www.ThePharmaJournal.com

The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23

TPI 2022; SP-11(9): 350-353 © 2022 TPI

www.thepharmajournal.com Received: 09-07-2022

Received: 09-07-2022 Accepted: 13-08-2022

Shivakumar R

Ph.D. Scholar, Department of Veterinary Microbiology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

S Wilfred Ruban

Associate Professor and Head, Department of Livestock Products Technology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

Srikrishna Isloor

Professor, Department of Veterinary Microbiology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru. Karnataka. India

D Rathnamma

Professor & Head, Department of Veterinary Microbiology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

Ahlen IG Barry

Junior Research Fellow, Department of Livestock Products Technology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

Sharada R

Associate Professor, Department of Veterinary Microbiology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

Sudarshan S

Assistant Professor, Department of Livestock Products Technology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

HB Shivappa Nayaka

Ph.D. Scholar, Department of Poultry Science, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

Corresponding Author: Shivakumar R

Ph.D. Scholar, Department of Veterinary Microbiology, Veterinary College, Karnataka Veterinary Animal and Fisheries Sciences University, Bengaluru, Karnataka, India

Antibiogram pattern of *E. coli* isolated from healthy adult pigs in pig farms located in and around Bengaluru, India

Shivakumar R, S Wilfred Ruban, Srikrishna Isloor, D Rathnamma, Ahlen IG Barry, Sharada R, Sudarshan S and HB Shivappa Nayaka

Abstract

A cross sectional study was carried out to evaluate the antimicrobial susceptibility pattern of E. coli isolated from healthy adult pigs from 10 organized pig farms located in and around Bengaluru, Karnataka, India. Faecal samples from 40 pigs (4 adult pigs from each farm) were collected and were used for isolation and characterization of E. coli based on biochemical methods. E. coli was isolated from all the samples. A total of 104 E. coli isolates obtained from healthy adult pigs and were screened for antimicrobial susceptibility testing based on disc diffusion assay against 13 commonly used antimicrobials in human and veterinary medicine. Out of the 104 E. coli isolates from adult pig fecal samples, 95.19 per cent isolates exhibited resistance towards doxycycline (Tetracyclines) followed by Enrofloxacin (70.19%), ciprofloxacin (66.35%), ampicillin (61.54%), cotrimoxazole (56.73%), Cefotaxime + clavulanic acid (34.62%), chloramphenicol (33.65%), ceftazidime + clavulanic acid (24.04%), cefpodoxime + clavulanic acid (23.08%), amoxicillin (13.46%), neomycin (4.81%) and gentamycin (2.88%). In this study, all the E. coli isolates tested were found to be susceptible to amikacin, imipenem and meropenem. The phenotypic characterization confirmed the presence of Extended spectrum β-lactamase E. coli at 34.62 per cent. In all, 86.54 per cent (90/104) of E. coli isolates were Multidrug resistant (MDR) were being resistant to one antimicrobial agent in three or more antimicrobial classes. The study clearly indicated higher presence of multidrug resistant E. coli in healthy adult pigs indicating its public health significance and potential of such adult pigs being a major source in dissemination of AMR E. coli in the entire pork production chain.

Keywords: AMR, E. coli, adult pigs, multidrug resistant, antibiotics

Introduction

The ever increasing human population in India and in other parts of the world has triggered an increase in demand for food from agriculture and animal products to cater for economic, social and dietary demand of the population. This has resulted in producers shifting from low profit, low productivity traditional extensive system of animal rearing to intensive production in both small and large scale as it is considered to be a more profitable way of ensuring animal welfare and a solution for food security worldwide (Robinson *et al.*, 2011) [22]. However, with all the advantages the intensive animal production carries it has also been implicated with the development and spread of zoonotic diseases, antimicrobial resistance (AMR), environmental change resulting from pressure on natural resources and change in livelihoods of small and marginal farmers (Otte *et al.*, 2019) [20].

Antimicrobial resistance (AMR) is one of the major and growing public health threat worldwide and indiscriminate use of antimicrobials in food-producing animals for prophylaxis and as growth promoter has been documented to be the primary reason in emergence and spread of AMR in animal production system (Lim *et al.*, 2014) [15]. In developing countries like India, rearing of livestock especially pigs and poultry, which plays an important role in improving the socio-economic status of the poor and marginal farmers has been practiced as family subsistence system or under backyard system with lower inputs in terms of feed and lower antimicrobial usage (Vinodh Kumar *et al.*, 2019) [24]. In the recent past, there has been a significant transformation in pig farming in India from backyard system of rearing to small/medium or larger commercial farms owing to the high demand for animal protein. However, with the increase in scale of productivity, animals have been exposed to various infections and hence use of antibiotics is often regarded as the simplest way to maintain healthy and productive animals (Manyi-Loh *et al.*, 2018) [18].

In addition, over the counter availability of antimicrobials have also contributed to the extensive use of antimicrobials by the livestock owners, which in turn has contributed to the emergence of AMR in livestock production system (Kotwani *et al.*, 2021) ^[10].

Escherichia coli is an abundant commensal enteric bacterium in intestinal tract of domestic animals and has been used as an indicator organism for studying AMR in the food chain as it is found in all the compartments of the production system. It has been documented that commensal bacteria in gastrointestinal tract of both animals and humans are considered to be good indicators of AMR as they are exposed to selection pressure driven by any antimicrobial use in the host (Blake et al., 2003) [2]. However, the role of these commensal bacteria in carriage of AMR in healthy adult pigs has been neglected. Hence, the present study has been carried out to evaluate the occurrence of antibiotic resistance in fecal E. coli isolates from healthy adult pigs in commercial pig farms in and around Bengaluru, Karnataka, India.

Materials and Methods

The present cross sectional study was carried out in 10 commercial pig farms located in and around Bengaluru, Karnataka, India. A total of 40 adult pigs (4 each from one farm) were used in this study. Rectal swabs were taken from these adult pigs using sterile cotton swabs and were transported to the laboratory for isolation and characterization of E. coli. Isolation of E. coli was done as per the standard procedure (Quinn et al. 2002) [20]. In brief, samples were preenriched in Brain heart infusion and enriched for 8-10 hr at 37 °C. The enriched samples were streaked onto Eosin Methylene Blue and MacConkey agar (Himedia, India) and isolates were gram stained and identified up to species level using standard bacteriological techniques, including colony morphology on MacConkey and Eosin Methylene Blue Agar, oxidase, catalase, urease and indole tests as per Quinn et al. (2002) [20]. The E. coli isolates were then subjected to antibiotic susceptibility testing of isolates to a panel of 13 antimicrobial agents (Gentamicin (GEN = 10 µg) Amikacin $(AK = 30 \mu g)$, Neomycin $(N = 10 \mu g)$, Ciprofloxacin $(CIP = 5 \mu g)$ μ g), Enrofloxacin (EX = 5 μ g), Doxycycline (DO = 30 μ g), Trimethoprim-sulfamethoxazole {COT = $25(23.75/1.25 \mu g)$ }, Chloramphenicol (C = $30 \mu g$), Ampicillin (AMP = $10 \mu g$), Amoxicillin + Clavulanic acid (AMC = 20/10 μg), Imipenem (IMP=), Meropenem (MRP=), Cefotaxime (CTX = 30 µg), Cefotaxime + Clavulanic acid (CEC = $30/10 \mu g$)) using the Kirby-Bauer method (disc diffusion method) (Bauer et al. 1966) [25]. Interpretation of the results was carried out as per European committee on Antimicrobial Susceptibility Testing (EUCAST, 2020) [8] and Clinical and Laboratory Standard Institute (CLSI, 2018) [5] wherever the EUCAST breakpoints were not available. There are no breakpoints for Doxycycline, and instead the breakpoint for Tetracycline were used. The standard reference strain of E. coli ATCC 25922, was used as the quality control strain. Intermediate isolates were grouped with resistant isolates. Resistance profiles were generated and isolates were classified as MDR if they showed resistance to one antimicrobial agent in three or more antimicrobial classes (Magiorakos et al., 2012) [16].

Result and discussion

In the present study, *E. coli* was isolated from all the rectal samples from the healthy adult pigs from all the farms, as *Escherichia coli* is a commensal flora in human and animal

intestines (Dohmen *et al.*, 2017) ^[7]. A total of 104 *E. coli* were isolated from 40 rectal samples and similar isolation have been carried out Lalzampuia *et al.* (2013) ^[14] who isolated 102 *E. coli* from 53 fecal samples from pigs in Mizoram and Lalruatdiki *et al.* (2018) ^[13] who isolated 867 *E. coli* from 228 fecal samples from pigs of Meghalaya and Assam. The isolation rate of *E. coli* varies with the type of samples, media used, isolation protocol and person to person variation. In this study, 3-4 suspected colonies were randomly picked and used for identification based on biochemical characterization.

Antibiotics especially with broad spectrum of activity are extensively used in intensive pig production system for effective disease control, to ensure good health and welfare of the animals, which results in emergence of AMR (Diana et al., 2017) [6]. The results of the Antibiogram pattern of E. coli isolated from healthy adult Pigs indicated that out of the 104 E. coli isolates, 95.19 per cent isolates exhibited resistance doxycycline (Tetracyclines) Enrofloxacin (70.19%), ciprofloxacin (66.35%), ampicillin (61.54%). The results of the study are in concurrence with the findings of Abubakar et al. (2019) [1] in adult pigs (73.1%) of Pretoria, Van Den Bogaard et al. (2000) [23] in Netherland who observed higher resistance towards tetracycline. In similar lines, Cho et al. (2006) [4] and Lim et al. (2014) [15] in Korea reported that the rate of resistance of E. coli from pigs to Tetracycline was the highest (97.8%), followed by Ampicillin (89.1%). The higher resistance observed in this study may be attributed to the wider use of tetracyclines and fluoroquinolones in the treatment of commonly observed pig diseases, their ease of availability drug over the counter without any prescription and broad spectrum of activity.

Table 1: Antibiogram pattern of *E. coli isolated* from healthy adult pigs from pig farms in and around Bengaluru

Class of antibiotic	Name of antibiotics	E. coli isolates from piglets (n=104)	
		Sensitive	Resistant
Aminoglycosides	GEN	101 (97.12)	3 (2.88)
	AK	104 (100)	0.00
	N	99 (95.19)	5 (4.81)
Fluoroquinolones	CIP	35 (33.65)	69 (66.35)
	EX	31 (29.81)	73 (70.19)
Tetracycline	DO	5 (40.81)	99 (95.19)
Folate pathway inhibitors	COT	45 (43.27)	59 (56.73)
Phenicols	С	69 (66.35)	35 (33.65)
Penicillin/ β-	AMP	40 (38.46)	64 (61.54)
lactamase inhibitors	AMC	90 (86.54)	14 (13.46)
Carbapenems	IMP	104 (100)	0.00
	MRP	104 (100)	0.00
Extended-spectrum β-lactamase	CTX-CEC	68 (65.38)	36 (34.62)

The isolates in the present study revealed complete sensitivity to carbapenem class of antibiotics (Imipenem and meropenem) and Amikacin followed by Gentamicin (97.12%0, Neomycin (95.19%), Amoxicillin + Clavulanic acid (86.54%) and chloramphenicol (66.35%). Similar observations have been documented by Kyung-Hyo *et al.* (2020) [11], who have reported that ban on use of antibiotics as growth promoter in pigs caused a decrease in resistance to gentamicin, neomycin, ciprofloxacin, norfloxacin and amoxicillin/clavulanic acid in *E. coli* isolated from in Korea. This is supported by Liu *et al.* (2014) [15] who opined that

AMR is dependent on the level of antimicrobial usage and in the present study farms aminoglycosides and chloramphenicol were seldom used in treatment of pigs substantiates its low resistance or higher sensitivity.

The prevalence of ESBL E. coli in the healthy adult pigs in this study was 34.62 per cent. Extended spectrum βlactamases (ESBLs) have been reported worldwide, most frequently in Enterobacteriaceae (Lalak et al., 2016) [12]. Similar prevalence have been recorded in pigs and piglets by Dohmen et al. (2017) [7] in Netherland (24.7%) and Galina et al. (2021) [9] in Lativa (26.9%). However, contrary to the findings of this study Mandakini et al. (2020) [17] observed that 65.89 per cent of E. coli isolates from organized and unorganized pig farms in Meghalaya were ESBLs producer and VinodhKumar et al. (2019) [24] who reported 64 per cent prevalence of ESBL E. coli in organized pig farms across India. In this study we observed that none of the farms used cephalosporins for treatment of the pigs. Among the isolates screened 86.54 per cent (90/104) of the isolates were multidrug resistant (MDR) being resistant to one antimicrobial agent in three or more antimicrobial classes tested. Similarly higher rate of MDR E. coli have been reported by Sanjukta et al. (2012) in apparently health pigs in North East (90%), Kyung-Hyo et al. (2020) [11] in weaned piglets (95%), Momtaz et al. (2012) [19] in commercial chicken meat (64.91%) and Brower et al. (2017) [3] in layer chicken (60%).

Conclusion

The results of the present study indicated that commensal *E. coli* from apparently healthy adult pigs were found to be resistant to antibiotics commonly used in human medicine and occurrence of resistance towards cephalosporins (ESBL) are of major concern as these isolates may be involved in dissemination of AMR to farm workers. In addition, presence of MDR *E. coli* in pigs indicates that animals are being exposed to varying environment and the resistance could have been environmentally acquired, which requires further investigation.

Acknowledgements

The author duly acknowledges the Karnataka Veterinary, Animal and Fisheries Sciences University and Veterinary College, Hebbal, Bangalore for providing facilities for conduct of the research. This study is part of Ph.D thesis submitted by the first author to Tamil Nadu KVAFSU, Bidar.

References

- Abubakar RH, Madoroba E, Adebowale O, Fasanmi OG, Fasina FO. Antimicrobial usage in pig production: Effects on Escherichia coli virulence profiles and antimicrobial resistance. The Onderstepoort Journal of Veterinary Research. 2019;86(1):e1-e11.
- 2. Blake DP, Hillman K, Fenlon DR, Low JC. Transfer of antibiotic resistance between commensal and pathogenic members of the Enterobacteriaceae under ileal conditions. J Appl. Microbiol. 2003;95:428-436.
- 3. Brower CH, Mandal S, Hayer S, Sran M, Zehra A, Patel SJ, Kaur R, *et al.* The Prevalence of Extended-Spectrum Beta-Lactamase-Producing Multidrug-Resistant *Escherichia coli* in Poultry Chickens and Variation According to Farming Practices in Punjab, India. Environ Health Perspect. 2017;125(7):077015.
- 4. Cho JK, Ha JS, Kim KS. Antimicrobial drug resistance of

- Escherichia coli isolated from cattle, swine and chicken. Korean J Vet. Public Health. 2006;30:9-18.
- CLSI. Performance standards for antimicrobial disk and dilution susceptibility tests for bacteria isolated from animals. CLSI document VET01 5th edition. Jun 2018, Clin. Lab. Stand. Inst; c2018.
- 6. Diana A, Manzanilla EG, Díaz JAC, Leonard FC, Boyle LA. Do weaner pigs need in-feed antibiotics to ensure good health and welfare? PLoS One. 2017;12:e0189434.
- 7. Dohmen W, Dorado-Garcia A, Bonten MJ, Wagenaar JA, Mevius D, Heederik DJ. Risk factors for ESBL-producing *Escherichia coli* on pig farms: A longitudinal study in the context of reduced use of antimicrobials. PLoS One. 2017;12(3):e0174094.
- 8. EUCAST, European Committee on Antimicrobial Susceptibility Testing Breakpoint tables for interpretation of MICs and zone diameters; c2020. p. 0-77.
- 9. Galina D, Balins A, Valdovska A. The Prevalence and Characterization of Fecal Extended Spectrum-Beta-Lactamase-Producing *Escherichia coli* Isolated from Pigs on Farms of Different Sizes in Latvia. Antibiotics. 2021 Sep 11;10(9):1099.
- Kotwani A, Joshi J, Lamkang AS. Over-the-Counter Sale of Antibiotics in India: A Qualitative Study of Providers' Perspectives across Two States. Antibiotics. 2021 Sep 17;10(9):1123.
- Kyung-Hyo D, Jae-Won B, Wan-Kyu L. Antimicrobial Resistance Profiles of *Escherichia coli* from Diarrheic Weaned Piglets after the Ban on Antibiotic Growth Promoters in Feed. Antibiotics. 2020 Oct 29;9(11):755.
- 12. Lalak A, Wasyl D, Zajac M, Skarzynska M, Hoszowski A, Samcik I, *et al*. Mechanisms of cephalosporin resistance in indicator *Escherichia coli* isolated from food animals. Vet. Microbiol. 2016 Oct 15;194:69-73.
- 13. Lalruatdiki A, Dutta TK, Roychoudhury P, Subudhi P.K. Extended-spectrum b-lactamase producing multidrug resistant *Escherichia coli*, *Salmonella* and *Klebsiella pneumonia* in pig population of Assam and Meghalaya, India. Vet. World. 2018;11(6):868-873.
- 14. Lalzampuia H, Dutta TK, Warjri I, Chandra R. PCR based detection of extended-spectrum b-lactamases (blaCTX-M and blaTEM) in *Escherichia coli*, *Salmonella* spp, *Klebsiella pneumoniae* isolated from pigs in North-Eastern India (Mizoram) Indian J Microbiol. 2013;53(3):291-296.
- 15. Lim SK, Lee JE, Lee HS, Nam HM, Moon DC, Jang GC, *et al.* Trends in antimicrobial sales for livestock and fisheries in Korea during 2003–2012. Korean J Vet. Res. 2014;54(2):81-86.
- 16. Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, *et al*. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. Clin Microbiol Infect. 2012;18(3):268-81.
- 17. Mandakini R, Roychoudhury P, Subudhi PK, Kylla H, Samanta I, Bandyopadhayay S, *et al.* Higher prevalence of multidrug-resistant extended-spectrum β-lactamases producing *Escherichia coli* in unorganized pig farms compared to organized pig farms in Mizoram, India. Veterinary world. 2020;13(12):2752-2758.
- 18. Manyi-Loh, C, Mamphweli S, Meyer E, Okoh A. Antibiotic Use in Agriculture and Its Consequential Resistance in Environmental Sources: Potential Public

- Health Implications. Molecules (Basel, Switzerland), 2018;23(4):795.
- 19. Momtaz H, Rahimi E, Moshkelani S. Molecular detection of antimicrobial resistance genes in *E. coli* isolated from slaughtered commercial chickens in Iran. Vet. Med. 2012;57(4):193-197.
- Otte J, Pica-Ciamarra U, Morzaria S. A comparative overview of the livestock-environment interactions in Asia and Sub Saharan Africa. Front. Vet. Sci. 2019 Feb 22;6:37.
- 21. Quinn PJ, Markey BK, Carter ME, Donnelly WJC, Leonard FC. Blackwell Scientific Publications; Oxford, London: Veterinary Microbiology and Microbial Diseases; c2002. p. 240-245.
- Robinson TP, Thornton PK, Franceschini G, Kruska, Chiozza F, Notenbaert AM. Global Livestock Production Systems; FAO: Rome, Italy; ILRI: Nairobi, Kenya; c2011.
- 23. Van Den Bogaard AE, London N. Stobberingh EE. Antimicrobial resistance in pig faecal samples from the Netherlands (five abattoirs) and Sweden. The Journal of Antimicrobial Chemotherapy. 2000;45(5):663-671.
- 24. VinodhKumar O, Singh B, Sinha D, Pruthvishree B, Tamta S, Dubal Z, *et al.* Risk factor analysis, antimicrobial resistance and pathotyping of *Escherichia coli* associated with pre- and post-weaning piglet diarrhoea in organised farms, India. Epidemiology and Infection. 2019;147:E174.
- 25. Bauer AW. Antibiotic susceptibility testing by a standardized single disc method. Am J clin pathol. 1966;45:149-58.