone meal should be administered daily. Additionally recommended are parenteral injections of copper (120 mg/cow). Tea leaves can be used as a supportive treatment along with toldimfos sodium (4-dimethyle 1-2 methyl phosphorus acid sodium salt) as an alternative to sodium acid phosphate therapy because of their astringent action, which helps to clear up urine staining (Durrani *et al.*, 2010) ^[20].

Prevention and control

Animals should be provided a balanced diet rich in minerals. After parturition mineral mixture should be given @ 50 g/day orally for 1 month. A decrease in the incidence of the disease is reported after copper supplementation of cattle in a copperdeficient area.

Milk fever

Milk fever is metabolic disease also known as parturient paresis, parturient apoplexy, calving paralysis and post parturient hypocalcaemia (Littledike et al., 1981)^[46]. Most frequently develops within 48 hours after parturition, especially in high milk producing dairy cows. The incidence of milk fever is highest in jersey breed (Littledike et al; 1981) $[^{46]}$. Over-conditioned cows (body condition score >3.5, in a scale of 1-5) are at increased risk of hypocalcaemia (Heuer et al., 1999) ^[33]. Ionized calcium levels in tissue fluids are decreasing which is the primary biochemical defect in this diesase. Clinical signs of milk fever include weakness, recumbency, shock and mortality. Diesase commonly occurs in 5-10 years age group and most marked in third to seventh parity (Singh and Singh, 2017)^[70]. Dietary calcium intake influences both the mobilisation of calcium from bone and the absorption of calcium from the gut, which is regulated by 1, 25(OH)2D3. The efficiency of intestinal absorption and the mobilisation of calcium from bone both decline when dietary Ca concentration and total dietary calcium consumption rise. (Ramberg, 1995)^[60]. In reaction to decreased blood Ca levels. the parathyroid glands release parathyroid hormone (PTH). PTH controls the kidney's ultimate hydroxylation of 25hydroxycholecalciferol (25-hydroxy- D3) to 1, 25(OH)2D3. PTH speeds up the release of calcium from bone. Ten litres of colostrum produced at calving will lose 23 g of calcium in a single milking (Horst et al., 1997)^[34].

Clinical findings

Three stages of milk fever in cattle are commonly recognized and described.

Stage 1(stage of excitement)

The cow is still standing at this point. At this stage, symptoms include ataxia, stiffness in the hind legs, normal to slightly elevated rectal temperature, hypersensitivity and muscular tremor in the head and limbs.

Stage 2 (stage of sternal recumbency)

The cow appears sleepy in sternal recumbency, frequently with the head turned into the flank or the neck twisted to the side ('S' posture) (Singh and Singh, 2017)^[70]. Important findings at this stage include subnormal temperature, a noticeable increase in heart rate (about 80 bpm), a decrease in the absolute intensity of the heart sounds, a weak or

nonexistent pupillary light reaction, a weak arterial pulse, low venous pressure and ruminal stasis. A typical milk fever consequence is uterine prolapse.

Stage 3 (stage of lateral recumbency)

The cow is nearly unconscious and heart beat is elevated 120 beats per minute and pulse is barely audible. In this stage animal was in lateral recumbency due to bloat is common.

Diagnosis

The history of a recent parturition and clinical indicators are used to make the diagnosis. Adult cows typically have a calcium level of 8 to 12 mg/dl. In this condition is indicated by hypocalcemia (5 mg/dl) and high serum creatinine phosphokinase (CPK). Long-term lateral recumbency causes ischemic muscle necrosis and increases in the levels of the muscle enzymes aspartate aminotransferase (AST) and creatine phosphokinase (CPK) in the serum (AST).

Line of treatment

The best time for treatment is when the disease is still in its early stages. The incidence of the downer cow syndrome caused by persistent ischemic muscle necrosis increases with the amount of time between when the cow first gets recumbent and when it receives therapy. For cattle, 400-500 mL of 25% solution of calcium borogluconate is the usual dose for dairy cattle weighing 300-400 kg. Cows with milk fever exhibit a typical pattern of response to calcium borogluconate if the response is favorable, including:

- 1. Belching
- 2. Muscle tremor, particularly of the flanks and often extending to the whole body
- 3. Slowing and improvement in the amplitude and pressures of the pulse
- 4. Increase in the intensity of the heart sounds
- 5. Sweating of the muzzle
- 6. Defecation.

The slow intravenous method is used to provide 50% of the predicted dose, while the subcutaneous route is used for the remaining 50%. Giving a higher calcium dose intravenously is not beneficial (Doze *et al.*, 2008) ^[13]. With initial calcium therapy, composite solutions comprising calcium, phosphorus, and glucose are advised for nonresponsive and recurrence cases (Sharma *et al.*, 2009) ^[65].

Prevention and control

The incidence of hypocalcemia and milk fever in dairy cows can be significantly reduced by lowering the amount of absorbable food cations (Na, K, Ca, and Mg) and/or increasing the amount of absorbable dietary anions (Cl, sulphate, and phosphate) (Block, 1984) ^[9]. When fed to parturient cows, oral calcium formulations can significantly reduce the incidence of milk fever cases. It has been proposed that another crucial element in the emergence of milk fever is the Ca/P ratio. In the last month of pregnancy, feeding a high phosphorus/low calcium ration (3.3:1) may help prevent the condition. Prevention of milk fever also benefits from the administration of vitamin D and its metabolites.

Ketosis

Negative energy balance is the main metabolic disease known as ketosis, which is characterised by relatively high ketone body concentrations and a simultaneous drop in blood glucose

levels. Dairy cows are particularly vulnerable to poor energy balance during late pregnancy and early lactation due to a significant increase in energy demand (Youssef et al., 2010) ^[75]. Within four weeks after parturition, cows that don't adjust well to negative energy balance (NEB) may experience hyperketonemia (clinical or subclinical ketosis). During this time, a significant amount of body fat is being used as energy to support milk production. This causes a marked mobilisation of lipids and a noticeable increase in circulating non-esterified fatty acids and ketone bodies in tissues and milk. (Herdt, 2000) [32]. Increases in the prevalence of postpartum disorders such as retained placenta, milk fever, metritis, mastitis, clinical ketosis, and displaced abomasum as well as decreased milk yield and reproductive performance, severe negative energy balance in the puerperal period are all linked to subclinical ketosis, which is defined by an elevated concentration of ketone bodies in the absence of clinical signs (Duffield et al., 2009; LeBlanc, 2010; McArt et al., 2012)^{[17,} 44, 50]. These factors cause significant economic loss (Shin et al., 2015; Suthar et al., 2013) [67, 72]. According to Duffield et al. (1998)^[18], subclinical ketosis can occur at a rate of 26 to 60% and clinical ketosis at a rate of 2 to 15% (Duffield, 2000) ^[16]. High yielding cows in India are more vulnerable to ketosis than the non-descript cows that produce less milk (Thirunavukkarasu *et al.*, 2010)^[73]. Early lactation after a late gestation has been linked to a high frequency of production disorders (Mulligan and Doherty, 2008)^[54]. High NEFA or BHBA concentrations were reported to induce immune dysfunction, which in turn is a major component of inflammatory disease in the reproductive tract (LeBlanc, 2012)^[45]. At calving, a body condition score (BCS) of 3.5 or higher was linked to an increased risk of ketosis. Dairy farmers suffer financial losses as a result of a high prevalence of clinical and subclinical ketosis due to decreased milk production and a fast decline in the milk's Solid Not Fat (SNF) composition (Tufani *et al.*, 2011)^[74]. The development of ketosis during the postpartum period may be characterized by a decrease in dry matter intake around parturition, an increase in glucose needs, and insufficient propionate synthesis during the early postpartum period (Drackley and Dann. 2005) ^[14]. Due to ketosis-induced hypoglycemia, lactose synthesis and milk output decline (Simensen et al., 1990) [68].

Clinical findings

There are two different types of symptoms: nervous and intestinal. Loss of appetite (Grohn et al., 1983)^[28], a sharp decline in milk production (Mir and Malik, 2002) [52], selective feeding (Lean et al., 1991)^[43], a lack of rumination (Hungerford, 1990)^[35] constipation (Bihani, 2001)^[8] and firm mucus-coated faeces are signs of the digestive type (Bhuin and Chakrabarti, 1993)^[7]. The apparent atrophy and loss of skin flexibility caused by what is likely the removal of subcutaneous fat has given affected cows a woody look (Sharma, 2006)^[64]. According to (Bihani, 2001)^[8] urine, milk, and breath all exhibit an acetone-like odour,. Ketotic cows have been seen to exhibit neurotic symptoms such as hyperesthesia (Chakrabarti, 2006)^[11], salivation (Sharma et al., 2004) [66] licking their faces continuously, trembling, and strange walking patterns. Ketosis, which is characterised by a decrease in feed intake and milk production, is brought on by the generation of a greater quantity of ketone bodies (BHBA) as a result of insufficient fatty acid oxidation in the liver (Duffield, 2011)^[19].

Diagnosis

Diagnosis is based on the history of recent calving and clinical signs. Ketosis can be diagnosed by measuring the concentration of ketone bodies (β -hydroxybutyrate [BHBA], acetone and acetoacetate), intermediate metabolites of fatty acid oxidation present in blood, urine or milk. A definition of ketosis greater than 1,400 µmol of BHBA/L of serum and a cutoff of \geq 200 µmol/L of milk resulted in a sensitivity of 59% and a specificity of 90% (Geishauser *et al.*, 2000)^[23].

Accurate diagnosis would require estimation of blood glucose and ketone levels however, in clinical practice the Ross modification of the Rothera test has been used and described to be reliable (Fox, 1971)^[22]. The reagent consists of 99 g of ammonium sulfate mixed with 1 g of sodium nitroprusside. Approximately 1 g of this mixture is added to 5 to 7 ml of urine in a standard test tube and after dissolving 1 ml of ammonium hydroxide solution or a flake of sodium hydroxide is added. After keeping for 3 to 5 minutes, the test is read:

No color change : (negative)

Slight lavender : 1+

Deep lavender : 2+

Beet red or purple : 3+

Deep beet red or purple and opaque (strongly positive) : 4+

A milk BHBA test strip (Keto-Test) was used by the participating veterinarians for screening a milk sample from cows. The strip was dipped into the milk sample for 3 seconds, read after 1 minute and recorded according to the color scale provided with the test. The color scale shows 6 colors corresponding to nominal levels of 0, 50, 100, 200, 500, and 1,000 μ mol of BHBA/L of milk. A recording of 100 μ mol of BHBA/L of milk or higher was defined as ketosis (Berge and Vertenten, 2014) ^[6].

Line of treatment

A large number of studies have been undertaken to identify effective treatments and evaluate glucose precursors oral propylene glycol (PG), glucocorticoid, niacin, insulin, recombinant bovine somatotropin, butaphosphan, cyanocobalamin and combined therapies (McArt et al., 2011; Gohary et al., 2015; Nuber et al., 2016) [51, 26, 57]. Glucocorticoids have been used in ketosis treatment because of their ability to produce hyperglycemia as a result of changes in glucose use (Herdt and Emery, 1992)^[31]. Steroids also block the effects of insulin, allowing for increased catabolism of fat and protein stores. Plasma concentrations of both glucose and insulin increase significantly about 48 hours after injection with dexamethasone (Jorritsma et al., 2004). Insulin is used in the treatment of ketosis because of the anabolic effects of the hormone (Hayirli, 2006)^[30]. Insulin decreases fat breakdown, increases fat synthesis, and increases use of ketone bodies as energy sources, which should decrease the level and consequences of ketonemia.

Administration of vitamin B_{12} may increase gluconeogenesis by increasing the activity of methylmalonyl-coenzyme A (CoA) mutase, a vitamin B_{12} -dependent enzyme and important component of the Krebs or tricarboxylic acid (TCA) cycle (Kennedy *et al.*, 1990) ^[41]. Butaphosphan, an organic phosphorus source, has been also been used because of its presumed role in gluconeogenesis (Rollin *et al.*, 2010) ^[61].

Treatment of ketotic animals with 300 g of propylene glycol daily should be considered the base of ketosis treatment. It is generally given as an oral drench once a day. When propylene glycol enters the rumen, it is either absorbed directly or converted to propionate (Nielsen and Ingvartsen, 2004)^[56]. Propylene glycol that is absorbed directly enters the TCA cycle to increase oxidation of acetyl CoA and stimulate gluconeogenesis. Propionate from propylene glycol can also be used for gluconeogenesis and helps stimulate insulin release. There is a significant increase in insulin by 15 minutes after administration, and insulin remains increased for 2 hours or more after drenching. This spike in insulin helps decrease fat breakdown and hepatic ketone body production (Studer *et al.*, 1993)^[71].

Prevention and control

Fattening of animals during pregnancy must be avoided and the score should be maintain between 3.5 and 4.0 on 5 point scale. The pregnant animal should be kapt on balanced diet and concentrate more than 1 kg/50 kg b.wt. should not be given. The ration should have a minimum of 20% crude fibre along with supplementation of essential vitamins and minerals. Monensin is known to reduce ketosis in herd as it alters the ratio of volatile fatty acids produced by rumen fermentation. (Sauer *et al.*, 1989)^[62].

Downer cow syndrome

The condition, which most frequently manifests in the first two or three days following calving in high-producing dairy cows immediately after milk fever, is clinically recognized by persistent recumbency even after two successive calcium treatments. Following treatment for milk fever, cows who do not fully recover and stand but instead remain recumbent frequently develop ischemic necrosis of the major muscles of the pelvic limbs, injuries to the tissues around the hip joint, and lesions to the obturator muscles. The majority of the time, alert downer cows are in a recumbent position as a result of musculoskeletal or neurological injuries, such as lesions of the sciatic or obturator nerve brought on by dystocia (calving paralysis), long bone or pelvic fractures, hip luxation, or muscle damage brought on by either primary trauma or prolonged recumbency. Cows that are unsteady during parturition or who are made to stand or walk on a slick floor right before or after parturition are also more likely to sustain injuries to their musculoskeletal system. The other cause is electrolyte imbalance brought on by prolonged recumbency after delivery, such as hypophosphataemia, hypokalaemia, and hypomagnesaemia.

Clinical findings

The vast majority of the affected animals are awake and aware, ruminating, pooping, and urination normally. They try to get up with their forelimbs, but they are unsuccessful. These cows are known as creeper cows (Sharma *et al.*, 2009)^[65]. The cow's hind limbs may often stretch to the elbows on each side of the cow. The medial thigh muscles of the cow are under a great deal of pressure in this position, which can lead to ischemic necrosis, coliform mastitis, decubitus ulceration, particularly over the prominences of the hock and elbow joint, and traumatic injuries around the tuber coxae brought on by the hip slings. After 7 days, the prognosis for the reclining animals is bleak, and the majority of them pass away from myocarditis.

Diagnosis

The blood levels of calcium, phosphorus, magnesium, and glucose are all within the normal range, and the outcomes of

haematological tests typically match those of healthy cows that have recently given birth. By 18 to 24 hours after the commencement of recumbency, the levels of the enzymes creatine phosphokinase (CPK) and aspartate aminotransferase (AST) are typically noticeably raised and continue to rise during the following few days. CPK levels that keep rising are a sign of ongoing muscle injury. By 18 to 24 hours after the commencement of recumbency, a significant proteinuria is typically detectable.

Line of treatment

Along with medical management, a Downer cow's good nursing care and management are priorities. Good bedding, especially enough straw or a soft ground surface, is helpful in healing. When downer cows try to stand, the best ground surface for them to stand on is a sand or mud pack. Frequent side-to-side rolling of the affected animal is important to reduce ischemia necrosis. Supplementing with a composite solution combining calcium, magnesium, phosphorus, and electrolyte solution is a part of therapeutic management. Early healing is aided by the use of nervine tonics such vitamin B complex.

Prevention and control

The incidence and severity of downer cow syndrome will be decreased by the early diagnosis and treatment of milk fever. Every few hours, if they are unable to stand, they should be turned from side to side. Prior to calving, dairy cows should be kept in a cosy, well-bedded box stall, and if milk fever occurs, they should remain there for at least 48 hours after partition.

Conclusion

The dairy business continues to face a significant threat from metabolic illness. For dairy farmers, metabolic illness is a significant source of financial loss. Systems for managing the health and performance of dairy cows are centred on the early detection and subsequent avoidance of production disorders through the treatment of afflicted animals or by enhancing the diet of the herd. Avoid overfeeding the animals and making sudden changes to their feeding plan. Give the animals the appropriate amounts of protein, vitamins, and minerals, as well as adequate amounts of high-quality roughage.

References

- 1. Akhtar MZ, Khan A, Khan MZ, Muhammad G. Haemato-biochemical aspects of parturient haemoglobinuria in buffalo. Turkish J Vet. Anim. Sci. 2007b;31:119-123.
- Akhtar MZ, Khan A, Sarwar M, Javaid A. Influence of soil and forage minerals on buffalo parturient haemoglobinuria. Asian-Aust. J Anim. Sci. 2007;20:393-98.
- 3. Akhtar MZ. Aetiopathology of Parturient Haemoglobinuria in Buffaloes. Ph.D. Thesis. Department of Veterinary Pathology, University of Agriculture, Faisalabad; c2006.
- Banerjee GC. A Textbook of Animal Husbandry. 8th ed. Oxford and IBH Publishing Co. Pvt. Ltd, New Delhi; 1998. p. 517-536.
- 5. Bareille N, Beaudeau F, Billon S, Robert A, Farverdin P. Effects of health disorders on feed intake and milk production in dairy cows. Livst. Prod. Sci. 2003;83:53-62.

- 6. Berge AC, Vertenten G. A field study to determine the prevalence, dairy herd management systems, and fresh cow clinical conditions associated with ketosis in western European dairy herds. J Dairy Sci. 2014;97(4):2145-54.
- 7. Bhuin S, Chakrabarti A. A note on prevalence of ketosis in West Bengal. Ind. Vet. J. 1993;70(6):582-583.
- 8. Bihani DK. Clinico-biochemical studies on ketosis in cattle, Ph.D. Thesis, RAU, Bikaner; c2001.
- Block E. Manipulating dietary anions and cations for prepartum dairy cows to reduce incidence of milk fever. J Dairy Sci. 1984;67:2939.
- Brechbuhl M, Meylan M, Kunz-Kirchhofer C, Bodmer M, Michel A, Kaufmann T. Post parturient haemoglobinuria in cows kept in the Swiss Alpine region. Tierarztliche Praxis Ausgabe G: Grosstiere – Nutztiere. 2008;36:236-40.
- Chakrabarti A. Text book of clinical veterinary medicine (2nd ed.) Ludhiana: Kalyani publishers; c2006. p. 564-557.
- Choudhary D, Kolte AY, Manohar DS, Chaturvedi M, Chaudhari M, Jakhar A. Evaluation of Oral And Parentral Therapy In Recently Parturited Hypocalcaemic And Hypoglycaemic Buffaloes. Haryana Vet. 2013;52:79-81.
- 13. Doze JG, Donders R, Kolk JHVD. Effects of intravenous administration of two volumes of calcium solution on plasma ionized calcium concentration and recovery from naturally occurring hypocalcemia in lactating dairy cows. American J Vete. Res. 2008;69(10):1346-50.
- 14. Drackley JK, Dann HM. Carnitine palmitoyltransferase in liver of periparturient dairy cows: effects of prepartum intake, postpartum induction of ketosis and periparturient disorders. J Dairy Sci. 2005;88:3851-59.
- 15. Dua K. Importance of micronutrients and relevance of their supplementation in buffaloes. Pakistan J Zool. 2009;9:541-549.
- Duffield T. Subclinical ketosis in lactating dairy cattle. Vet. Clin. North Am. Food Anim. Pract. 2000;16:231-253.
- 17. Duffield TF, Lissemore KD, McBride BW, Leslie KE. Impact of hyperketonemia in early lactation dairy cows on health and production. J Dairy Sci. 2009;92:571-580.
- Duffield TF, Sandals D, Leslie KE, Lissemore K, McBride BW, Lumsden JH, *et al.* Efficacy of monensin for prevention of subclinical ketosis in lactating dairy cows. J Dairy Sci. 1998;81:2866-73.
- Duffield TF. Prevention of metabolic diseases for special patients. 63rd CVMA convention, Halifax, Nova Scotia, Canada; c2011. p. 6-9.
- 20. Durrani AZ, Kamal N, Shakoori AR, Younus RM. Prevalence of post parturient haemoglobinuria in buffalo and therapeutic trials with toldimfos sodium and tea leaves in Pakistan. Turk. J Vet. Anim. Sci. 2010;34:45-51.
- Ellison RS, Young BJ, Read DH. Bovine post-parturient haemoglobinuria: two distinct entities in New Zealand. N Z. Vet. J. 1986;34:7-10.
- 22. Fox FH. Clinical Diagnosis and Treatment of Ketosis. J Dairy Sci. 1971;54(6):974-978.
- 23. Geishauser T, Leslie K, Tenhag J, Bashiri A. Evaluation of eight cow-side ketone tests in milk for detection of subclinical ketosis in dairy cows. J Dairy Sci. 2000;83:296-299.
- 24. Ghanem MM, El-Deeb WM. Lecithin cholesterol acyltransferase (LCAT) activity as predictor for ketosis

https://www.thepharmajournal.com

and parturient haemoglobinuria in Egyptian water buffaloes. Res. Vet. Sci. 2010;88:20-25.

- 25. Goel P, Malik KS, Dwarkanath PK, Chugh SK. Role of Oxygen releasers in Post parturient haemoglobinuria in buffaloes. Indian Vet. J. 1988;69:823-26.
- 26. Gohary K, Leslie KE, Ford J, Capel M, LeBlanc SJ, Duffield TF. Effect of administration of recombinant bovine somatotropin on health and performance of lactating dairy cows diagnosed with hyperketonemia. J Dairy Sci. 2015;98:4392-400.
- 27. Grasso F, Terzano GM, Rosa G De, Quarantelli T, Serpe L, Bordi A. Influence of housing conditions and calving distance on blood metabolites in water buffalo cows. Italian J Anim Sci. 2004;3:275-282.
- Grohn Y, Lindberg LA, Bruss ML, Farver TB. Fatty infiltration of liver in spontaneously ketotic cows. J Dairy Sci. 1983;66(11):2320-28.
- 29. Gupta S, Bihani DK, Singh AP, Tanwar RK, Fakhruddin. Clinical Studies on Post Parturient Haemoglobinuria in Buffaloes. Vet. Pract. 2010;11:2.
- 30. Hayirli A. The role of exogenous insulin in the complex of hepatic lipidosis and ketosis associated with insulin resistance phenomenon in postpartum dairy cattle. Vet. Res. Commun. 2006;30(7):749-74.
- Herdt TH, Emery RS. Therapy of diseases of ruminant intermediary metabolism. Vet Clin North Am Food Anim. Pract. 1992;8(1):91-106.
- Herdt TH. Ruminant adaptation to negative energy balance. Influences on the etiology of ketosis and fatty liver. Vet. Clin. North Am. Food Anim. Pract. 2000;16:215-30.
- 33. Heuer C, Schukken YH, Dobbelaar P. Postpartum body condition score and results from the first test day milk as predictors of disease, fertility, yield and culling in commercial dairy herds. J Dairy Sci. 1999;82:295-304.
- Horst RL, Goff JP, Reinhardt TA and Buxton DR. Strategies for preventing milk fever in dairy cattle. J Dairy Sci. 1997;80:1269-1280.
- 35. Hungerford TG. Diseases of livestock: 9th ed. McGraw Hill book company, Sydney; c1990. p. 336, 344-347.
- Huzzey JM, Veira DM, Weary DM, von Keyserlingk MAG. Prepartum behavior and dry matter intake identify dairy cows at risk for metritis. J Dairy Sci. 2007;90:3220-33.
- 37. Jain RK, Saksule CM, Dhakad RK. Nutritional Status and Probable Cause of Haemoglobinuria in Advanced Pregnant Buffaloes of Indore District of Madhya Pradesh. Buffalo Bulletin. 2012;31:1.
- 38. Jalali MT, Nouri M, Rasooli A, Haji H, Shahryari A, Shirazi MR. Hepatic triacylglycerols and serum nonesterified fatty acids (NEFA) variations in indigenous water buffalo (*Bubalus bubalis*) in the province of Khuzestan, Iran. Int. J Vet. Res. 2011;5:151-155.
- 39. Jorritsma R, Thanasak J, Houweling M. Effects of a single dose of dexamethasone-21-isonicotinate on the metabolism of heifers in early lactation. Vet. Rec. 2011;155(17):521-3.
- 40. Kahn CM, Line S. The Merck Veterinary Manual, 9th edition. Merck and Co., Inc. Whitehouse station, N.J., U.S.A, 2005, 816.
- 41. Kennedy DG, Cannavan A, Molloy A. Methylmalonyl-CoA mutase (EC 5.4.99.2) and methionine synthetase (EC 2.1.1.13) in the tissues of cobaltvitamin 12 deficient sheep. Br J Nutr. 1990;64:721-32.

- 42. Khan A, Akhtar MZ. Hemato-biochemical and clinicoepidemiological aspects of parturient hemoglobinuria in Nili-Ravi buffaloes. Ital. J Anim. Sci. 2007;6:953-56.
- 43. Lean IJ, Bruss ML, Baldwin RL, Troutt HF. Bovine ketosis: a review. I. Epidemiology and pathogenesis. Vet. Bu. 1991;11(62):1209-18.
- 44. LeBlanc S. Monitoring metabolic health of dairy cattle in the transition period. J Reprod. Dev. 2010;56(S):S29-S35.
- 45. LeBlanc SJ. Interactions of metabolism, inflammation, and reproductive tract health in the postpartum period in dairy cattle. Reprod. Domest. Anim. 2012;47:18-30.
- Littledike ET, Young JW, Beitz DC. Common Metabolic Diseases of Cattle: Ketosis, Milk Fever, Grass Tetany and Downer Cow Complex1. J dairy Sci. 1981;64(6):1465-82.
- 47. Mahmood A, Khan MA, Younus M, Khan MA, Ahad A, Ahmad M, *et al.* Haematological and Biochemical Risk Factors Of Parturient Haemoglobinuria In Buffaloes. J Anim. Plant Sci. 2013;23:364-68.
- 48. Mata MM, Bhardwaj RM. Possible alterations in erythrocyte metabolism and integrity in post-parturient haemoglobinuria. Ind, J Vet. Med. 1985;5:67-72.
- 49. McArt JA, Nydam DV, Oetzel GR. A field trial on the effect of propylene glycol on displaced abomasum, removal from herd, and reproduction in fresh cows diagnosed with subclinical ketosis. J Dairy Sci. 2012;95:2505-12.
- McArt JA, Nydam DV, Oetzel GR. Epidemiology of subclinical ketosis in early lactation dairy cattle. J Dairy Sci. 2012;95:5056-66.
- 51. McArt JAA, Nydam DV, Ospina PA, Oetzel GR. A field trial on the effect of propylene glycol on milk yield and resolution of ketosis in fresh cows diagnosed with subclinical ketosis. J Dairy Sci. 2011;94:6011-20.
- Mir AQ, Malik HU. Prevalence and Clinico-pathological studies in bovine ketosis. National symposium and XX ISVM convention, Bikaner, 2002, 125.
- 53. Monteiro BM, Yasouka MM, Pogliani FC, Ayres H, Viana RB, Birgel EH. Lipid and glucose profiles of dairy buffaloes during lactation and dry period. Revue de Ciencia Agraria. 2012;55:33-39.
- 54. Mulligan FJ, Doherty ML. Production diseases of the transition cow. Vet. J. 2008;176:3-9.
- 55. Neto B, Oliveira C, Duarte D, Albernaz T, Júnior DO, Riet-Correa G, *et al.* Phosphorus deficiency in buffaloes in the state of Pará, Northern Brazil. Italian J Anim. Sci. 2007;6:971-973.
- 56. Nielsen NI, Ingvartsen KL. Propylene glycol for dairy cows. Anim. Feed Sci. Technol. 2004;115(3-4):191-213.
- 57. Nuber U, Van Dorland HA, Bruckmaier RM. Effects of butafosfan with or without cyanocobalamin on the metabolism of early lactating cows with subclinical ketosis. J Anim Physiol. Anim. Nutr. 2016;100:146-55.
- Purohit GN, Gaur M, Saraswat CS, Dihani BK. Metabolic disorders in the parturient buffalo. In: Bubaline Theriogenology. Eds: GN Purohit, International Veterinary Information Service, Ithaca NY (www.ivis.org), Last updated: 8-Oct-2013; A5721.1013
- Radostits OM, Gay CC, Hinchcliff KW, Constable PD. Veterinary Medicine: A Textbook of the Diseases of Cattle, Sheep, Pigs, Goats and Horses, 10th Ed. Elsevier's Health Sciences 1600 John F. Kennedy Boulevard, Suite 1800, Philadelphia, PA 19103 – 2899, U.S.A.; c2007. p.

1682-83.

- Ramberg CF. Kinetic overview: modeling calcium metabolism in pregnant and lactating cows. In: Siva Subramanian, K.N., Wastney, M.E. (Eds.), Kinetic Models of Trace Element and Mineral Metabolism During Development. CRC Press, Boca Raton, Florida, USA; 1995. p. 11-28.
- 61. Rollin E, Berghaus RD, Rapnicki P. The effect of injectable butaphosphan and cyanocobalamin on postpartum serum beta-hydroxybutyrate, calcium, and phosphorus concentrations in dairy cattle. J Dairy Sci. 2010;93(3):978-87.
- 62. Sauer FP, Kramer JKG, Cantwell WJ. Antiketogenic effects of monensis in early lactation. J dairy Sci. 1989;72:436.
- 63. Sebastian L, Mudgal VD, Nair PG. Comparative efficiency of milk production by Sahiwal cattle and Murrah buffalo. J Anim. Sci. 1970;30:253-256.
- 64. Sharma BL. Studies on some biochemical and hormonal changes in ketotic cows in Bikaner region. M.V.Sc.; Thesis, RAU, Bikaner, 2006.
- 65. Sharma MC, Kumar M, Sharma RD. Metabolic diseases in textbook of clinical veterinary medicine. first edition, ICAR, NEW DELHI 110002, 2009, 436-469.
- 66. Sharma T, Dixit SK, Singh AP. Metabolic aspects of ketosis in relation with toxicity and management. Vet. Pract. 2004;5(1):37-39.
- 67. Shin EK, Jeong JK, Choi IS, Kang HG, Hur TY, Jung YH. Relationships among ketosis, serum metabolites, body condition, and reproductive outcomes in dairy cows. Theriogenol. 2015;84:252-60.
- 68. Simensen E, Halse K, Gillund P, Lutnaes B. Ketosis treatment and milk yield in dairy cows related to milk acetoacetate levels. Acta Vet. Scand. 1990;31:433-440.
- 69. Singh N, Kumari R, Akbar MA. Biochemical-changes in blood metabolites in buffalos with indigestion. Indian Vet. J. 1989;66:923-26.
- 70. Singh RSV, Singh NK. Impact of metabolic diseases on milk production in lactating cows: A review. The Blue Cross Book. 2017;36:11-18.
- 71. Studer VA, Grummer RR, Bertics SJ. Effect of prepartum propylene glycol administration on periparturient fatty liver in dairy cows. J Dairy Sci. 1993;76(10):2931-9.
- 72. Suthar VS, Canelas-Raposo J, Deniz A, Heuwieser W. Prevalence of subclinical ketosis and relationships with postpartum diseases in European dairy cows. J Dairy Sci. 2013;96:2925-38.
- Thirunavukkarasu M, Kathiravan G, Kalaikannan A, Jebarani W. Prevalence of ketosis in dairy farm. Tamilnadu J Vet. Anim. Sci. 2010;6(4):193-195.
- 74. Tufani NA, Hafiz A, Muhee A, Makhdoomi DM. Therapeutic management of ketosis in bovine. Indian J Vet. Med. 2011;31:38-39.
- 75. Youssef MA, El-Khodery SA, El-deeb WM, El-Amaiem WE. Ketosis in buffalo (*Bubalus bubalis*): clinical findings and the associated oxidative stress level. Tropical animal health and production. Trop. Anim. Health Prod. 2010;42(8):1771-7.