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Department of Entomology, School of Agriculture, Lovely Professional University, Phagwara, Punjab, India Benefits of introducing amphibians and fishes in paddy ecosystem, harmful effects of neonicotinoids on amphibians and fishes

P Sai Prasad

Abstract

Amphibians and their ancestor fishes have flourished eons of years ago; their presence in paddy ecosystem is lot more beneficial as frogs and fishes help to promote exponential yield. This is achieved by their ability to control insects, pests, weeds and chironomid larvae, mosquitos. Apart from these their presence facilitates rise in soil permeability, aeration, soil nutrient uptake. The need for using fertilisers which are source of Greenhouse gases (GHG) are also minimized. Thus addition of aquatic organisms like fishes, amphibians promotes more sustainable, eco-friendly farming. However these animals are threatened by variety of chemicals such as Neonicotinoids. These pesticides increase the animal's susceptibility for pathogens, render them open to predators. They also cause morphological, anatomical, histological changes thus contributing to global amphibian decline. Current review focuses on the ecological services rendered by fishes and frogs and harmful effects of Neonicotinoids on these aquatic animals.

Keywords: Amphibians, fishes, paddy ecosystem, neonicotinoids

1. Introduction

With extensive use of pesticides round the globe residues are becoming a major part in all water bodies' viz. rivers, oceans and in urban areas. (Guruge & Tanabe, 2001; Sanchez- Bayo, 2012) ^[31]. Although the insecticides do control variety of insect pest, the repercussions are undoubtedly faced by the society. The effects are manifested in form of toxicity on non-target animals thus affecting them in various degrees (Sanchez-Bayo, 2012; Aliko & Baba, 2011)^[1]. In general the toxicity arises when the insecticides do show their effect on physiological functioning of animal (Sanchez-Bayo, 2012). Different insecticides have got different mode of action. Some are neurotoxic some are respiratory inhibitors thus impeding the process of oxidative phosphorylation in mitochondria (a mechanism found in all organisms). Some act as growth regulators thus barricading the arthropods from commencing metamorphosis. But when it comes to aquatic organisms, the tables are turned. Aquatic organisms mostly invertebrates are ancestral. Despite sharing common respiratory and digestive systems with hat of land dwelling insects, the former have poor detoxification systems (Sanchez-Bayo, 2012; Walker, 2011) ^[71]. Land dwelling insects have highly efficient iso-enzymes that facilitate efficient removal of toxins from the body. (Sanchez-Bayo, 2012). One such example is the use of propargite, a miticide is highly toxic for aquatic life as compared to terrestrial arthropods due to later developing highly advanced detoxifying systems. Speaking of aquatic systems, rice is most widely known man made agriculture system. Water that runs from oceans, lakes, rains, along with aerial drift of spraying contaminates all the forms of life. Many articles have been proposed that state the presence of pesticides in the water bodies. (Goulson, 2013; Tapparo et al., 2012; Marzaro et al., 2011; Krupke et al., 2012) ^[30, 65, 46, 39]. The sad reality is that still now these insecticides are being applied in tropics illegally. The most targeted organisms that succumb to these chemicals directly or indirectly (Ayoola, 2008 and Framnklin et al., 2010) are fishes. In the fishes gills which are the major source of respiration and osmoregulation. They also aid in assessing the quality of water (Fanta, 2003) ^[22]. Hence by histopathological analysis damage to cells can be screened at the targeted organs. Apart from fishes amphibians are other organisms that atone for indiscriminate use of pesticides. It is imperative to know that since 1970 most of amphibians are subjected to population decline. Although many reasons attribute to their decline such as climate changes (Pounds et al., 2001) ^[54] pathogenic microbes (Carey, 2000; Johnson et al., 1999) ^[9, 36] and losses of habitat

Corresponding Author P Sai Prasad Department of Entomology, School of Agriculture, Lovely Professional University, Phagwara, Punjab, India (Blaustein et al., 1994; Dunson et al., 1992) [6, 18] pesticides are yet another reason for their decline. Although more studies need to be documented, however many studies on other animals depict that most of the agriculture pesticides can cause significant changes in morphology or functional immune system of amphibians (Leubke et al., 1997; O'Halloran et al., 1996; Zelikoff et al., 1996) [45, 53, 74]. When contaminants affect the animals, behaviour is altered by immunomodulation thus defensive functions are supressed against variety of pathogens. (Fournier, 1998; Krzystynaik et al., 1985) [25, 40]. To understand their impact knowledge of these animals in rice ecosystem needs to be fully understood. Hence the review focuses on the importance of introducing frogs and fishes into rice ecosystem and use of chemical pesticides as detrimental factor determining their survival (Fulton and Chambers, 1985; Berril et al., 1994; Sparling et al., 2001) [28, 4, 64].

2. Importance of Paddy in India

Being consumer by over half of world's current population, paddy is then principle staple cereal crop (Xia et al., 2016, p.13, 4569-4579; Zhou et al., 2017, p. 115, 21-34) [72, 76]. India has been exporting 44, 14,562 Metric Tons of Basmati rice in the year of 2018-2019 whose worth can be accounted to 32.8 crores of Rupees (www. apeda.gov.in). This proves that India has enough rice area making itself reliant as well has symbolizing its strength in agrarian economy. Talking in terms of area India stands first and in terms of production it is below china making it second largest (Saha, 2006, p. 51(4):304-06). Dey *et al.* (2020) ^[57, 17] reported the increasing trend of area, production and productivity of paddy with its growth rate of 0.5 percent, 2.4 percent and 1.9 percent. Hence being one of most intensively crop grown especially in India, paddy is also subjected to large number of pest attacks. As per DWR. Vision 2050 Losses by pest, weed, insect, diseases and other pests did contribute to about 33%, 12.5%, 9.5%, 6.5% and 4.5% respectively. Mondal et al. (2017) [49] had conducted experiments estimating yield loss assessment of rice due to different biotic stress under SRI system and as per their estimates of crop losses, weeds occupied highest losses followed by insects later by in diseases. Hence forth taking all these above foresaid data, we conclude that management practices are therefore essential in increasing the productivity of paddy in India. However increase in use of pesticides and fertilizers are paving way for bio- accumulation, biomagnifications. One of classical example is Handigodu syndrome caused by non-judicious usage of arsenicals in Karnataka. This diseases affects joints, deformities in joints, hips etc. Besides putting human beings life at risk it also causes threat to biodiversity as in case of kasargod disaster where aerial spraying of endosulfan has vanquished most of aquatic, amphibian and caused many herbivores to become deformed. Apart from all the risks the pesticides and fertilizers pose to the environment, paddy fields are again important source of greenhouse gas emissions. Carlson et al. (2017) ^[10] has estimated that by releasing nitrous oxide and methane, paddy fields do account for 48% of global scale of greenhouse gas emissions. To tackle all these problems demand for use of eco- friendly management practices is called for. Research all over the world over use of frogs, fish

and shrimp has being done and benefits have been evaluated as well. This practice results in excess production of both fishes and frogs and also it is assumed that both weeds and pests are controlled by frogs (Teng *et al.*, 2015)^[66].

3. Effects of introducing fishes and frogs in paddy ecosystem

The practice of rice fish farming dates back to 1200 in china. While some resources say that it was practised 1700 years back (Li K, 1992; Cai *et al.*, 1995) ^[8]. However rice frog fish system has been a recent practise in south part of Asia. Rice fish farming system is a practise that is believed to augment the yields by ecological sustainable agriculture. (Jintong, 1996) ^[35]. Fishes and frogs serve as additional benefits by serving as a predator of insects, weeds. In addition to it they nourish the soil by their excrement. (Shugen *et al.*, 1995: Lightfoot, 1992; Frei and Becker; 2005) ^[60, 26]. Thus making a more profitable and sustainable countries like Africa, Zimbabwe are gaining momentum quickly thus raising the standards of living farming system. Detailed benefits of introducing frogs and fishes are discussed below.

3.1. Effect on CH₄ emissions

Rice fields are the major contributors to the global CH4 and N₂O emissions (Carlson et al., 2017)^[10]. Such drastic increase in the climate changes can have detrimental impact on the humanity (Mora et al., 2017) [50]. Main contributor to the GHG (greenhouse gasses emissions) is the addition of soil fertilizers (Liang et al., 2013) [41]. In rice fields, IRFF dramatically enhanced Do (Dissolved oxygen), soil Eh, TOC (Total organic) content, and soil C:N (carbon nitrogen) ratio (Fang et al., 2019)^[21]. Yuan et al. (2018) Xu et al. (2017) and Zhan et al. (2009) ^[75] investigated the GWP of GHG from an integrated rice-duck farming system. While the introduction of ducks in paddy fields can raise N2O emissions from duck faeces, it also increases the concentration of DO in the water layer and reduces CH4 emissions, according to their findings. Overall, the integrated rice-duck farming system reduces GWP in rice fields, according to their research. Frei et al. (2004), Datta et al. (2009), and Bhattacharyya et al. (2013)^{[26,} ^{14, 5]} on the other hand, found that carp generation in paddy fields boosts CH4 diffusion and discharge through the river.

3.2. Effect on insect pests

Liu *et al.* (2013) ^[43] carried out an experiment of raising bullfrogs in paddy fields. The results showed that the application of 900 and 1500 bullfrogs per hectare decreased the plant hopper population by 60% to 70% in paddy fields. Bull frogs specifically target insect pests such as *Spodoptera*, *Hieroglyphus*, *Melantis* and especially paddy stem borer thus increasing the yield considerably (Kharat, 1985) ^[37]. Fishes especially carps feed on the chironomid larvae (responsible for stealing soil nutrients) (Ikiwama & Otsuki, 1991) ^[34]. Plant hoppers which were responsible for rice sheath blight was controlled by using exotic frogs (Teng, 2015) ^[66]. Thereby augmenting the yield by controlling the actual fungi. Various researcher who have documented the various Genera of frogs and fishes and their prey insects listed order wise are briefly entailed in summarised way in Table.1.

Aquatic organism fish /frog	Insects preyed	Documented by
Unnamed frogs (exotic) reared from frog breeding farm in Zhejiang province	Cnaphalocrocis medinalis (rice leaf folder)	(Teng <i>et al.</i> , 2015) [66]
(Channa spp.) murrels	Mosquito larvae	(Usha 2018)
Aquatic organism fish /frog	Insects preyed	Documented by
Crucian Carp (Carassius spp.)	Insects belonging to Hemiptera, Ephemeroptera, Aphididae, Lycaneidae, Diptera, Tipulidae, chironomidae, Ichneumonidae, Formacidae and Braconidae of Hymenoptera	(Tsurata <i>et al.</i> , 2010)
<i>Rana tigrina</i> (Bull frog)	Spodoptera mauritia (Boisd.), Pelopidas mathias (F.) (Parnera mathias), Rhinyptia and Holotrichia spp., Melanitis leda ismene (Cram.) (M. ismene), Hieroglyphus banian (F.) and Scirpophaga incertulas (Wlk.) (Tryporyza incertulas).	(Kharat, 1985) ^[37]

Table 1: Different species of fishes and frogs controlling various insect orders

4. Neonicotinoids

Neonicotinoids represents chemical class with systemic nature and offering long time crop protection because of their extended half-life in soil (7-6931 days, Goulson, 2013) ^[30] aquatic systems 30-120 days, Lewis et al. (2016). They contribute to 25% of insecticide sales throughout the world (Jeschke et al., 2011; Sparks 2013) ^[19, 44]. Their traces have been reported across wider geographic range throughout the world on account of their worldwide usage and persistency (Sanchez-Bayo et al., 2016). Since last decade there has been exponential increase in usage of neonicotinoids against wide range of pests. Neonicotinoids on account for their high solubility as in imidacloprid (EPA 2003) ^[20] have higher potential for exposure for amphibians. Among all aquatic life particularly invertebrates suffer consequences despite having same, yet deficient detoxification, neurological, respiratory systems as that of terrestrial insects (sanchez-bayo.2012). Neonicotinoids along with widely used fipronil are agonist to acetylcholine for nicotinic receptors. As fipronil binds to gamma amino butryic acid receptors, neurons fire continuously thus exhausts cell energy and results in death of insect (Simon-delso et al., 2015; Velisak & Stara, 2018)^[62,]. Detrimental effects in terrestrial ecosystems are targeted on bees impairing their ability to discriminate floral scents (mustard et al., 2020)^[52], affecting navigation (Tison et al., 2016) ^[68] etc. Vertebrates and invertebrates although show differences in sub units for nicotinic receptors, former has low receptors with high affinity as opposed to latter (Simon-delso et al., 2015) [62]. In aquatic environments, Neonicotinoids concentrations frequently surpass standards for protection against short-term acute effects (0.2 g/L). Chronic long-term effects (0.035 g/L) on aquatic invertebrate populations (Morrissey et al., 2015). Although vertebrates are assumed to be immune to neonicotinoids, a growing body of research has found that exposed fishes had lower activity (Crosby et al. 2015; Finnegan et al. 2017) and growth (Hayasaka et al., 2012; DeCant &Barrett, 2010) ^[13, 24, 32, 16]. Neonicotinoids are most likely introduced to amphibians through their very permeable skin (Van Meter et al., 2014) [69]. Hence amphibians are considered as ideal indicator species in determining overall health of ecosystem (Mason et al., 2003; Rios et al., 2017)^[47, 55]. Despite tadpoles having high LC 50 values for neonicotinoids (100-219 mg/L, Feng et al., 2004; Sanchez- Bayo 2012; Anderson et al., 2015) ^[23, 2], sub lethal doses can have dire consequences (Boone & semlitsch, 2002)

^[7]. Holtswarth et al. (2019) ^[33] had shown that exposure to concentration of neonicotinoids even at lowest dosage 0.25ug/L, tadpoles became less active with little swimming distance. Such less reactive behaviours can be detrimental as it makes them susceptible to predation due to less foraging; (Boone and semlitsh, 2002) ^[7]. Apart from these assessing corticosterone concentrations has become popular tool in determining the stress levels and immune systems of amphibians (Belden and kiesecker, 2005; Davis et al., 2008; Mcmahon et al., 2011) ^[3, 15]. When an animal encounters stress glucocorticosteroids suppress the important functions and mediates energy requirements (Romero, 2002)^[56]. When the concentrations of chemicals with long half-lives of 7-353 d for thiamethoxam and 148-6932 days, energy needed for other body functions is depleted affecting immunity as well (Belden & keisecker 2005) ^[3]. Davis *et al.*, 2008 ^[15] has reported that both neutrophils whose response to stress, inflammation (Davis et al., 2008; Shutler et al., 2009)^[15] and leucocytes antibody producer (Davis et al., 2008; shutler et al., 2009)^[15] account for 80% of leukocytes.

5. Conclusion

Thus amphibians and fishes help in rice field ecosystem in plethora of ways. The use of IFF integrated rice fish frog farming has proven to be the best in terms of increasing the yield, reducing the insect pests however ecological rehabitation of frogs should be taken care of because most of the frog species are subjected to foreign trade. Hence by adding both the fishes and frogs raises rice yields significantly can thus contributing to a sustainable agriculture.

6. References

- 1. Aliko V, Biba A. Micronuclei induction in ranidae & buffonidae tadpoles by the pyrethroid insecticide lambdacyhalothrin. Journal of Ecosystems and Ecology Science (IJEES). 2011;1:43-48.
- 2. Anderson JC, Dubetz C, Palace VP. Neonicotinoids in the Canadian aquatic environment: A literature review on current use products with a focus on fate, exposure, and biological effects. Science of the Total Environment. 2015;505:409-422.
- 3. Belden LK, Kiesecker JM. Glucocorticosteroid hormone treatment of larval tree frogs increases infection by *Alaria* sp. trematode cercariae. Journal of Parasitology. 2005;91(3):686-688.

- Berrill M, Bertram S, McGilliray L, Kolohon M, Pauli B. Effects of low concentrations of forest-use pesticides on frog embryos and tadpoles. Environmental Toxicology and Chemistry: An International Journal. 1994;13(4):657-664.
- Bhattacharyya P, Sinhababu DP, Roy KS, Dash PK, Sahu PK, Dandapat R, *et al.* Effect of fish species on methane and nitrous oxide emission in relation to soil C, N pools and enzymatic activities in rainfed shallow lowland ricefish farming system. Agric. Ecosyst. Environ. 2013;176:53-62. doi: 10.1016/j.agee.2013.05.015.
- 6. Blaustein AR, Han BA, Relyea RA, Johnson PT, Buck JC, Gervasi SS, *et al.* The complexity of amphibian population declines: understanding the role of cofactors in driving amphibian losses. Annals of the New York Academy of Sciences. 2011;1223(1):108-119.
- 7. Boone MD, Semlitsch RD. Interactions of an insecticide with competition and pond drying in amphibian communities. Ecological applications. 2002;12(1):307-316.
- 8. Cai R. Rice-fish culture in China: the past, present, and future. Rice-fish culture in China, 1995, 3-14.
- Carey C. Infectious disease and worldwide declines of amphibian populations, with comments on emerging diseases in coral reef organisms and in humans. Environmental Health Perspectives. 2000;108(1):143-150.
- Carlson KM, Gerber JS, Mueller ND, Herrero M, MacDonald GK, Brauman KA, *et al.*, Greenhouse gas emissions intensity of global croplands. Nat. Clim. Change. 2017;7:63-68.

https://doi.org/10.1038/nclimate3158

- 11. Cataño HC, Carranza E, Huamaní C, Hernández AF. Plasma cholinesterase levels and health symptoms in Peruvian farm workers exposed to organophosphate pesticides. Archives of environmental contamination and toxicology. 2008;55(1):153-159.
- 12. Connolly CN. Nerve agents in honey. Science. 2017;358(6359):38-39.
- 13. Crosby EB, Bailey JM, Oliveri AN, Levin ED. Neurobehavioral impairments caused by developmental imidacloprid exposure in zebrafish. Neurotoxic ology and teratology. 2015;49:81-90.
- Datta A, Nayak DR, Sinhababu DP, Adhya TK. Methane and nitrous oxide emissions from an integrated rainfed rice–fish farming system of Eastern India. Agric. Ecosyst. Environ. 2009;129:228-237. doi: 10.1016/j.agee.2008.09.003.
- 15. Davis AK, Maerz JC. Comparison of hematological stress indicators in recently captured and captive paedomorphic mole salamanders, *Ambystoma talpoideum*. Copeia, 2008;(3):613-617.
- DeCant J, Barrett M. Clothianidin registration of prosper T400 seed treatment on mustard seed (Oilseed and condiment) and Poncho/Votivo seed treatment on cotton. US Environmental Protection Agency, Washington DC. 2010.
- 17. Dey A, Dinesh R. Rice and wheat production in India: An overtime study on growth and instability. Journal of Pharmacognosy and Phytochemistry. 2020;9(2):158-161.
- Dunson WA, Wyman RL, Corbett ES. A symposium on amphibian declines and habitat acidification. Journal of Herpetology, 1992, 349-352.
- 19. DWR Vision (2050). Directorate of Weed Research,

Indian Council of Agricultural Research. New Delhi. 1st to 2 nd 2015. Jeschke, P., Nauen, R., Schindler, M., & Elbert, A. Overview of the status and global strategy for neonicotinoids. Journal of agricultural and food chemistry. 2011;59(7):2897-2908.

- 20. EPA (Environmental protection agency). Name of chemical: thiacloprid reason for issuance: conditional registration Date issued: September 26, 2003; Office of Prevention and Toxic substances (7501C), Oregon state; Salem, OR, USA, 2003.
- 21. Fang K, Yi X, Dai W, Gao H, Cao L. Effects of Integrated Rice-Frog Farming on Paddy Field Greenhouse Gas Emissions. International journal of environmental research and public health. 2019;16(11):1930.

https://doi.org/10.3390/ijerph16111930

- 22. Fanta E, Rios FSA, Romão S, Vianna ACC, Freiberger S. Histopathology of the fish *Corydoras paleatus* contaminated with sublethal levels of organophosphorus in water and food. Ecotoxicology and environmental safety. 2003;54(2):119-130.
- Feng S, Kong Z, Wang X, Zhao L, Peng P. Acute toxicity and genotoxicity of two novel pesticides on amphibian, Rana N. Hallowell. Chemosphere. 2004;56(5):457-463.
- 24. Finnegan MC, Baxter LR, Maul JD, Hanson ML, Hoekstra PF. Comprehensive characterization of the acute and chronic toxicity of the neonicotinoid insecticide thiamethoxam to a suite of aquatic primary producers, invertebrates, and fish. Environmental toxicology and chemistry. 2017;36(10):2838-2848.
- 25. Fournier M, Chevalier G, Nadeau D, Trottier B, Krzystyniak K. Virus-pesticide interactions with murine cellular immunity after sublethal exposure to dieldrin and aminocarb. Journal of Toxicology and Environmental Health, Part A Current. 1988;25(1):103-118.
- Frei M, Becker K. Integrated rice-fish production and methane emission under greenhouse conditions. Agric. Ecosyst. Environ. 2005;107:51-56. doi: 10.1016/j.agee.2004.10.026.
- Frei M, Becker K. Integrated rice-fish culture: Coupled production saves resources. In Natural Resources Forum. Oxford, UK: Blackwell Publishing, Ltd. 2005 May;29(2):135-143.
- 28. Fulton MH, Chambers JE. The toxic and teratogenic effects of selected organophosphorus compounds on the embryos of three species of amphibians. Toxicology Letters. 1985;26(2-3):175-180.
- Gongpeng LIU, Yuzhu Zhang, Kailin Chen, Yang LIU, Guoqi ZHU, Baohua Fang. Effects of rice-bullfrog mixed cultivation on rice planthoppers and rice yield. Chinese Journal of Biological Control. 2013;29(2):207.
- Goulson D. An overview of the environmental risks posed by neonicotinoid insecticides. Journal of Applied Ecology. 2013;50(4):977-987.
- 31. Guruge KS, Tanabe S. Contamination by persistent organochlorines and butyltin compounds in the west coast of Sri Lanka. Marine Pollution Bulletin. 2001;42(3):179-186.
- 32. Hayasaka D, Korenaga T, Suzuki K, Sánchez-Bayo F, Goka K. Differences in susceptibility of five cladoceran species to two systemic insecticides, imidacloprid and fipronil. Ecotoxicology. 2012;21(2):421-427.
- 33. Holtswarth JN, Rowland FE, Puglis HJ, Hladik ML, Webb EB. Effects of the neonicotinoid insecticide

leopard frog (*Rana* 48. McMahon TA,

clothianidin on southern leopard frog (*Rana sphenocephala*) tadpole behavior. Bulletin of Environmental Contamination and Toxicology. 2019;103(5):717-722. http://ijarse.com/images/fullpdf/1519301763_SVCET206 9ijarse.pdf

- 34. Iwakuma T, Otsuki A. Role of chironomid larvae in reducing rate of nutrient release from lake sediment: Evaluation by a mathematical model. Internationale Vereinigung für theoretische und angewandte Limnologie: Verhandlungen. 1991;24(5):3056-3062.
- 35. Jintong Y. Rice fish culture and its macro development in ecological agriculture. In Rice fish culture in China. IDRC, Ottawa, ON, CA. 1995.
- Johnson PT, Lunde KB, Ritchie EG, Launer AE. The effect of trematode infection on amphibian limb development and survivorship. Science. 1999;284(5415):802-804.
- Kharat SB, Manjrekar MD, Dumbre RB, Dalvi CS. Role of Indian bull frog, in controlling rice pests. Journal of Maharashtra Agricultural Universities. 1983;8(3):223-225.
- Konradsen F, VAN der Hoek W, Cole DC, Hutchinson G, Daisley H, Singh S, *et al.* Reducing acute poisoning in developing countries-options for restricting the availability of pesticides. Toxicology. 2003;192(2-3):249-261.
- Krupke CH, Hunt GJ, Eitzer BD, Andino G, Given K. Multiple routes of pesticide exposure for honey bees living near agricultural fields. PLoS one. 2012;7(1):e29268.
- 40. Krzystyniak K, Hugo P, Flipo D, Fournier M. Increased susceptibility to mouse hepatitis virus 3 of peritoneal macrophages exposed to dieldrin. Toxicology and applied pharmacology. 1985;80(3):397-408.
- 41. Liang X, Li H, Wang S, Ye Y, Ji Y, Tian G, *et al.* Nitrogen management to reduce yield-scaled global warming potential in rice. Field Crops Res. 2013;146:66-74. doi: 10.1016/j.fcr.2013.03.002.
- 42. Liang X, Li H, Wang S, Ye Y, Ji Y, Tian G, *et al.* Nitrogen management to reduce yield-scaled global warming potential in rice. Field Crops Res. 2013;146:66-74. doi: 10.1016/j.fcr.2013.03.002.
- 43. Liu GP, Zhang YZ, Huang ZN, Chen KL, Liu Y, Zhu GQ, *et al.* Effects of Rice-Bullfrog Mixed Cultivation on Rice Planthoppers and Rice Yield. Chin. J Biol. Contr. 2013;29:207-213.
- 44. Longhurst C, Babcock JM, Denholm I, Gorman K, Thomas JD, Sparks TC. Cross-resistance relationships of the sulfoximine insecticide sulfoxaflor with neonicotinoids and other insecticides in the whiteflies *Bemisia tabaci* and *Trialeurodes vaporariorum*. Pest Management Science. 2013;69(7):809-813.
- 45. Luebke RW, Hodson PV, Faisal M, Ross PS, Grasman KA, Zelikoff J. Aquatic pollution-induced immunotoxicity in wildlife species. Fundamental and Applied Toxicology. 1997;37(1):1-15.
- 46. Marzaro M, Vivan L, Targa A, Mazzon L, Mori N, Greatti M, *et al.* Lethal aerial powdering of honey bees with neonicotinoids from fragments of maize seed coat. Bulletin of Insect ology. 2011;64(1):119-126.
- 47. Mason NW, MacGillivray K, Steel JB, Wilson JB. An index of functional diversity. Journal of Vegetation Science. 2003;14(4):571-578.

- 48. McMahon TA, Halstead NT, Johnson S, Raffel TR, Romansic JM, Crumrine PW, *et al.* The fungicide chlorothalonil is nonlinearly associated with corticosterone levels, immunity, and mortality in amphibians. Environmental health perspectives. 2011;119(8):1098-1103.
- 49. Mondal D, Ghosh A, Roy D, Kumar A, Shamurailatpam D, Bera S, *et al.* Yield loss assessment of rice (*Oryza Sativa* L.) due to different biotic stresses under system of rice intensification (SRI). Journal of Entomology and Zoology Studies. 2017;5(4):1974-1980.
- Mora C, Spirandelli D, Franklin EC, Lynham J, Kantar MB, Miles W, *et al.*, Broad threat to humanity from cumulative climate hazards intensified by greenhouse gas emissions. Nat. Clim. Chang. 2018;8:1062-1071. doi: 10.1038/s41558-018-0315-6.
- 51. Morrissey CA, Mineau P, Devries JH, Sanchez-Bayo F, Liess M, Cavallaro MC, *et al.* Neonicotinoid contamination of global surface waters and associated risk to aquatic invertebrates: A review. Environment international. 2015;74:291-303.
- 52. Mustard JA, Gott A, Scott J, Chavarria NL, Wright GA. Honeybees fail to discriminate floral scents in a complex learning task after consuming a neonicotinoid pesticide. Journal of Experimental Biology. 2020;223(5):jeb217174.
- 53. O'Halloran K, Ahokas JT, Wright PFA. *In vitro* responses of fish immune cells to three classes of pesticides. Modulators of immune response: the evolutionary trail, Fair Haven, NJ: SOS Publications, 1996, 535-8.
- 54. Pounds JA, Fogden MP, Campbell JH. Biological response to climate change on a tropical mountain. Nature. 1999;398(6728):611-615.
- 55. Rios FM, Wilcoxen TE, Zimmerman LM. Effects of imidacloprid on Rana catesbeiana immune and nervous system. Chemosphere. 2017;188:465-469.
- 56. Romero LM. Seasonal changes in plasma glucocorticoid concentrations in free-living vertebrates. General and comparative endocrinology. 2002;128(1):1-24.
- 57. Saha S. Comparative study on efficacy of sulfonylurea herbicides and traditional recommended herbicides in transplanted rice (*Oryza sativa* L.). Indian Journal of Agronomy. 2006;51(4):304-06.
- 58. Sánchez-Bayo F. Insecticides mode of action in relation to their toxicity to non-target organisms. J Environ. Anal. Toxicol. 2012;S4:S4-002.
- 59. Sánchez-Bayo F, Goka K, Hayasaka D. Contamination of the aquatic environment with neonicotinoids and its implication for ecosystems. Frontiers in Environmental Science. 2016;4:71.
- 60. Shugen P, Zhechun H, Jicheng Z. Ecological mechanisms for increasing rice and fish production. In Rice fish culture in China. IDRC, Ottawa, ON, CA, 1995.
- 61. Shutler D, Smith TG, Robinson SR. Relationships between leukocytes and *Hepatozoon* spp. in green frogs, Rana clamitans. Journal of Wildlife Diseases. 2009;45(1):67-72.
- 62. Simon-Delso N, Amaral-Rogers V, Belzunces LP, Bonmatin JM, Chagnon M, Downs C, *et al.* Systemic insecticides (neonicotinoids and fipronil): trends, uses, mode of action and metabolites. Environmental Science and Pollution Research. 2015;22(1):5-34.
- 63. Simon-Delso N, Amaral-Rogers V, Belzunces LP,

Bonmatin JM, Chagnon M, Downs C, *et al.* Systemic insecticides (Neonicotinoids and fipronil): trends, uses, mode of action and metabolites. Environmental Science and Pollution Research. 2015;22(1):5-34.

- 64. Sparling DW, Fellers GM, McConnell LL. Pesticides and amphibian population declines in California, USA. Environmental Toxicology and Chemistry: An International Journal. 2001;20(7):1591-1595.
- 65. Tapparo A, Marton D, Giorio C, Zanella A, Soldà L, Marzaro M, *et al.* Assessment of the environmental exposure of honeybees to particulate matter containing neonicotinoid insecticides coming from corn coated seeds. Environmental science & technology. 2012;46(5):2592-2599.
- 66. Teng Q, Hu XF, Luo F, Cheng C, Ge X, Yang M, *et al.* Influences of introducing frogs in the paddy fields on soil properties and rice growth. Journal of Soils and Sediments. 2016;16(1):51-61.
- 67. Teng Q, Hu XF, Luo F. *et al.* Influences of introducing frogs in the paddy fields on soil properties and rice growth. J Soils Sediments. 2016;16:51-61. https://doi.org/10.1007/s11368-015-1183-6
- Tison L, Hahn ML, Holtz S, Rößner A, Greggers U, Bischoff G, *et al.* Honey bees' behavior is impaired by chronic exposure to the neonicotinoid thiacloprid in the field. Environmental science & technology. 2016;50(13):7218-7227.
- 69. Van Meter RJ, Glinski DA, Hong T, Cyterski M, Henderson WM, Purucker ST. Estimating terrestrial amphibian pesticide body burden through dermal exposure. Environmental Pollution. 2014;193:262-268.
- Velisek J, Stara A. Effect of thiacloprid on early life stages of common carp (*Cyprinus carpio*). Chemosphere. 2018;194:481-487.
- 71. Walker CH. Organic pollutants: an ecotoxicological perspective. CRC press, 2008.
- 72. Xia L, Lam SK, Chen D, Wang J, Tang Q, Yan X. Can knowledge-based N management produce more staple grain with lower greenhouse gas emission and reactive nitrogen pollution? A meta-analysis. Global Change Biology. 2017;23(5):1917-1925.
- 73. Xia L, Li X, Ma Q, Lam SK, Wolf B, Kiese R, *et al.* Simultaneous quantification of N2, NH3 and N2O emissions from a flooded paddy field under different N fertilization regimes. Global Change Biology. 2020;26(4):2292-2303.
- 74. Zelikoff JT, Wang W, Islam N, Flescher E, Twerdok LE. Immune responses of fish as biomarkers to predict the health of aquatic pollution: application of laboratory assays for field studies. In Ecotoxicology: response, biomarkers and risk assessment, an OECD workshop. SOS Publications, Fair Haven, NJ, 1997, 263-279p.
- 75. Zhan M, Cao CG, Wang JP, Li CF, Yuan WL. Greenhouse gas emission from an integrated rice-duck system and its global warming potentials. Acta Scientiae Circumstantiae. 2009;29:420-426.
- 76. Zhou M, Zhu B, Wang X, Wang Y. Long-term field measurements of annual methane and nitrous oxide emissions from a Chinese subtropical wheat-rice rotation system. Soil Biology and Biochemistry. 2017;115:21.