



ISSN (E): 2277-7695

ISSN (P): 2349-8242

NAAS Rating: 5.23

TPI 2022; 11(6): 52-56

© 2022 TPI

[www.thepharmajournal.com](http://www.thepharmajournal.com)

Received: 06-02-2022

Accepted: 13-05-2022

**Amit Prakash Nayak**

Ph.D. Research Scholar,  
Department of Forestry, College  
of Agriculture, Indira Gandhi  
Agricultural University, Raipur,  
Chhattisgarh, India

**Dr. Rajendra Kumar Prajapati**

Professor Forestry, Department  
of Forestry, College of  
Agriculture, Indira Gandhi  
Agricultural University, Raipur,  
Chhattisgarh, India

**Dr. MD Omprakash**

Assistant Professor, Faculty of  
Ecosystem and Environment  
Management, Indian Institute of  
Forest Management, Bhopal,  
Madhya Pradesh, India

**Dr. Manish Mishra**

Research Associate, Faculty of  
Ecosystem and Environment  
Management, Indian Institute of  
Forest Management, Bhopal,  
Madhya Pradesh, India

**Dr. Dharmendra Khokhar**

Scientist, Department of Plant  
Physiology, Agri. Bio-chemistry  
and Medicinal Aromatic Plants  
Agricultural, College of  
Agriculture, Indira Gandhi  
Agricultural University, Raipur,  
Chhattisgarh, India

**Chandrashikha Patel**

Ph.D. Research Scholar,  
Department of Forestry, College  
of Agriculture, Indira Gandhi  
Agricultural University, Raipur,  
Chhattisgarh, India

**Corresponding Author**

**Amit Prakash Nayak**

Ph.D. Research Scholar,  
Department of Forestry, College  
of Agriculture, Indira Gandhi  
Agricultural University, Raipur,  
Chhattisgarh, India

## Analysis of various physical properties of oleoresin obtained from *Pinus patula* in Amarkantak, Madhya Pradesh, India

**Amit Prakash Nayak, Dr. RK Prajapati, Dr. MD Omprakash, Dr. Manish Mishra, Dr. Dharmendra Khokhar and Chandrashikha Patel**

### Abstract

Oleoresin are among the most important NTFPs in the World. They are one of the oldest tree products used by humans. The *Pinus patula* tree planted globally as a dominant plantation forestry crop across various parts of the world especially in the high altitude tropical environment of India. Analysis of various physical properties were carried out in the laboratory conditions. The findings revealed that the oleoresin samples exhibited a number of properties that could be useful to the chemical and pharmaceutical sectors, but no commercial oleoresin tapping experiments have indeed been conducted in this species to far. The resin samples were found to be highly soluble in Chloroform and having high rosin content of (91.48%).

**Keywords:** Oleoresin, *Pinus patula*, Tropical pines, Turpentine oil, Viscosity

### 1. Introduction

Resin is mostly a mixture of volatile monoterpenes and non-volatile diterpenic resin acids. Through a distillation process, resin can be separated into turpentine (30%) and rosin (70%) (Hawley and Palmer 1912) [7]. *Pinus patula* Schlecht. & Cham commonly known as Mexican weeping pine or Jelecote pine native to Mexico (from Tamaulipas to Oaxaca region). This species has been widely planted for plantation forestry program on a large scale in Africa, Columbia, Brazil, Argentina, India, Nepal, New Zealand (Farjon, 2017) [6]. It grows fast and produces a long, straight bole, for most part free of branches and the wood obtained from this tree is soft, light colored and easily worked, therefore it finds application in flooring and paneling, plywood and other wood based industries. In India first trial plantation of *Pinus patula* was raised in West Bengal in 1929 at Ramam which was located at an elevation of 2270 m, Whereas in the present study site it was planted at Different compartments of Amarkantak range during 1970 at an elevation of 1080 m amsl. This oleoresins obtained from *Pinus* tree act as one of the most important non-wood forest products As per records, more than 80 species of pine trees are tapped for the production of resin throughout the world (Hillis, 1987). The purpose of this study was to determine physical properties of *Pinus patula* oleoresin samples. This properties affects the processing and makes this nature of studies necessary so as to determine the predictability of the end product and consequently quality of the output.

### 2. Materials and Methods

Trees selected for oleoresin tapping were located in Amarkantak Forest Range, Madhya Pradesh at an elevation of 1060 m amsl and lies at 22° 41' N latitude and 81° 43' longitude. These areas were selected for provenance trial of many tropical pine plantations in early 1970s under a Co-ordinate research program of Forest Research Institute. For oleoresin tapping, three distinct methods were used to collect samples, that were then examined in the laboratory for various parameter estimations.

#### 2.1 Determination of Turpentine and Rosin content

Twenty-five grams of oleoresin sample from each tree was taken in a round bottom flask and sufficient amount of water was added to it. The flask was fixed with Clevenger's apparatus and was heated with frequent agitation. Heating was continued for about one and half hour and after cooling for at least five minutes; the volume of turpentine in the graduated portion of the tube was noted. The distillation was continued until successive readings of the volume of turpentine did not differ (Persad, 1983) [13].

### Calculation of per cent Rosin & turpentine content

$$\text{Turpentine (\%)} = \frac{X}{\text{Sample weight (25gm)}} \times 100$$

Where: x is the volume of turpentine in ml.

#### 2.2 Rosin content

The non-volatile solid material called rosin which remained in the flask after distillation was allowed to cool until it was solidified. The rosin was weighed and the percentage of rosin was calculated as under:

$$\text{Rosin(\%)} = \frac{X}{\text{Sample weight(25gm)}} \times 100$$

Where: X is the weight of rosin in gram.

#### 2.3 Determination of solubility

Solubility is defined as the amount of substance that passes into solution to achieve a saturated solution at constant temperature and pressure. Solubility is expressed in terms of maximum volume or mass of the solute that dissolve in a given volume or mass of a solvent. The solubility of the resin was determined in, acetone, chloroform and benzene. 1.0 g sample of the resin was added to 50 ml of each of the above-mentioned solvents and left overnight. 25 ml of the clear supernatants were taken in small pre-weighted evaporating dishes and heated to dryness over a digital thermostatic water bath. The weights of the residue with reference to the volume of the solutions were determined using a digital top loading balance (Model.XP-3000) and expressed as the percentage solubility of the resins in the solvents (Mohsenin,1978).

$$\text{Water solubility index (\%)} = \frac{\text{Weight of dissolve resin sample in solution(g)}}{\text{Weight of resin sample(g)}}$$

#### 2.4 Determination of Moisture content

Moisture content of resin samples was determined by drying 5g of the resin sample to constant weight at 80 °C using hot air oven. Dried samples were cooled in desiccators before weighing. Moisture content was expressed as percentage of mass loss from the original mass as described by Yusuf (2011) [18].

$$\frac{\text{Weight of total sample} - \text{Weight of dry sample}}{\text{Weight of total sample}} \times 100$$

#### 2.5 Determination of Ash content

Ash content of the resin samples was determined by burning 1g of resin sample in a muffle furnace at 550 °C for 4 hours. The ash content was expressed as % ratio of the weight of ash to weight of the sample.

Ash value is designed to measure the total amount of material remaining after ignition. It includes both physiological (derived from plant tissue itself) and non-physiological (residue of the extraneous matter like sand that are adhering to plant substances. It also used to determine the critical levels of foreign matter, acid insoluble matter, salts of calcium, potassium and magnesium (Mocak *et al.* 1998).

$$\text{Ashcontent(\%)} = \frac{W3 - W1}{W2 - W1} \times 100$$

Where

W1 = weight of the empty crucible,

W2 = weight of crucible + sample and

W3 =weight of crucible + ash

#### 2.6 Determination of Viscosity

The viscosity of a liquid is its resistance to shearing, to stirring in capillary tube. Viscosity was considered as one of the most important analytical and commercial parameters, since it is a factor involving the size and the shape of the macro-molecule. The viscosity was determined and calculated for the 4%, concentration of all the resin sample at room temperature and rotational speed of 100 rpm by using spindle 62 and Spindle 63 of digital Brookfield DV-E viscometer (Sinha, 2017) [16].

#### 2.7 Determination of Colour and Odour

The collected resin sample was visually analysed for colour identification and aroma authentication through smells.

### 3. Results and Discussions

#### 3.1 Turpentine and Rosin content

Both components were separated from the resin using steam distillation using Clevengers apparatus. The two major products obtained from pine tree resin are turpentine oil and rosin. In the present experiment it was found that turpentine oil which is volatile in nature was found to be 8.52% and rosin content was 91.48% as shown in figure 2. Rosin or colophony is the solid fraction and has numerous uses, such as paints, varnishes, coatings, adhesives and other applications related to polymers (Maiti *et al.* 1989; Arrieta *et al.* 2017; Narayanan *et al.* 2017) [10, 2, 12]. Whereas the turpentine oil finds a wide variety of industrial uses such as in perfume industry, disinfectants and synthetic pine oil etc. The colour of rosin samples were found to be light yellow in colour. Coppen and Hone (1995) [5] reported that Rosin is usually translucent with a color ranging from light yellow to brown. Among different diameter class selected for resin tapping diameter classes of 35-40 cm yielded better resin yield. Teshome (2011) [17], observed that diameter class of 25-30 cm produced highest turpentine oil content.

#### 3.2 Solubility

The resin samples of *Pinus patula* was found to be highly soluble in Chloroform (1.28%), followed by Acetone and Benzene shown in figure 1. Schafer and Zandbiglari (2002) [15] revealed that Epoxy resin was highly soluble in Chloroform than Eucalyptus oil. The current finding also found satisfactory as compared to the report made by above workers.

#### 3.3 Moisture content

The moisture content of resin samples was determined using hot air oven. It was observed that the moisture content of resin sample was 1.1%. Jalali *et al.* (2011) [9] also reported that the moisture content in oleo gum resin of *Ferula gummosa* was 6.7%. Whereas moisture content in resin samples of *Pinus roxburghii* collected from Kashmir Valley was 3% (Ahmad *et al.* 2011) [1]. These figures were far higher than those observed in the current experiment.

#### 3.4 Ash content

The ash content in the resin was determined by muffle furnace. The values of Ash content in Rosin samples and Resin samples were found to be 21% & 1% respectively. The higher value of ash in rosin samples was because of due to the

volatile component turpentine was separated by distillation method. Whereas in resin samples turpentine oil was there. Saeidy *et al.* (2018) <sup>[14]</sup> reported in the oleo gum resin of Asafoetida (*Ferula assafoetida*) having ash content of 2.75%.The resin of *Pinus patula* had a 1% ash content in the current experiment; this difference could be attributed to the chemical characteristics of the two species being different.

### 3.5 Viscosity

The viscosity of different resin samples was calculated by using Brookfield viscometer at a rotation of 100 rpm with the use of spindle (No. 62 and 63) and at room temperature (approximately 32°C).From the laboratory analysis it was found that the viscosity value by using 62 and 63 number needles are 0.90 cp and 1.20 cp (Centipoise) respectively. Bourdeau and Schopmeyer 1958 reported that rate of oleoresin flow seems to be determined by Viscosity, number and size of resin ducts.

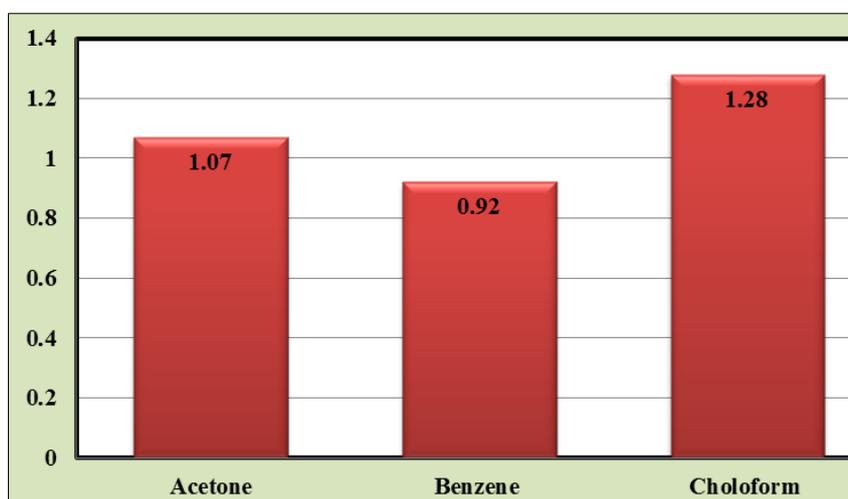
### 3.6 Colour and odour

*Pinus patula* having strong camphor like aroma with creamy white and yellowish white color respectively. Cabaret *et al.*

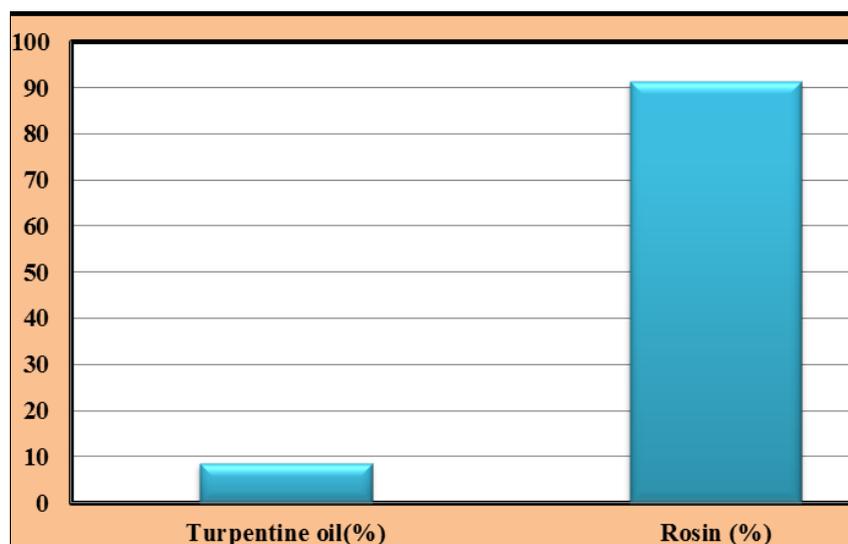
(2019) <sup>[4]</sup> conducted Spectro-colorimetry study in maritime pine (*Pinus pinsater*) and revealed that two different colour observed in this species. In the present experiment similar colour variation was observed. This colour variation illustrated a possible difference in the chemical composition.

**Table 1:** Physical properties of *Pinus patula* resin samples

| Physical properties         | Value                            |
|-----------------------------|----------------------------------|
| 1. Turpentine oil (%)       | 8.52                             |
| 2. Rosin (%)                | 91.48                            |
| 3. Solubility (%)           |                                  |
| a. Acetone                  | 1.07%                            |
| b. Benzene                  | 0.92%                            |
| c. Chloroform               | 1.28%                            |
| 4. Moisture content (%)     | 1.1                              |
| 5. Ash content in resin (%) | 1                                |
| 6. Ash content in rosin (%) | 21                               |
| 7. Viscosity                |                                  |
| a. 62 number needle         | 0.90 cp                          |
| b. 63 number needle         | 1.20 cp                          |
| 8. Colour                   | Yellowish white and Creamy white |
| 9. Aroma                    | Strong camphor                   |



**Fig 1:** Solubility of resin samples in different organic solvent



**Fig 2:** Rosin and Turpentine oil content in *Pinus patula* resin samples



Fig 3: Preparation for solubility test

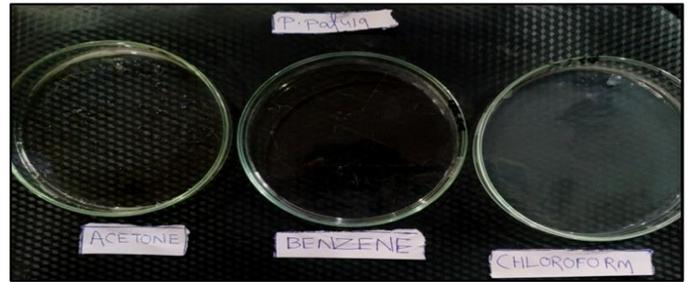


Fig 8: Solubility test in different organic solvent (Acetone, Benzene, Chloroform)



Fig 4: Turpentine oils after distillation Exudation of oleoresin from *Pinus patula* tree



Fig 5: Estimation of Viscosity of resin Samples by using Brookfield DV-E Viscometer



Fig 6: Estimation of Ash content of resin samples in Muffle furnace



Fig 7: Separation of Rosin and Turpentine oil by Clevenger apparatus (L), Ash content in Rosin samples

#### 4. Conclusions

Physical properties of the pine oleoresin are vary from species to species this is because of due to genetic parameters. This species is categorized as Least Concern by the IUCN Red List, and its global population is robust. As a result, a sustainable method of resin tapping should be developed so that additional bio-physical investigations in this species can be conducted on a global scale.

#### 5. Acknowledgment

The authors would like to thank Madhya Pradesh Forest Department for the permission to collect the resin samples and also thankful to Er. Prashant S. Pisalkar (Department of Agricultural Processing and Food Engineering, IGKV, Raipur) for providing the necessary facilities for preparing the part of Research.

#### 6. References

1. Ahmad Mir I, Jahan N. Standardization and Characterization of Oleo Resin of Pine obtained from *Pinus roxburghii* Sarg. *American Journal of Pharmatech Research*, 2017;7(6):211-227.
2. Arrieta MP, Samper MD, Jiménez-López M, Aldas M, López J. Combined effect of linseed oil and gum rosin as natural additives for PVC. *Ind. Crops Prod.* 2017;99:196–204.
3. Bourdeau PF, Schopmeyer. Oleoresin exudation pressure in Slash pine: its measurement, heritability, and relation to oleoresin yield. In the physiology of forest trees. Edited by K. Thaiman. Ronald press Co., New York. 1958, 313-319.
4. Cabaret T, Harfouche N, Leroyer Leo, Ledeuil JB, Martinez H, Charrier B. A study of the physico-chemical properties of dried maritime pine resin to better understand the exudation process. *Holzforschung*. 2019, 73(12)
5. Coppen JJW, Hone GA. Gum naval stores: turpentine and rosin from pine resin. *Non-wood forest products*. 1995;2:71.
6. Farjon A. *A Handbook of the Worlds Conifers*. Koninklijke Brill NV Publishers, Leiden, The Netherlands. 2017;981069. ISBN 9789004324497.
7. Hawley LF, Palmer RC. Distillation of resinous wood by saturated steam. *J. Ind. Eng. Chem.* 1912;4:789-795
8. Hillis W. The future of forest chemicals. *Chinese Academy of Forestry, Chemistry and Industry of Forest Products*. 1987;7(1):1-13.
9. Jalali HT, Ebrahimian ZJ, Evtuguin DV, Pascoal Neto C. Chemical composition of oleo-gum-resin from *Ferula gummosa*. *Industrial Crops and Products*. 2011;33(2):549-553.
10. Maiti S, Ray SS, Kundu AK. Rosin: a renewable resource

- for polymers and polymer chemicals. Prog. Polym. Sci. 1989;14:297-338
11. Mocak J, Jurasek P, Phillips GO, Varga S, Casadei E, Chikemai BN. The classification of natural gums. X. Chemometric characterization of exudate gums that conform to the revised specification of the gum Arabic for food use, and the identification of adulterants. Food Hydrocoll. 1998;12:141–150.
  12. Narayanan M, Loganathan S, Valapa RB, Thomas S, Varghese TO. UV protective poly(lactic acid) rosin films for sustainable packaging. Int. J. Biol. Macromol. 2017;99:37–45.
  13. Persad V. Rill method of resin tapping and storage obtained from chirpine (*Pinus roxburghii* Sargent). M.Sc. Thesis, HPKV, Palampur. 1983, 68p.
  14. Saeidy S, Nasirpour A, Keramat J, Desbrières J, Cerf DL, Pierre G. Structural characterization and thermal behavior of a gum extracted from *Ferula assafoetida* L. Carbohydr Polym. 2018;181:426-432. doi: 10.1016/j.carbpol.2017.10.096
  15. Schäfer E, Zandbiglari T. A comparison of the effectiveness of chloroform and eucalyptus oil in dissolving root canal sealers. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2002;93(5):611-6. doi: 10.1067/moe.2002.121899. PMID: 12075213.
  16. Sinha G. Studies on Drying Characteristics of Babul Gum (*Acacia nilotica*). M.Sc (Agril. Engg.) Thesis. Faculty of Agricultural Engineering, Indira Gandhi Krishi Vishwavidyalaya, Raipur (C.G.). 2017.
  17. Teshome T. Analysis of resin and turpentine oil constituents of *Pinus patula* grown in Ethiopia. Agriculture and Forestry issue. 2011;3(1):38-48.
  18. Yusuf AK. Studies on Some Physicochemical Properties of the Plant Gum Exudates of *Acacia senegal* (Dhakwara), *Acacia sieberiana* (Farar Kaya), and *Acacia nilotica* (Bagaruwa). 2011, 9(2).