www.ThePharmaJournal.com

# The Pharma Innovation



ISSN (E): 2277-7695 ISSN (P): 2349-8242 NAAS Rating: 5.23 TPI 2022; SP-11(7): 279-282 © 2022 TPI

www.thepharmajournal.com Received: 13-05-2022 Accepted: 16-06-2022

## Pooja Solanki

Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

#### S Choudhary

Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

#### AP Singh

Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

## JP Kachhawa

Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

## Rajkumar Soni

Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

## Savita

Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

## Corresponding Author

Pooja Solanki Department of Clinical Veterinary Medicine Ethics and Jurisprudence, College of Veterinary and Animal Science, RAJUVAS, Bikaner, Rajasthan, India

## Prevalence of subclinical mastitis in dairy cows in the area of Bikaner district of Rajasthan

## Pooja Solanki, S Choudhary, AP Singh, JP Kachhawa, Rajkumar Soni and Savita

## Abstract

The objective of this study was to determine the prevalence of subclinical mastitis in dairy cows of the area of Bikaner district of Rajasthan. A total of 392 quarters milk sample of 100 HF crossbred cattle were collected for study which were subjected to physical examination and afterward screened for subclinical mastitis using four indirect tests *viz*. modified california mastitis test (MCMT), somatic cell count, electric conductivity and pH. Overall prevalence of subclinical mastitis (SCM) in 100 lactating dairy cows found in this study on the basis of MCMT, SCC, EC and pH were 53 per cent, 57 per cent, 51 per cent and 49 per cent on animal basis and 29.33 per cent, 32.14 per cent, 28.57 per cent, 27.29 per cent, respectively on quarter basis. Hind quarters were found more affected than fore quarters.

Keywords: Dairy cows, prevalence, subclinical mastitis

## Introduction

Mastitis is an inflammation of the mammary gland that can be caused by various physical or chemical agents but the majority of the causes are infectious and usually caused by bacteria that invade the udder, multiply and produce several toxins that are harm to the mammary gland (Radostits et al., 2007) [15]. The disease is complex and multi-factorial, and its occurrence is influenced by a host of variables, including the animal, the pathogen, and the environment. Mastitis occurs in two forms; clinical and subclinical mastitis. Subclinical mastitis is a nonobservable form of mastitis, such as no visible abnormalities in udder tissues as well as in milk (Kathiriya et al., 2014)<sup>[9]</sup>. Sub clinical mastitis remains a herd problem, without visible clinical signs or changes in milk grossly, which may be detected by indirect testing like the Modified California Mastitis Test (MCMT), total somatic cell count (TSCC), electrical conductivity (EC) and a cultural examination of the milk. Staphylococcus spp. is found to be the main etiological agents of clinical and subclinical mastitis in cows while, Staphylococcus aureus and E. coli are most frequently isolated pathogen from the clinical mastitis, coagulase negative Staphylococci (CNS) are the most commonly isolated pathogens from the subclinical mastitis (Contreras et al., 2003)<sup>[4]</sup>. The dairy industry suffers much greater losses due to subclinical mastitis (SCM) than clinical mastitis (Kader et al., 2002)<sup>[7]</sup>. Many risk factors have been identified for clinical and subclinical mastitis in the dairy animals such as breed, parity, age, stage of lactation, nutritional status, physical condition of teat, udder hygiene, genetic factor and housing system (Constable et al., 2017)<sup>[3]</sup>. It is important to determine the prevalence of SCM in dairy herds and also to demarcate the main factors responsible for it.

## Materials and Methods

## **Collection of milk samples**

For the present study, 100 apparently healthy lactating cattle from private dairy farms, Livestock Research Station, Veterinary Clinical Complex of College of Veterinary and Animal Science, Bikaner and animals of individual holding in and around the Bikaner were screened for subclinical mastitis. The tests performed immediately after collection of milk. Udder and teats were washed with water and air-dried and then wiped off by spirit swab. First three-four stripping of fore milk were discardedand then 10 ml of milk taken into sterilized test tubes with rubber cap.

## Physical examination of milk samples

Physical examination performed immediately after collection of milk samples to determine any abnormalities in colour, odour, consistency and presence of flakes, clot, blood and any other visible abnormalities.

## Modified California Mastitis test

The MCMT was carried out on milk samples using the Schalm and Noorlander method (1957)<sup>[18]</sup>. In present study, as an anionic active surface agent, Ezee was used in place of aryl sulphates (Chahar, 2001 and Savita, 2016)<sup>[2, 17]</sup>.

The test was carried out with 3 ml of milk samples collected from each quarter into the respective 4 cups in CMT paddle. Then equal amount of MCMT reagent was added to each cup of paddle. The content was mixed by gentle circular motion of paddle in the horizontal plane. Depending on the degree of precipitation and formation of gel, readings of positive test were categorized as negative, trace, weak positive (+), distinct positive (++) and strong positive (+++), respectively.

## Somatic Cell Count

About 0.01 ml (10  $\mu$ l) milk was withdrawn with the help of micropipette and spread evenly in 1 square cm area on a grease free glass slide. The smear was allowed to dried in air. Then, a few drops of xylene poured over the milk smear. Then, the smear was air dried and fixed with 99 per cent methanol for 2 minutes and washed with distilled water. After fixing, the smear was stained with Giemsa stain for 30 minutes. Then smear was kept in phosphate buffer solution (pH 7.0) in coupling jar for 5 minutes and bloat dried. This smear was used for somatic cell count under oil immersion and the cells were counted in total 20 fields. The average number of cells (per square cm) area was calculated. For counting of cells per ml of milk the average numbers of cells per field were multiplied by microscopic factor.

## **Electric Conductivity**

Electrical conductivity of milk samples was determined by Pen type EC-035 (ATC) Conductivity meter of ERMA instruments.

## pН

Milk pH was measured by using single electrode Pen type digital pH meter immediately.

## Statistical analysis

Statistical analysis of the collected observations was done on

the basis of the methods described by Snedecor and Cochran (1989)<sup>[23]</sup> and by using SPSS 20.0.0 version.

## **Results and Discussion**

In the present study, a total number of 100 HF crossbred cows were screened on the basis of MCMT, SCC, EC and pH for subclinical mastitis. Out of 400 quarters of 100 cows 392 functional quarters were examined during the study. Based on MCMT prevalence of sub-clinical mastitis was 53 per cent (53/100) on animal basis and 29.33 per cent (115/392) on quarter basis (Table 1). Prevalence of present study was accordance to the study of Tanwar *et al.* (2001) <sup>[26]</sup>, Kumar (2010) <sup>[10]</sup>, Marwaha (2018) <sup>[11]</sup> and Kachhawa (2019) <sup>[6]</sup> wherein they reported animal-wise prevalence as 54 per cent, 54.09 per cent, 49.78 per cent and 48 per cent, respectively and Kumar (2010) <sup>[10]</sup>, Saidi *et al.* (2013) <sup>[16]</sup>, Mir *et al.* (2014) <sup>[13]</sup>, Savita (2016) <sup>[17]</sup> and Kachhawa (2019) <sup>[6]</sup> reported quarter-wise prevalence 31.02 per cent, 29.20 per cent, 30.73 per cent, 29.50 per cent and 27.31 per cent, respectively.

Prevalence of subclinical mastitis on basis of somatic cell (SCC) count was recorded as 57 per cent (57/100) on cow basis and 32.14 per cent (126/392) on quarter basis (Table 1). SCC is reported to be the most reliable test as it is the closest to bacteriological results by Sharma *et al.* (2010) <sup>[20]</sup>. As SCC rises in the first few days of lactation (Atakan, 2008) <sup>[1]</sup> and may continue to rise up to the end of the first month, it is considered physiological that the level increases towards lactation's end.

On the basis of electric conductivity, prevalence of subclinical mastitis was 51 per cent (51/100) and 28.57 per cent (112/392) animal-wise and quarter-wise, respectively (Table 1). The almost similar results were reported by Savita (2016) <sup>[17]</sup> and Marwaha *et al.* (2018) <sup>[11]</sup> which was 56 per cent and 52 per cent, respectively. According to Kasikci *et al.* (2012) in the detection of subclinical mastitis, EC showed comparable results with both CMT and SCC; its reliability would increase further when combined with the other methods.

According to pH, the prevalence of subclinical mastitis was recorded as 49 per cent (49/100) and 27.29 per cent (107/392) animal-wise and quarter-wise, respectively (Table 1). The prevalence of SCM in present study was similar to the study of Shahid *et al.* (2011) <sup>[19]</sup> and Sunder *et al.* (2013) <sup>[25]</sup> who reported prevalence 40.8 per cent and 48 per cent, respectively. During mastitis, the pH in the milk is higher because of increased permeability of the udder tissue to blood components like bicarbonate ions.

Diagnostic tests	Subclinical mastitis affected	Percentage	Subclinical mastitis affected	Percentage
	cows (out of 100 cows)	(%)	quarters (out of 392 quarters)	(%)
Modified California Mastitis Test	53	53	115	29.33
Somatic Cell Count	57	57	126	32.14
Electric Conductivity	51	51	112	28.57
pH	49	49	107	27.29

Table 1: Results of various diagnostic tests used for detection of subclinical mastitis (cow-wise and quarter-wise)

The prevalence of SCM affected hindquarters was 32.98 per cent which was higher than the forequarters (27.77 per cent). The prevalence of right-side quarters were more than the left side quarters which 36.47 per cent (71/195) and 24.37 per cent (48/197), respectively (Table 2). Sudhan *et al.* (2005)<sup>[24]</sup>, Joshi *et al.* (2006)<sup>[5]</sup> and Sharma *et al.* (2012)<sup>[21]</sup> also reported

that hind quarters were affected more than fore-quarters. The higher prevalence of subclinical mastitis in hindquarters than in forequarters may be due to their higher production capacity (Radostits *et al.*, 2000) and greater chances of hind quarters being soiled with urine or dung from the tail; thus, increasing their chance to get infected (Singh, 2015)<sup>[22]</sup>.

Table 2: Quarter-wise prevalence of subclinical masti	tis in cattle
---	---------------

Quarters affected	No. of affected quarters/no. of screened quarters	Percentage (%)
Right Fore	32/99	32.32
Right Hind	39/96	40.62
Left Fore	23/99	23.23
Left Hind	25/98	25.51

In the recent study, a high prevalence might be explained by the absence of dry cow therapy and post milking teat dipping, invariable hold milking practice, low culling rates of chronically ill cows, poor hygiene standards of the dairy environment, coupled with a large proportion of cows confined to zero-grazing production systems. Other factors that could influence the occurrence of SCM could be due to genetic variation in disease resistance amongst the age, breeds, immune responses, milk production capacity, udder conformation, teat structure, use of different methods of diagnosing of subclinical mastitis and the definition of infection (Mdegela *et al.*, 2005)<sup>[12]</sup>.

## Conclusions

The present study concluded that prevalence of subclinical mastitis in area of Bikaner was found to be high. There may be differences in prevalence rates of SCM among different dairy herds as a result of different management and hygienic practices associated with several animal factors such as population size, breed, climatic conditions, lactation stage, milk yield, udder morphology and sanitation.

## References

- 1. Atakan KO. Factors influencing daily yield, somatic cell count and non-fat dry matter content of milk. Indian Veterinary Journal. 2008;85:630-2.
- Chahar A. Studies on some epidemiological and diagnostic aspects of bovine subclinical and clinical mastitis. PhD diss., Ph.D. Thesis submitted to Rajasthan Agricultural University, Bikaner, 2001.
- 3. Constable PD, Hinchcliff KW, Done SH, Grünberg W. Veterinary Medicine. A textbook of diseases of cattle, horses, sheep, pigs and goats. 11th ed. 3251 Riverport Lane St. Louis, Missouri 63043, 2017, 1912.
- 4. Contreras A, Luengo C, Sanchez A, Corrales JC. The role of intramammary pathogens in dairy goats. Livestock Production Science. 2003;79:273-283.
- 5. Joshi S, Gokhale S. Status of mastitis as an emerging disease in improved and periurban dairy farms in India. Annals of the New York Academy of Sciences. 2006;1081(1):74-83.
- 6. Kachhawa JP, Marwaha S, Savita, TC, Singh AP, Chahar A, Gupta SR. Prevalence study on subclinical mastitis in cross-bred cattle in Bikaner city of Rajasthan. 2019;49:27-31.
- Kader MA, Samad MA, Saha S, Taleb MA. Prevalence and aetiology of sub-clinical mastitis with antibiotic sensitivity to isolated organisms among milch cows in Bangladesh. Indian Journal of Dairy Science. 2002;55:218-223.
- KAŞIKÇI G, ÇEtİn Ö, BİNGÖL, EB, GÜndÜz MC. Relations between electrical conductivity, somatic cell count, California mastitis test and some quality parameters in the diagnosis of subclinical mastitis in dairy cows. Turkish Journal of Veterinary and Animal Sciences. 2012;36(1):49-55.
- 9. Kathiriya JB, Kabaria BB, Saradava DA, Sanepara DP.

Prevalence of subclinical mastitis in dairy cattle in Rajkot district of Gujarat. International Journal of Science and Nature. 2014;5:433-436.

- Kumar S. Development of herbal remedies for management of subclinical mastitis in bovines. Thesis, M.V.Sc. G. B. Pant University of Agriculture and Technology, Pantnagar (India), 2010.
- 11. Marwaha S. Therapeutic studies of *Piper nigrum* in subclinical mastitis in cattle. M.V.Sc. thesis submitted to Rajasthan University of Veterinary and Animal Sciences, Bikaner, 2018.
- Mdegela RH, Karimuribo E, Kusiluka LJM, Kabula B, Manjurano A, Kapaga AM, *et al.* Mastitis in smallholder dairy and pastoral cattle herds in the urban and periurban areas of the Dodoma municipality in Central Tanzania. Livestock Research Rural Development. 2005;17(11):123.
- 13. Mir AQ, Bansal BK, Gupta DK. Subclinical mastitis in machine milked dairy farms in Punjab: prevalence, distribution of bacteria and current antibiogram. Veterinary World, 2014, 7(5).
- Radostitis OM, Gay CC, Blood DC, Hinchcliff KW. Diseases caused by viruses and Chlamydia-II. Veterinary Medicine: a Textbook of the Disease of Cattle, Sheep, Pigs, Goats and Horses. 9th ed. WB Saunders company Ltd, London, 2000, 115-1260.
- Radostits OM, Gay CC, Hinchcliff KW, Constable PD. A textbook of the diseases of cattle, horses, sheep, pigs and goats. 10th ed. Saunders, Elsevier, London, 2007, 2045-2050.
- Saidi R, Khelef D, Kaidi R. Subclinical mastitis in cattle in Algeria: Frequency of occurrence and bacteriological isolates. Journal of the South African Veterinary Association. 2013;84(1):00-00.
- 17. Savita. Studies on some aspect of subclinical mastitis in cattle. M.V.Sc. Thesis submitted to Rajasthan University of Veterinary and Animal Science, 2016.
- Schalm OW, Noorlander DO. Experiments and observations leading to the development of the California mastitis test. Journal of American Veterinary Medical Association. 1957;130:199-204.
- Shahid M, Sabir N, Ahmed I, Khan RW, Irshad M, Rizwan M, Ahmed S. Diagnosis of subclinical mastitis in bovine using conventional methods and electronic detector. Journal of Agricultural and Biological Science. 2011;6(11):18-22.
- 20. Sharma A, Singh R, Beigh SA, Bhardwaj RK. Prevalence of subclinical mastitis in cross breed cattle from Jammu region. Veterinary Practitioner. 2012;13(2):356-357.
- 21. Sharma N, Pandey V, Sudhan NA. Comparison of some indirect screening tests for detection of subclinical mastitis in dairy cows. Bulgarian Journal of Veterinary Medicine, 13(2), 98-103.
- 22. Singh D. Diagnostic and therapeutic studies on subclinical mastitis in cows. M.V.Sc. Thesis submitted to Rajasthan University of Veterinary and Animal Science, Bikaner, Rajasthan, 2015.

- 23. Snedecor GW, Cochran WG. Statistical methods, 8<sup>th</sup>Edn. Ames: Iowa State University Press Iowa. 1989;54:71-82.
- 24. Sudhan NA, Singh R, Singh M, Soodan JS. Studies on prevalence, etiology and diagnosis of subclinical mastitis among crossbred cows. Indian Journal of Animal Research. 2005;39(2):127-130.
- 25. Sunder J, De AK, Jeyakumar S, Kundu A. Effect of feeding of Morinda citrifolia fruit juice on the biophysical parameters of healthy as well as mastitisaffected cow milk. Journal of Applied Animal Research. 2013;41(1):29-33.
- 26. Tanwar RK, SK Vyas, Fakhruddin, Singh AP. Comparative efficacy of various diagnostic tests in diagnosis of SCM in Rathi cows. In: Proceedings of round table conference of the Indian Association for the Advancement of Veterinary Research (IAAVR) on Mastitis, 2001, 161-163.