



ISSN (E): 2277-7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2022; SP-11(6): 2412-2416
© 2022 TPI
www.thepharmajournal.com
Received: 20-04-2022
Accepted: 24-05-2022

Sharad Kumar
Division of TVCC, FVSc &AH,
SKUAST-J, R. S. Pura, Jammu,
Jammu and Kashmir, India

Utsav Sharma
Division of VGO, FVSc &AH,
SKUAST-J, R. S. Pura, Jammu,
Jammu and Kashmir, India

Sudhir Kumar
Division of VGO, FVSc &AH,
SKUAST-J, R. S. Pura, Jammu,
Jammu and Kashmir, India

JS Soodan
Division of TVCC, FVSc &AH,
SKUAST-J, R. S. Pura, Jammu,
Jammu and Kashmir, India

Corresponding Author
Sharad Kumar
Division of TVCC, FVSc &AH,
SKUAST-J, R. S. Pura, Jammu,
Jammu and Kashmir, India

Estrus induction and fertility in response to exogenous hormonal administration protocols in postpartum anestrus murrah buffaloes

Sharad Kumar, Utsav Sharma, Sudhir Kumar and JS Soodan

Abstract

The objectives were to determine the estrus and conception responses to the controlled internal drug release (CIDR), Crestar and Polyurethane sponge (vaginal implant containing 1.5 gm of Natural Micronized Progesterone) along with prostaglandin F_{2α} and PMSG protocol and to compare the efficacy of this protocol. In Group I twelve buffaloes received CIDR inserts for 7 d and were treated with PGF_{2α} on day 6th and inj PMSG (600 IU) immediately after CIDR withdrawal. In Group II, CIDR was replaced with Crestar, While In Group III and IV, exogenous progesterone source were Polyurethane sponge (vaginal implant containing 1.5 gm of Natural Micronized Progesterone). In Group I and III, 100% of buffaloes showed estrus responses, while Group II and IV buffaloes showed 91.67% estrus response. The overall conception rate were 50.00, 63.64, 66.67 and 54.55% respectively. Intra-vaginal sponge impregnated with progesterone with conception rate of (66.67%) provided excellent fertility. The intra-vaginal sponge impregnated with progesterone can replace CIDR and Crestar in buffaloes and costs 1/10th the price, thus reducing the financial implications to livestock owner.

Keywords: CIDR, Crestar, polyurethane sponge estrus induction, ovulation, conception, progesterone and buffalo

1. Introduction

Post-partum anestrus is the period after calving during which dairy animals do not exhibit estrus. It is the prevalent, frustrating and challenging problem encountered in dairy animals. Field survey on reproductive disorder have revealed that anestrus is most common single cause of infertility in buffaloes among which inactive or non-functional ovaries are single cause of anestrus (EL-Wishy, 2007) [12]. Incidence of anestrus is more in buffaloes than cattle and problem is more severe in summers (Singh *et al.*, 2010) [41]. In comparison to cows, buffaloes have lesser number of pre antral and antral follicles and greater tendency of follicular atresia (Danell, 1987) [8]. Different methods are employed to reduce the postpartum anestrus and subsequent inter-calving period in order to increase the fertility during the low breeding season. Exogenous administration of progestins are considered appropriate for non-cyclic or anestrus post-partum cows (Yaniz *et al.*, 2004) [48] and buffaloes (Lakra *et al.*, 2003) [20], for resumption of estrus which normal cycle length (Rhodes *et al.*, 2003) [35]. Progesterone both exogenous and endogenous suppress the estrus and ovulation by inhibiting the release of Leutinizing Hormone (LH) (Peter and Lamming, 1984) [34] and its withdrawal results in a gradual rise in plasma LH concentration culminating in a preovulatory LH surge which occurs approximately after 48 hours (Garcia-Winder *et al.*, 1986) [13]. Crestar is a recently used therapy for estrus induction in postpartum anestrus buffaloes (Nayak *et al.*, 2009; Malik *et al.*; 2011; Nakrani *et al.*, 2014) [29, 21, 25]. The CIDR-B is the most common intravaginal progesterone available and contains 1.38 g of progesterone and is extensively used in various protocols for estrus synchronization, induction, fixed-time AI, fixed-time embryo transfer, and super stimulation programs. Polyurethane sponges impregnated with varying amounts of progesterone and of various lengths, diameters and densities have been used in cattle. In India, at present no locally made commercial progesterone-releasing intravaginal device is available for treatment of post-partum anestrus in buffaloes. Based on these views, the present study was proposed to evaluate the progesterone implants in post-partum anestrus buffaloes in relation to reproductive parameters.

2. Materials and Methods

The present study was carried out on 48 postpartum anestrus buffaloes from different villages of R. S. Pura area of Jammu district. History of each animal was recorded before including in the investigation. Buffaloes were selected on the basis of history, which had not exhibited any signs of estrus for 90 days or more postpartum. The anestrus period varied from 3 months to 6 months duration. Buffaloes were examined per rectally twice at 10 days interval to ascertain inactive, smooth ovaries, flaccid uterine horns and pale vestibule. Pathological causes of anestrus were ruled out. On the basis of selection criteria, 48 post-partum anestrus buffaloes were included in the trial and randomly divided into 4 groups consisting of 12 animals in each group. In Group I buffaloes progesterone were supplemented with CIDR (Controlled Intravaginal Drug Release, Pfizer Ltd.), In Group II progesterone was supplemented by Crestar ear implant, In Group III exogenous source of progesterone was Polyurethane sponge, a vaginal implant (containing 1.5 gm of Natural Micronized Progesterone) and In Group IV source of progesterone was Polyurethane sponge, a vaginal implant (containing 1.5 gm of

Natural Micronized Progesterone along with 1% carboxymethylcellulose). All post-partum buffaloes in each group were treated with an injection of 600 I.U. PMSG (Folligon) administered intramuscularly on the day of removal of source of exogenous progesterone. The buffaloes exhibiting estrus (behavioural symptoms) within 10 days after treatment were considered to have responded positively to treatment and were also examined per-rectally (Nautiyal, 2000) [28]. Artificial Insemination (AI) was done with good quality frozen thawed semen 12 h after the onset of estrus. The animals not conceiving at first estrus were again inseminated on subsequent estrus. Statistical Analysis was done by using analysis of variance (ANOVA) and General Linear Models of SPSS 16.0 computer programme. The significance was assayed at 5% ($P < 0.05$).

3. Results and Discussion

Response of post-partum anestrus buffaloes to different hormone protocols *viz* CIDR (Group I), Crestar (Group II), Intra-vaginal sponge (Group III) and Intra-vaginal sponge with CMC (Group IV) is summarized in table 1.

Table 1: Response to estrus induction treatment

| Sr. No. | Particulars | Group I (N=12) | Group II (N=12) | Group III (N=12) | Group IV (N=12) |
|---------|--------------------------------|----------------------------------|---------------------------------|---------------------------------|---------------------------------|
| 1 | Estrus Induction Response | 12 (100%) | 11 (91.67%) | 12 (100%) | 11 (91.67%) |
| 2 | Onset of estrus (hrs) | 37.07±1.58 ^{ab} (28-42) | 33.32±2.23 ^a (23-40) | 39.33±2.24 ^b (28-48) | 40.33±2.41 ^b (35-50) |
| 3 | Duration of estrus (hrs) | 22.31±0.79 ^a (18-27) | 21.08±0.69 ^a (18-26) | 27.08±0.69 ^b (24-31) | 23.64±0.50 ^c (20-29) |
| 4 | Conception rate | | | | |
| | First Service Conception Rate | 3/12 (25.00%) | 4/11 (36.36%) | 5/12 (41.67%) | 3/11 (27.27%) |
| | Second Service Conception Rate | 3/9 (33.33%) | 3/7 (42.86%) | 3/7 (42.86%) | 3/8 (37.50%) |
| | Overall Conception Rate | 6/12 (50.00%) | 7/11 (63.64%) | 8/12 (66.67%) | 6/11 (54.55%) |

Means bearing different superscript within a row differ significantly ($P < 0.05$).

In Group I, the estrus induction response of buffaloes treated with CIDR and PMSG was 100%. The present findings are in collaboration with observations of Caesar *et al.* (2011) [7] and Kajaysri *et al.* (2015) [18] in buffaloes. The average time required for onset of estrus in Group I was 37.07±1.58 hrs (28-42 hrs). Which are in the agreement with the findings of Kajaysri *et al.* (2015) [18] and Lakra *et al.* (2003) [20] who reported 36-72 hrs time required for inducing estrus in buffaloes after CIDR and PMSG treatment. In the present study, the average duration of estrus in Group I was 22.31±0.79 hrs (18-27 hrs). These finding are in collaboration with Kajaysri *et al.* (2015) [18] and Saini *et al.* (2017) [36]. In Group I, out of 12 buffaloes, 3 (25.00%) conceived at first estrus. 3 (33.33%) out of 9 conceived at second estrus. The overall pregnancy rate was 6/12 (50.00%). These findings are in agreement with Lakra *et al.* (2003) [20] and Naseer *et al.* (2013) [26] in buffaloes. High progesterone in the blood supply suppresses GnRH neurons in the tonic center of the hypothalamus with a flow-on to suppression of both LH and FSH from the pituitary. Upon CIDR removal the rapid drop in circulating concentrations of progesterone promotes the release of a surge of GnRH, followed by FSH and LH from the anterior pituitary causing the resumption of ovarian cyclicity (Zaabel *et al.*, 2009; Azawi *et al.*, 2012) [49, 4].

In Group II, the estrus induction response of buffaloes treated with Crestar and PMSG was 91.67%. Following removal of implant, the resumption of follicular development and maturation might be due to flux of the gonadotropins from the pituitary. The use of PMSG along with Norgestomet ear implant may be useful as PMSG is known to increase blood estrogen level and in turn leads to induction of behavioral

estrus signs (Singh *et al.*, 2004a) [40]. The present findings are in close collaboration with Nakrani *et al.* (2014) [25], Parmar *et al.* (2015) [31] and Jerome *et al.* (2016) [17]. The average time required for onset of estrus in Group II was 33.32±2.23 hrs (23-48 hrs). The time interval of estrus expression in this study corroborated with findings of Thangapandian *et al.* (2015) [45] and Dhalkari *et al.* (2018) [10]. The time interval to exhibit estrus after hormonal treatment observed in the present study might be due to the administration of PMSG in combination with progesterone and estrogen (Nath *et al.*, 2004) [27]. The average duration of estrus in Group II was 21.08±0.69 hrs (18-26 hrs), which collaborate with finding of Parmar *et al.* (2015) [31] and Thangapandian *et al.* (2015) [45]. In Group II, out of 11 buffaloes, 4 (36.37%) conceived at first estrus. 3 (42.86%) out of 7 conceived at second estrus. The overall pregnancy rate was 7/11 (63.64%). The findings in the present study are in collaboration with Vikas *et al.* (2014) [46], Parmar *et al.* (2015) [31] and Kumar *et al.* (2016) [19].

In Group III and IV, the estrus induction response of buffaloes treated with Intra-vaginal sponge and PMSG was 100 and 91.67% respectively. Progesterone sponge was found to be more effective in anestrus buffaloes by slow release of progesterone from the sponge and absorbed through blood vessel in vagina. It has a negative feedback effect on anterior pituitary gland to inhibit estrous cycle (Thammakarn, 2010) [44]. When the sponge is removed from vagina, the progesterone in blood stream was decreased immediately and the reproductive cycle of cattle becomes active again (Devipriya *et al.*, 2015) [9]. The present findings are in collaboration with Visha *et al.* (2014) [47] and Jayachandran and Muralidharan (2015) [16]. In the present study, the average

time required for onset of estrus in Group III and IV were 39.33 ± 2.24 hrs (28-48 hrs) and 40.33 ± 2.41 (35-50 hrs) respectively. These findings are in collaboration with Saini *et al.* (2017)^[36]. The average time required for onset of estrus in Group III and IV was significantly higher compared to Group II. This may be due to increased half-life and delayed metabolism of micronized progesterone (Malik and Krishnaprasad, 2016)^[22]. The average duration of estrus in Group III and IV were 27.08 ± 0.69 hrs (24-31 hrs) and 23.64 ± 0.50 (20-29 hrs) (Table 4.1) respectively. On the contrary, Saini *et al.* (2017)^[36] reported lower duration of estrus. Estrus duration in Group IV was significantly higher compared to Group III as CMC may have adhered to vaginal wall even after removal of sponge leading to not much abrupt fall in progesterone concentration. The mean duration of estrus was significantly higher ($P < 0.05$) in Group III and IV compared to Group I and II. The duration of estrus was higher in these group due to micronized natural progesterone having increased half-life (Malik and Krishnaprasad, 2016)^[22] and due to suprabasal progesterone, which causes prolonged estrus (Bage *et al.*, 2002)^[5]. In Group III, out of 12 buffaloes, 5 (41.67%) conceived at first estrus. 3 (42.87%) out of 7 conceived at second estrus. The overall pregnancy rate was 8/12 (66.67%) and in Group IV, out of 11 buffaloes, 3 (27.27%) conceived at first estrus. 3 (37.50%) out of 8 conceived at second estrus. The overall pregnancy rate was 6/11 (54.54%). These findings are in collaboration with Devipriya *et al.* (2015)^[9] and Jaychandran and Muralidharan (2015)^[16]. On the other hand lower pregnancy rates were reported by Singh *et al.* (2010b)^[42]. The pregnancy rate was higher (66.67%) in intravaginal sponge without CMC compared to intravaginal sponge with CMC (54.54%). In group IV (intravaginal sponge with CMC) has led to local infection and which might have resulted in inadequate / excess absorption of micronized natural progesterone in some of animal which in turn led to insufficient follicular development (Hixon *et al.*, 1981)^[14] or prolongation of follicular development attributed to low pregnancy rate (Stock and Stolla, 1995)^[43]. Further, 30-40 per cent conception obtained at second cycle in anestrus buffaloes induced to cycle proved that all the estrus induction protocols used to induce estrus and established normal cyclicity in treated animals, resulting into conceptions in subsequent cycles like normal cycling/breeding buffaloes. These observations further supported the previous observations on use of similar protocols in anestrus cows and buffaloes by many workers (Naikoo *et al.*, 2010; Bhoraniya *et al.*, 2012; Ammu *et al.*, 2012; Parmar 2013; Patel *et al.*, 2013; Savalia *et al.*, 2013; Dhami *et al.*, 2014)^[24, 6, 3, 30, 32, 38, 11].

Suboptimal functioning of the hypothalamus-pituitary-gonadal axis and low FSH and LH peaks have been related by several authors to anestrus in buffalo under tropical and subtropical conditions (Janakiraman *et al.*, 1980, Aboul-Elah *et al.*, 1983 and Saini *et al.*, 1988)^[15, 1, 37]. The high progesterone primes the brain for the actions of estrogen and addition of eCG subsequent to progesterone treatment increased plasma concentration of LH and FSH and enhanced the LH peak in non-cyclic buffalo heifers (Saini *et al.*, 1988)^[37] due to rapid decline in systemic progesterone concentration on its removal (Perry *et al.*, 2004)^[33] leading to induction of behavioral estrus signs (Singh *et al.*, 2004a)^[40] due to synergistic effect of exogenous gonadotropin on folliculogenesis, oocytes maturation and development of corpus luteum (Agarwal *et al.*, 2005)^[2]. Thus, administration

of eCG probably helps complete the recovery of hypothalamus-pituitary-gonadal axis function already stimulated by the progesterone treatment (Murugavel *et al.*, 2009)^[23].

4. Conclusion

Amongst the different hormonal protocol CIDR and Intra-vaginal sponge impregnated with progesterone treatment group resulted in 100% estrus induction response while Intra-vaginal sponge impregnated with progesterone with conception rate of (66.67%) provided excellent fertility. It can be concluded that intra-vaginal sponge impregnated with progesterone can replace CIDR and Crestar in buffaloes and costs 1/10th the price, thus reducing the financial implications to livestock owner.

5. References

1. Aboul-Ela MB, El-Keraby FE, Chesworth JM. Seasonal variation in the LH release in response to GnRH in the buffalo. *Animal Reproduction Science*. 1983;6:229-32.
2. Agarwal SK, Singh SK, Rajkumar R. Reproductive disorders and their management in cattle and buffalo: A review. *Indian Journal Animal Science*. 2005;75(7):858-873.
3. Ammu R, Dhami AJ, Naikoo M, Parmar BC, Divekar BS. Oestrus induction and fertility response in postpartum anoestrus Gir cows. *Indian Journal of Animal Reproduction*. 2012;33(1):37-42.
4. Azawi OI, Ali MD, Oday SA, Al-Hadad AS, Mouayad SJ, Hussien ASA. Treatment of anoestrus in Iraqi buffaloes using Ovsynch alone or in combination with CIDR. *Journal of Advance Veterinary Research*. 2012;2(1):68-72.
5. Bage R, Gustafson H, Larson B, Forsberg M, Radriguez-Martinez H. Repeat breeding in dairy heifers: Follicular dynamics and estrous cycle characteristics in relation to sexual hormone pattern. *Theriogenology*. 2002;57:2257-2269.
6. Bhoraniya HL, Dhami AJ, Naikoo M, Parmar BC, Sarvaiya NP. Effect of oestrus synchronization protocols on plasma progesterone profile and fertility in postpartum anoestrus Kankrej cows. *Tropical Animal Health Production*. 2012;44(6):1191-1197.
7. Caesar NK, Shukla SN, Agrawal S, Shrivastava OP, Agrawal S, Agrawal RG. Study on fertility response in anoestrus buffaloes using a modified CIDR- based synchronization protocol. *Buffalo bulletin*. 2011;30(3):184-187.
8. Danell B. Oestrus behaviour, ovarian morphology and cyclical variation in the follicular system and endocrine pattern in Water Buffalo heifers. PhD Dissertation, Swedish University of Agricultural Science, Uppsala, 1987.
9. Devipriya K, Eswari S, Nanjappan K. Induction of estrus and fertility response in true anoestrus buffaloes using intravaginal progesterone sponge during summer. *Journal of Animal Research*. 2015;5(2):389-391.
10. Dhalkari S, Sahatpure SK, Patil AD, Pati P, Gawande AP. Estrus induction response following hormone treatment in Murrah buffalo. *Indian Journal of Animal Reproduction*. 2018;39(1):61-62.
11. Dhami AJ, Panchal MT, Hadiya KK, Patel JA, Shah RG. Use of controlled breeding techniques under field conditions for estrus synchronization and conception in

- anoestrus crossbred cows and buffaloes. Proc. 2nd Annual Meeting of SVSBT and National Seminar on Biotechnological approaches to challenges in animal health & production, Vet. College, DUVASU, Mathura (UP), India, 2014 March 6-7, 86.
12. El-Wishy AB. The postpartum buffaloes II. Acyclicity and anestrus. *Animal Reproduction Science*. 2007;97:216-236.
 13. Garcia-Winder M, Lewis PE, Deaver DR, Smith VG, Lewis GS, In Skeep EK. Endocrine profiles associated with life span and induced corpora lutea in postpartum beef cows. *Journal of Animal Science*. 1986;62:1353-1362.
 14. Hixon DL, Kesler DJ, Troxel TR, Vincent DL, Wiseman BS. Reproductive hormone secretions and first service conception rate subsequent to ovulation control with Synchro-Mate B. *Theriogenology*. 1981;16:219-229.
 15. Janakiraman K, Desai MC, Amin DR, Sheth SR, Mudbidri SB, Wadedakar KB. Serum gonadotrophin levels in buffaloes in relation to phases of estrus and breeding periods. *Indian Journal of Animal Science*. 1980;50:606-606.
 16. Jayachandran S, Muralidharan J. Estrus induction and conception percentage in anestrus buffaloes treated with progesterone. *Shanlax International Journal of Veterinary Science*. 2015;3(1):12-19.
 17. Jerome A, Srivastava SK, Sharma RK. Study on follicular characteristics, hormonal and biochemical profile in norgestomet+PMSG treated acyclic buffaloes. *Indian Journal of Veterinary Research*. 2016;17(4):247-252.
 18. Kajaysri J, Chumchoung C, Photikanit G. Estrus and ovulation in anestrus postpartum swamp buffaloes following synchronization with a controlled internal drug release device and prostaglandin F₂ α based protocols. *Buffalo bulletin*. 2015;34(3):357-368.
 19. Kumar L, Phogat JB, Pandey AK, Phulia SK, Kumar S, Dalal J. Estrus induction and fertility response following different treatment protocols in Murrah buffaloes under field conditions. *Veterinary World*. 2016;9(12):1466-1470.
 20. Lakra BS, Luthra RA, Khar SK, Nanda T, Beniwal BS. Induction of cyclicity in anestrus buffaloes during non-breeding season. *Intas Polivet*. 2003;4:162-166.
 21. Malik RK, Singh P, Sharma RK, Phulia SK, Tuli RK. Efficacy of norgestomet ear implant for estrus induction on postpartum Murrah buffaloes (*Bubalus bubalis*). *Indian Journal of Animal Sciences*. 2011;81:7.
 22. Malik S, Krishnaprasad K. Natural micronized progesterone sustained release (SR) and luteal phase: Role redefined. *Journal of Clinical and Diagnostic Research*. 2016;10(2):4.
 23. Murugavel K, Antonie D, Raju MS, Lopez-Gatius F. The effect of addition of equine chorionic gonadotropin to a progesterone-based estrous synchronization protocol in buffaloes (*Bubalus bubalis*) under tropical conditions. *Theriogenology*. 2009;71:1120-1126.
 24. Naikoo M, Patel DM, Sarvaiya NP, Killedar A. Estrus synchronization in postpartum anoestrus Mehsana buffaloes using different hormone protocols. *Indian Journal of Field Veterinarians*. 2010;6(2):1-4.
 25. Nakrani BB, Panchal MT, Dhami AJ, Hadiya KK, Patel JA, Gosai RK, *et al*. Influence of controlled breeding techniques on estrus induction response, conception rate and plasma progesterone profile in anoestrus buffaloes. *Global Journal of Medical Research*. 2014;14(3):1-6.
 26. Naseer Zahid, Ejaz Ahmad, Nemat Ullah, Muhammad Yaqoob, Zeeshan Akbar. Treatment of anestrus Nili-Ravi buffaloes using eCG and CIDR protocols. *Asian Pacific Journal of Reproduction*. 2013;2(3):215-217.
 27. Nath HC, Dutta DJ, Dutta A, Biswas RK. Effect of Crestar administration on oestrus response in postpartum anoestrous cows. *Indian Journal of Animal Science*. 2004;74(1):22-24.
 28. Nautiyal H. Effect of synthetic progestagen implant with and without PGF₂ α / PMSG and GnRH for inducing ovarian cyclicity in cattle and buffaloes. PhD Thesis, IVRI, Izzatnagar, India, 2000.
 29. Nayak V, Agrawal RG, Srivastav OP, Thakur MS. Induction of estrus in true anoestrus buffaloes using Crestar implants alone and in combination with PMSG. *Buffalo Bulletin*. 2009;28:51-54.
 30. Parmar BN. Augmenting reproductive efficiency of infertile buffaloes using controlled breeding techniques in tribal areas. MVSc Thesis, Anand Agricultural University, Anand, Gujarat, India, 2013.
 31. Parmar SC, Khasatiya CT, Tyagi KK, Chaudharu JK, Patel RV. Estrus induction and serum progesterone and oestradiol-17 β profile in norgestomet primed postpartum anoestrus Surti buffaloes. *The Indian Journal of Veterinary Sciences and Biotechnology*. 2015;11(1):24-29.
 32. Patel KR, Dhami AJ, Hadiya KK, Savalia, Sarvaiya NP. Effect of CIDR and Ovsynch protocols on estrus response, fertility and plasma progesterone and biochemical profile in true anoestrus crossbred cows. *Indian Journal of Animal Production Management*. 2013;29(3-4):50-58.
 33. Perry GA, Smith MF, Geary T W. Ability of intravaginal progesterone inserts and melengestrol acetate to induce estrous cycles in postpartum beef cows. *Journal of Animal Science*. 2004;82:695-704.
 34. Peter AR, Lamming GE. Reproductive of the cow in the post-partum period, endocrine pattern and induction of ovulation. *British Veterinary Journal*. 2004;140:269-280.
 35. Rhodes FM, McDougall S, Burke CR, Verkerk GA, Macmillan KL. Treatment of cows with an extended postpartum anoestrus interval. *Journal of Dairy Science*. 2003;86:1876-1894.
 36. Saini A, Luthra RA, Pandey AK, Nanda T, Kumar L. Use of progesterone impregnated indigenous sponges along with PMSG for the induction of cyclicity in anoestrus Murrah buffalo. *Indian Journal of Animal Reproduction*. 2017;38(1):12-15.
 37. Saini MS, Galhotra MM, Kaker ML, Razdan MN. Induction of estrus and ovulation in non-cyclic buffalo (*Bubalus bubalis*) heifers with progesterone releasing intravaginal device and pregnant mare serum gonadotrophin and their gonadotrophin profile. *Theriogenology*. 1988;26:749-755.
 38. Savalia KK, Dhami AJ, Hadiya KK, Patel KR, Sarvaiya NP. Influence of Controlled Breeding Techniques on Fertility and Plasma Progesterone, Protein and Cholesterol Profile in True Anoestrus and Repeat Breeding Buffaloes. *Veterinary World*. 2014;7(9):727-732.
 39. Singh AS, Saxena MS, Prasad S. Efficacy of Crestar and its combination with Folligon on postpartum anoestrus in

- buffalo. *Indian Journal of Animal Reproduction*. 2004;25:43-44.
40. Singh AS, Saxena MS, Prasad S. Efficacy of Crestar and its combination with Folligon on postpartum anoestrus in buffalo. *Indian Journal of Animal Reproduction*. 2004a;25:43-44.
 41. Singh S, Kumar D, Naqvi SMK. Progesterone concentration in buffaloes after insertion of progesterone impregnated intra-vaginal sponges. *Veterinary Practitioner*. 2010;11(1):17-18.
 42. Singh S, Kumar D, Naqvi SMK. Progesterone concentration in buffaloes after insertion of progesterone impregnated intra-vaginal sponges. *Veterinary Practitioner*. 2010b;11(1):17-18.
 43. Stock AE, Stolla R. Der dominante Ovarfollikel beim Rind-Physiologische Zusammenhänge und praktische Bedeutung. *Tierarztl Umschau*. 1995;50:543-550.
 44. Thamakkarn C. Case report: large litter size in first parity does synchronize the estrus by intravaginal medroxyprogesterone sponge and PMSG injection. In *Proceedings, 16th Asian Agricultural Symposium and 1st International Symposium on Agricultural Technology*. KMITL, Bangkok, Thailand, 2010, 577-580.
 45. Thangapandian M, Pothiappan P, Palaniappan RM, Samuel EJ, Kathiresan D. Induction of estrus in anestrus Murrah buffaloes and programmed breeding. *Buffalo Bulletin*. 2015;34(2):241-244.
 46. Vikash MV, Malik RK, Pardeep Singh. Impact of CIDR in combination with different hormones for treatment of anestrus in buffaloes under field conditions in Haryana. *Haryana Veterinary*. 2014;53(1):28-33.
 47. Visha P, Jayachandran S, Selvaraj P, Nanjappan K, Sathes KS, Palanisammi A. Follicular growth pattern in buffaloes synchronized to estrus with progesterone impregnated intravaginal sponges. *International Journal of Science, Environment and Technology*. 2014;3(3):960-965.
 48. Yaniz JL, Murugavel K, Lopez-Gatius F. Recent developments in oestrous synchronization of postpartum dairy cows with and without ovarian disorders. *Reproduction of Domestic Animal*. 2004;39:86-93.
 49. Zaabel SM, Hegab AO, Montasser AE, El-Sheikh H. Reproductive performance of anestrus buffaloes treated with CIDR. *Animal Reproduction*. 2009;6(3):460-464.