



ISSN (E): 2277-7695

ISSN (P): 2349-8242

NAAS Rating: 5.23

TPI 2022; 11(5): 316-321

© 2022 TPI

www.thepharmajournal.com

Received: 08-03-2022

Accepted: 13-04-2022

Dharamraj Kumar

M.Sc. Student, Department of Vegetable Science, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India

Anjali Rani

M.Sc. Student, Department of Genetics and Plant Breeding, Sardar Vallabhbhai Patel University of Agriculture and Technology, Meerut, Uttar Pradesh, India

Jaswant Prajapati

M.Sc. Student, Department of Vegetable Science, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India

Sudipta Mahato

M.Sc Student, Department of Plant Pathology, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India

Narendra Pratap Verma

Research scholar, Department of Crop Physiology, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India

Astha Vishwaraj

M.Sc. Student, Department of Vegetable Science, Sardar Vallabhbhai Patel University of Agriculture and Technology, Meerut, Uttar Pradesh, India

Suman Kumar

M.Sc. Student, Department of Horticulture, Assam Agricultural University, Jorhat, Assam, India

Devesh Sanjay Pardhi

M.Sc Student, Department of Crop Physiology, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India

Corresponding Author:

Dharamraj Kumar

M.Sc. Student, Department of Vegetable Science, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj, Ayodhya, Uttar Pradesh, India

Breeding for biotic stresses resistance in tomato: A review

Dharamraj Kumar, Anjali Rani, Jaswant Prajapati, Sudipta Mahato, Narendra Pratap Verma, Astha Vishwaraj, Suman Kumar and Devesh Sanjay Pardhi

Abstract

Pests and diseases that affect tomato production have a major impact. Although fungicides and pesticides have helped to manage plant diseases and insects/pests, indiscriminate use of hazardous chemicals pollutes the environment, causes chemical residues in tomatoes to exceed threshold values, and leads to the establishment of novel races/biotypes. Breeding for the production of biotic resistant cultivars is thus a primary goal of vegetable breeders at this point. Basic research into the genetic origins of pest and disease resistance in tomato crops, as well as host-pathogen interactions, has resulted in the production of high-yielding genetically resistant cultivars. Sources of resistance, as well as genetic information, are required for the production of resistant cultivars and pre-breeding line.

Keywords: Inheritance of resistance, Resistance sources, Biotechnological approaches, Grafting, Molecular Marker

Introduction

Tomato (*Solanum lycopersicum* L.) is one of the leading vegetable crops of Solanaceae [1]. It has become an important commercial crop when we talk about the human nutrition. Tomatoes were originated in Peru (South America) and primary domesticated in Mexico on the basis of availability of numerous cultivated and wild Species of the tomato initiate in this area [2]. It provides nutrients like beta-carotene, lycopene, vitamin C and flavonoids. Moreover tomato has achieved high popularity especially in latest years because of lycopene's anti-oxidative activities and anti-cancer functions [3]. conventional cultivated tomato lack genetic diversity. Hence it has been suggested to transmit the desired resistance traits from their wild type Species [4]. Upon stress observation, transcription factors (TFs) attach to their target genes to regulate their expression and orchestrate biochemical and physiological modifications critical for stress tolerance and the alteration of plant growth [5]. The significant incidences of diseases and pests in tomato production are the main issue. Pesticides used indiscriminately to manage diseases, insect pests, and nematodes are detrimental to human health and the environment. On the contrary the progress in the development of insect resistance is very limited. Host plant resistance is the economical method but such resistance against insect pest is not at all stable due to population pressure of insects on the host, therefore there is evolution of novel biotypes and breakdown of resistance [6].

Varieties released till date are having one or more undesirable characters associated with them and hence have not gained the popularity. Insect-pests are accountable for falling about 40% yield in vegetables [7] Tomato (*Solanum lycopersicum* L.) possesses unique properties, as it is both an economically important crop, the first vegetable in production in the world (FAOSTAT 2011), and a model plant species, due to its diploid, relatively compact, and recently sequenced genome and its large genetic and genomic resources [8]. The biotrophic ascomycete *Oidium neolyopersici* (causing PM) is one of the economically most important foliar pathogens of tomato, both in the greenhouse and in open field conditions [9].

Table 1: Major Diseases and Insect-Pests Attacking Tomato Crop

Crop	Major Diseases	Major Insect-Pests
Tomato	ToL CV, CMV, late blight, early blight, bacterial wilt, root-knot, nematodes, septoria leaf spot, tomato spotted wilt virus	Fruit borer, white fly, aphid

Source: Dhall R.K (2015) [4]

The tomato leaf curl New Delhi virus (ToLCNDV) is a bipartite begomovirus species (genus *Begomovirus*, family *Geminiviridae*) whose isolates are transmitted in nature by the whitefly *Bemisia tabaci* (family *Aleyrodidae*, order *Hemiptera*) in a circulative and constant manner [10]. ToLCNDV is an economically significant begomovirus identified to cause overwhelming damage to tomato (*Solanum lycopersicum*) production, and it is more prevalent in northern India [11, 12, 13, 14]. Wilt diseases of tomatoes can be caused by fungal, bacterial, viral, and nematode pathogens, and a biotic factor. There are over 100 different special forms of *Fusarium*

oxysporum, each usually with a specific host on which they can cause disease by *Fusarium oxysporum*. The fungus is the soil borne hyphomycete and is one of more than 100 *F. oxysporum* that causes vascular wilts of flowering plants [15]. Tomatoes are attacked by a variety of insects, including mites, whiteflies, aphids, Lepidoptera (such as tomato fruitworm, beet armyworm, cotton bollworm, southern armyworm, soybean podworm, and Egyptian cottonworm), Coleoptera (such as Colorado potato beetle and tobacco flea beetle), Diptera (such as leafminers and fruit fly), thrips, sinkbugs, and cutworms).

Table 2: Biotic Stresses and their Source of Resistance in Tomato

Resistant gene	Resistance against	Resistance Source	References
Asc-1	<i>Alternaria alternata</i> f. sp. <i>lycopersici</i>	<i>S. Lycopersicum</i>	[16]
Am	Alfalfa mosaic virus	<i>S. Habrochaites</i>	[17]
Bs4	<i>Xanthomonas campestris</i>	<i>S. Lycopersicum</i>	[18]
Cmv	Cucumber mosaic virus	<i>S. Chilense</i>	[19]
Cf-1	<i>Cladosporium fulvum</i>	<i>S. Lycopersicum</i> Var <i>Cerasiforme</i>	[20]
Cf-2	<i>Cladosporium fulvum</i>	<i>S. Pimpinellifolium</i>	[21]
Cf-3	<i>Cladosporium fulvum</i>	<i>S. Pimpinellifolium</i>	[22]
Frl	<i>Fusarium oxysporum</i> f.sp. <i>radicis-lycopersici</i>		[23]
Hero	<i>Globodera rostochiensis</i>	<i>S. Pimpinellifolium</i>	[24]
I	<i>Fusarium oxysporum</i> formae speciales <i>lycopersici</i>	<i>S. Pimpinellifolium</i>	[25]
Mi-1.2	<i>Meloidogyne</i> spp	<i>S. Peruvianum</i>	[26]
Ph-1	<i>Phytophthora infestans</i>	<i>S. Pimpinellifolium</i>	[27]
Sw-5	Tomato spotted wilt virus, tomato chlorotic spot virus	<i>S. Peruvianum</i>	[28]
Sw-7	Tomato spotted wilt virus	<i>S. Chilense</i>	[29]
Ve1	<i>Verticillium dahliae</i>	<i>S. Lycopersicum</i>	[30]
Ty-1	Tomato yellow leaf curl virus	<i>S. Chilense</i>	[31]
Ty-2	Tomato yellow leaf curl virus	<i>S. Habrochaites</i>	[32]
Ty-3	Tomato yellow leaf curl virus, Tomato mosaic virus	<i>S. Chilense</i>	[33]
Ty-4	Tomato yellow leaf curl virus	<i>S. Chilense</i>	[34]
	Tomato yellow leaf curl virus	<i>S. Peruvianum</i>	[35]
ol-1	<i>Oidium neolyopersici</i>	<i>S. Habrochaites</i>	[36]
ol-2	<i>Oidium neolyopersici</i>	<i>S. Lycopersicum</i> Var <i>Cerasiforme</i>	[37]
ol-3	<i>Oidium neolyopersici</i>	<i>S. Habrochaites</i>	[38]

Biotechnological Interventions

Marker-assisted breeding can also be used to introduce certain desired genes from wild relatives into domesticated species, known as introgression. Because the desired gene is only found in one or a few places in the genome, markers indicating other parts of the wild type chromosomes can be selected against, resulting in the elimination of those (usually undesired) genes from the progeny. Genetic engineering has been attempted for developing plants resistant to various

biotic stresses such as viruses, bacteria, fungus and insect pests. Significant resistance to tobacco mosaic virus (TMV) infection termed "Coat protein mediated protection" has been achieved by expressing only the coat produced similar results in transgenic tomato plants against a broad spectrum of plant viruses including alfalfa mosaic virus, cucumber mosaic virus, (39) Progress in engineering insect resistance in transgenic tomato has been achieved [40, 41].

Table 3: Molecular markers used for tomato crop

Marker type	Crop	Reference
SNP	tomato	[42, 43, 44, 45, 46, 47, 48]
CAPS	tomato	[49]
SSR	tomato	[50, 51, 52, 53, 54, 55]
AFLP	tomato	[56]
RFLP	tomato	[57]
SRAP	tomato	[58, 59]
SCAR	tomato	[60]

Table 4: Transgenes Resistant to Viral Diseases in tomato Crop

Crop/Class of Transgene	Origin of Transgene	Resistant to
Cp gene	ToMV	Tomato mosaic virus
Cp gene	TYLCV	Tomato yellow leaf curl virus
Antisense RNA	ToMV	Tomato mosaic virus
Satellite RNA	CMV	Cucumber mosaic virus
N gene	TSWV	Tomato spotted wilt virus
Truncated CI gene	CMV	Cucumber mosaic virus
Two Cp genes	CMV	Cucumber mosaic virus

Source: Dhall R.K (2015) [4]

Grafting

To minimize main crop loss caused by infection of soil born disease exacerbated by successive cropping, grafted seedling vegetable production was developed in Japan and Korea. The vigorous roots of selected rootstock can show excellent tolerance to serious soil-borne diseases, such as those caused by *Fusarium*, *Verticillium*, *Pseudomonas*, *Phytophthora*, *Didymella bryoniae*, *Monosporascus cannonballus*, and nematodes [61, 62] yet though the degree of tolerance varies

considerably with the rootstocks. Even the scion infection of certain virus diseases (TMV races) could be markedly influenced by virus resistant rootstocks depending upon the level of resistance in scion and rootstocks. The disease tolerance in grafted seedlings may be entirely due to the tolerance of rootstock roots to such diseases. It is generally accepted, on the other hand, that the disease vulnerable characteristics of the scion are not transported to the rootstock [63, 64].

Table 5: Rootstocks for Controlled Diseases and Pests of tomato crop by Grafting

Crop	Rootstock	Disease Resistance
Tomato	<i>S. habrochaites</i>	CR
	Brinjal (EG-203 and EG-195) <i>S. Melongena</i> , <i>S. Lycopersicum L.</i> × <i>S. Habrochaites</i>	BW, NMR MDR

Source: Dhall R.K (2015) [4]

Solanum Pennellii: Resistance to *Alternaria alternata* f. sp. *Lycopersici* [65]

S. Habrochaites: Resistance to *Pseudomonas syringe* pv. tomato race 1 [66]

S. Chilense: Resistance to diseases (CMV, TYLCV) Zamir *et al.* [67]

S. Neorickii: Resistant to *Botrytis cinerea* [68]

S. Pimpinellifolium: Colour, quality, resistance to bacterial wilt (69), late blight [69]

S. Lycopersicum: Resistance to fungi and root rot [70]

Cerasiforme S. Peruvianum: Resistance to tomato spotted wilt virus and RKN [71, 72, 73]

Future Strategies

One of the best strategies for minimizing losses due to disease/insect occurrence is to develop resistant or tolerant cultivars. Although there are effective chemical control strategies for a variety of insect pests and diseases, the expense of such pesticides is quite high, and many chemicals have long-term impacts. There is a need to conduct an active research programme to make the most use of the country's existing germplasm, particularly wild relatives, for the establishment of pre-bred lines that may be used successfully against certain biotic stresses as and when resistance sources are required. When methods of artificial inoculation on plantlets are devised, breeding for disease and pest resistance becomes considerably more successful. In addition, through gene pyramiding, major efforts should be made to generate varieties or hybrids with various disease resistance. Breeding should be prioritised for integrated disease and insect pest management, such as leaf curl and TMV in tomatoes. An effective collaboration of breeders and plant pathologists/entomologists will be of high proposition to tackle this problem.

References

- Mueller LA, Tanksley SD, Giovannoni JJ, van Eck J, Stack S, Choi D, *et al.* The tomato sequencing project, the first cornerstone of the international Solanaceae Project (SOL). *Comp. Funct. Genomics.* 2005;6:153-158. doi: 10.1002/cfg.468.
- Rick CM. Origin of cultivated tomato, current status and the problem. *International Botanical Congress*, 1969,180p.
- Fentik DA. Review on Genetics and Breeding of Tomato (*Lycopersicon esculentum* Mill.). *Advances in Crop Science and Technology.* 2017;5:5.
- Dhall RK. Breeding for biotic stresses resistance in vegetable crops: a review. *Journal of Crop Science Technology.* 2015;4:13-27.
- Rick CM, Chetelat RT. Utilization of related wild species for tomato improvement. *Acta Horticulturae.* 1995;412:21-38.
- Hichri I, Muhovski Y, Zizkova E, Dobrev PI, Franco-Zorrilla JM, Solano R, *et al.* The *Solanum lycopersicum* zinc finger 2 cysteine-2/histidine-2 repressor-like transcription factor regulates development and tolerance to salinity in tomato and *Arabidopsis*. *Plant Physiol.* 2014;164:1967-1990. doi: 10.1104/pp.113.225920
- Singh M, Chakraborty S, Kumar S, *et al.* Genetic Engineering for Insect Resistance in Vegetable Crops. *Veg Sci.* 2000;27(2):105-16p.
- Ranjan A, Ichihashi Y, Sinha NR. The tomato genome: implications for plant breeding, genomics and evolution. *Genome Biol.* 2012;13:167
- Jones H, Whipps JM, Gurr SJ. The tomato powdery mildew fungus *Oidium neolyopersici*. *Mol Plant Pathol.* 2001;2:303-309
- Brown JK, Fauquet CM, Briddon RW, Zerbini FM, Moriones E, Navas-Castillo J. Family *Geminiviridae*. In *Virus Taxonomy 9th Report, Proceedings of the*

- International Committee on Taxonomy of Viruses, 2001.
11. King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ. Eds., Elsevier Academic Press: London, UK, 2012, 351-373p. [Google Scholar]
 12. Varma A, Malathi VG. Emerging geminivirus problems: A serious threat to crop production. *Ann. Appl. Biol.* 2003;142:145-164. [Google Scholar] [CrossRef]
 13. Zaidi SSS, Martin DP, Amin I, Farooq M, Mansoor S. Tomato leaf curl New Delhi virus: A widespread bipartite begomovirus in the territory of monopartite begomoviruses. *Mol. Plant Pathol.* 2016;18:901-911. [Google Scholar] [CrossRef] [PubMed]
 14. Chakraborty S. Tomato leaf curl viruses from India (*Geminiviridae*). In *Encyclopedia of Virology*; Mahy, B.W.J., Van Regenmortel, M.H.V., Eds.; Elsevier: London, UK, 2008,124-133p. [Google Scholar]
 15. Mansoor S, Khan SH, Saeed M, Bashir A, Zafar Y, Malik KA. Evidence for the association of a bipartite geminivirus with tomato leaf curl disease in Pakistan. *Plant Dis.* 1997;81:958. [Google Scholar] [CrossRef]
 16. Domsch KH, Gams W, Anderson TH. *Compendium of soil fungi*, Academic Press, New York, USA, 1980.
 17. Brandwagt BF, Kneppers TJ, Nijkamp HJ, Hille J. Overexpression of the tomato Asc-1 gene mediates high insensitivity to AAL toxins and fumonisin B1 in tomato hairy roots and confers resistance to *Alternaria alternata* f. sp. *lycopersici* in *Nicotiana umbratica* plants. *Mol Plant Microbe Interact.* 2002;15:35-42.
 18. Parrella G, Moretti A, Gognalons P, Lesage ML, Marchoux G *et al.* The Am Gene Controlling Resistance to Alfalfa mosaic virus in Tomato Is Located in the Cluster of Dominant Resistance Genes on Chromosome 6. *Phytopathol.* 2004;94:345-350.
 19. Schornack S, Ballvora A, Gurlebeck D, Peart J, Baulcombe D *et al.* The tomato resistance protein Bs4 is a predicted non-nuclear TIR-NB-LRR protein that mediates defense responses to severely truncated derivatives of AvrBs4 and overexpressed AvrBs3. *Plant J.* 2004;37:46-60.
 20. Stamova BS, Chetelat RT. Inheritance and genetic mapping of cucumber mosaic virus resistance introgressed from *Lycopersicon chilense* into tomato. *Theor Appl Genet.* 2000;101:527-537.
 21. Langford AN. The parasitism of *Cladosporium fulvum* Cooke and the genetics of resistance to it. *Can J Res.* 1937;15:10828
 22. Dixon MS, Jones DA, Keddie JS, Thomas CM, Harrison K, Jones JDG. The tomato Cf-2 disease resistance locus comprises two functional genes encoding leucine-rich repeat proteins. *Cell.* 1996;84:451-60.
 23. Hammond-Kosack KE, Jones JDG. Incomplete dominance of tomato Cf genes for resistance to *Cladosporium fulvum*. *Mol Plant Microbe Interact.* 1993;7:58-70.
 24. Vakalounalou DJ, Laterrot H, Moretti A, Ligoixakis EL, Smardas K. Linkage between Frl (*Fusarium oxysporum* f.sp. *radicis-lycopersici* resistance) and Tm-2 (tobacco mosaic virus resistance-2) loci in tomato (*Lycopersicon esculentum*). *Ann Appl Biol.* 1997;130:319-23.
 25. Ernst K, Kumar A, Kriseleit D, Kloos DU, Phillips MS *et al.* The broad spectrum potato cyst nematode resistance gene (Hero) from tomato is the only member of a large gene family of NBS-LRR genes with an unusual amino acid repeat in the LRR region. *Plant J.* 2002;31:127-36.22
 26. Sela-Buurlage MB, Budai-Hadrian O, Pan Q, Carmel-Goren L, Vunsch R *et al.* Genome-wide dissection of *Fusarium* resistance in tomato reveals multiple complex loci. *Mol Genet Genomics.* 2001;265:1104-1111.
 27. Milligan SB, Bodeau J, Yaghoobi J, Kaloshian I, Zabel P *et al.* The root knot nematode resistance gene Mi from tomato is a member of the leucine zipper, nucleotide binding, leucine-rich repeat family of plant genes. *Plant Cell.* 1998;10:1307-1319
 28. Bonde R, Murphy EF. Resistance of certain tomato varieties and crosses to late blight. *Maine Agr Exp Sta Bull.* 1952;497:5-15
 29. Brommenschkel SH, Tankslet SD. Map-based cloning of the tomato genomic region that spans the Sw-5 tospovirus resistance gene in tomato. *Mol Genet Genomics.* 1997;256:121-126.
 30. Dockter KG, O'neil DS, Price DL, Scott J, Stevens MR. Molecular Mapping of the tomato spotted wilt virus resistance gene Sw-7 in tomato. In. *American Society for Horticultural Science*. St. Louis, MO, 2009.
 31. Hammond-Kosack KE, Jones JDG. Incomplete dominance of tomato Cf genes for resistance to *Cladosporium fulvum*. *Mol Plant Microbe Interact.* 1993;7:58-70.
 32. Zamir D, Ekstein-Michelson I, Zakay Y, Navot N, Zeidan M *et al.* Mapping and introgression of a tomato yellow leaf curl virus tolerance gene, TY-1. *Theor Appl Genet.* 1994;88:141-146.
 33. Ji Y, Scott JW, Schuster DJ. Toward Fine Mapping of the Tomato Yellow Leaf Curl Virus Resistance Gene Ty-2 on Chromosome 11 of Tomato. *Hort Science.* 2009;44:614-8.
 34. Ji Y, Schuster DJ, Scott JW. Ty-3, a begomovirus resistance locus near the Tomato yellow leaf curl virus resistance locus Ty-1 on chromosome 6 of tomato. *Mol Breed.* 2007;20:271-284.
 35. Ji Y, Scott JW, Schuster DJ. Toward Fine Mapping of the Tomato Yellow Leaf Curl Virus Resistance Gene Ty-2 on Chromosome 11 of Tomato. *Hort Science.* 2009;44:614-8
 36. Anbinder I, Reuveni M, Azari R, Paran I, Nahon S *et al.* Molecular dissection of Tomato leaf curl virus resistance in tomato line TY172 derived from *Solanum peruvianum*. *Theor Appl Genet.* 2009;119:519-530.
 37. Huang CC, Cui YY, Weng CR, Zabel P, Lindhout P. Development of diagnostic PCR markers closely linked to the tomato powdery mildew resistance gene Ol-1 on chromosome 6 of tomato. *Theor Appl Genet.* 2000;101:918-24.
 38. De Giovanni C, Dell'orco P, Bruno A, Ciccacese F, Lotti C *et al.* Identification of PCR-based markers (RAPD, AFLP) linked to a novel mildew resistance gene (ol-2) in tomato. *Plant Sci.* 2004;166:41-8.
 39. Huang CC, Cui YY, Weng CR, Zabel P, Lindhout P. Development of diagnostic PCR markers closely linked to the tomato powdery mildew resistance gene Ol-1 on chromosome 6 of tomato. *Theor Appl Genet.* 2000;101:918-24.
 40. Shukla DD, Ward CW, Brunt AA *et al.* *Potyviridae*. *AAB Descriptions of Plant Viruses*, No. 366, 1998.
 41. Fischhoff DA, Bowdish KS, Perlak FJ *et al.* *Insect*

- Tolerant Transgenic Tomato Plants. *Biotechnology*. 1987;5:807-813.
42. Babu RM, Sajeena A, Seetharaman K *et al.* Advances in Genetically Engineered (Transgenic) Plants in Pest Management An Overview. *Crop Protection*. 2003;22:1071-1086.
 43. Jiménez-Gómez JM, Maloof JN. Sequence diversity in three tomato species: SNPs, markers, and molecular evolution. *BMC Plant Biology*. 2009;9(1):85.
 44. Sim SC, Robbins MD, Van Deynze A, Michel AP, Francis DM. Population structure and genetic differentiation associated with breeding history and selection in tomato (*Solanum lycopersicum* L.). *Heredity*. 2011;106(6):927-935.
 45. Hamilton JP, Hansey CN, Whitty BR, Stoffel K, Massa AN, Van Deynze A *et al.* Single nucleotide polymorphism discovery in elite North American potato germplasm. *BMC Genomics*. 2011;12(1):302.
 46. Sim SC, Durstewitz G, Plieske J, Wieseke R, Ganal MW, Van Deynze A *et al.* Development of a large SNP genotyping array and generation of high-density genetic maps in tomato. *PloS one*, 2012,7(7).
 47. Iquebal MA, Arora V, Verma N, Rai A, Kumar D. First whole genome based microsatellite DNA marker database of tomato for mapping and variety identification. *BMC Plant Biology*. 2013;13(1):197.
 48. Viquez-Zamora M, Vosman B, van de Geest H, Bovy A, Visser RG, Finkers R *et al.* Tomato breeding in the genomics era: insights from a SNP array. *BMC Genomics*. 2013;14(1):354.
 49. Kevei Z, King RC, Mohareb F, Sergeant MJ, Awan SZ, Thompson AJ. Resequencing at ≥ 40 -fold depth of the parental genomes of a *Solanum lycopersicum* \times *S. pimpinellifolium* recombinant inbred line population and characterization of frame-shift In Dels that are highly likely to perturb protein function. *G3: Genes, Genomes, Genetics*. 2015;5(5):971-981.
 50. Yang X, Caro M, Hutton SF, Scott JW, Guo Y, Wang X *et al.* Fine mapping of the tomato yellow leaf curl virus resistance gene Ty-2 on chromosome 11 of tomato. *Molecular Breeding*. 2014;34(2):749-760.
 51. He C, Poysa V, Yu K. Development and characterization of simple sequence repeat (SSR) markers and their use in determining relationships among *Lycopersicon esculentum* cultivars. *Theoretical and Applied Genetics*. 2003;106(2):363-373.
 52. Ruiz JJ, GarcíaMartínez S, Picó B, Gao M, Quiros CF. Genetic variability and relationship of closely related Spanish traditional cultivars of tomato as detected by SRAP and SSR markers. *Journal of the American Society for Horticultural Science*. 2005;130(1):88-94.
 53. Grushetskaya ZE, Lemesh VA, Poliksenova VD, Khotyleva LV. Mapping of the Cf-6 tomato leaf mould resistance locus using SSR markers. *Russian Journal of Genetics*. 2007;43(11):1266-1270.
 54. Sim SC, Robbins MD, Van Deynze A, Michel AP, Francis DM. Population structure and genetic differentiation associated with breeding history and selection in tomato (*Solanum lycopersicum* L.). *Heredity*. 2011;106(6):927-935.
 55. Ning L, Jing-Bin J, Jing-Fu L, Xiang-Yang X. Development of molecular marker linked to Cf-10 gene using SSR and AFLP method in tomato. *Journal of Northeast Agricultural University (English Edition)*. 2012;19(4):30-36.
 56. Todorovska E, Ivanova A, Ganeva D, Pevicharova G, Molle E, Bojinov B *et al.*, 2014.
 57. Ning L, Jing-Bin J, Jing-Fu L, Xiang-Yang X. Development of molecular marker linked to Cf-10 gene using SSR and AFLP method in tomato. *Journal of Northeast Agricultural University (English Edition)*. 2012;19(4):30-36.
 58. Tanksley SD, Ganal MW, Prince JP, De Vicente MC, Bonierbale MW, Broun P *et al.* High density molecular linkage maps of the tomato and potato genomes. *Genetics*. 1992;132(4):1141-1160.
 59. Ruiz JJ, GarcíaMartínez S, Picó B, Gao M, Quiros CF. Genetic variability and relationship of closely related Spanish traditional cultivars of tomato as detected by SRAP and SSR markers. *Journal of the American Society for Horticultural Science*. 2005;130(1):88-94.
 60. Al Shaye N, Migdadi H, Charbaji A, Alsayegh S, Daoud S, Wala AA *et al.* Genetic variation among saudi tomato (*Solanum lycopersicum* L.) landraces studied using sds-page and srp markers. *Saudi journal of biological sciences*. 2018;25(6):1007-1015.
 61. Yang X, Caro M, Hutton SF, Scott JW, Guo Y, Wang X *et al.* Fine mapping of the tomato yellow leaf curl virus resistance gene Ty-2 on chromosome 11 of tomato. *Molecular Breeding*. 2014;34(2):749-760.
 62. Edelstein M, Cohen R, Burger Y *et al.* Integrated Management of Sudden Wilt of Melons, Caused by *Monosporascus cannonballus*, Using Grafting and Reduced Rate of Methyl Bromide. *Plant Dis*. 1999;83:1142-1145.
 63. Hanson PM, Bernacchi D, Green S *et al.* Mapping of a Wild Tomato Introgression Associated with Tomato Yellow Leaf Curl Virus Resistance in a Cultivated Tomato Line. *J Amer Soc Hort Sci*. 2000;125:15-20.
 64. Ji Y, Scott JW, Hanson P *et al.* Sources of Resistance, Inheritance, and Location of Genetic Loci Conferring Resistance to Members of the Tomato Infecting Begomoviruses. In: *Tomato Yellow Leaf Curl Virus Disease: Management, Molecular Biology, Breeding for Resistance* (Ed. Czosnek H). Springer, The Netherlands, 2007,343-362p.
 65. Wang D, Bosland PW. The Genes of Capsicum. *Hort Sci*. 2006;41:1169-1187p.
 66. Van der Biezen EA, Glagotskaya T, Overduin B, Nijkamp HJ, Hille J. Inheritance and genetic mapping of resistance to *Alternaria alternata* f. sp. *lycopersici* in *Lycopersicon pennellii*. *Molecular Genetics and Genomics*. 1995;247:453-461.
 67. Thapa SP, Miyao EM, Davis MR, Coaker G. Identification of QTLs controlling resistance to *Pseudomonas syringae* pv. *tomato* race 1 strains from the wild tomato, *Solanum habrochaites* LA1777. *Theoretical and Applied Genetics*. 2015;128(4):681-92.
 68. Zamir D, Ekstein MI, Zakay Y, Navot N, Zeidan M, Sarfatti M *et al.* Mapping and introgression of a tomato yellow leaf curl virus tolerance gene, Ty1. *Theoretical and Applied Genetics*. 1994;88:141-146.
 69. Finkers R, Bai Y, van den Berg P, van Berloo R, Meijer-Dekens F, ten Have A *et al.* Quantitative resistance to *Botrytis cinerea* from *Solanum neorickii*. *Euphytica*. 2008;159:83-92.

70. Danesh D, Aarons S, McGill GE, Young ND. Genetic dissection of oligogenic resistance to bacterial wilt in tomato. *Molecular Plant-Microbe Interactions*. 1994;7:464-471.
71. Pierce LC. Linkage tests with Ph conditioning resistance to race O, *Phytophthora infestans*. Report of the Tomato Genetics Cooperative. 1971;21:30.
72. Balint-Kurti PJ, Dixon MS, Jones DA, Norcott KA, Jones JDG. RFLP linkage analysis of the Cf-4 and Cf-9 genes for resistance to *Cladosporium fulvum* in tomato. *Theoretical and Applied Genetics*. 1994;88:691-700.
73. Williamson VM, Ho JY, Wu FF, Miller N, Kaloshian I. A PCR-based marker tightly linked to the nematode resistance gene, Mi in tomato. *Theoretical and Applied Genetics*. 1994;87:757-763.
74. Rossi M, Goggin FL, Milligan SB, Kaloshian I, Ullman DE, Williamson VM. The nematode resistance gene Mi of tomato confers resistance against the potato aphid. *Proceedings of National Academy of Sciences USA*. 1998;95:9750-9754.
75. Yaghoobi J, Kaloshian I, Wen Y, Williamson VM. Mapping a new nematode resistance locus in *Lycopersicon peruvianum*. *Theoretical and Applied Genetics*. 1995;91:457-464.