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## Concept and breeding strategies for waterlogging and submerged tolerance in sesame (*Sesamum indicum*)

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### Abstract

Submergence or waterlogging is among the most damaging abiotic stresses in low-lying areas, and agricultural damages due to waterlogging are significant. Plant breeding techniques, either traditional or genetically engineered, may be a productive and cost-effective approach of producing crops that can grow effectively in waterlogged and submerged conditions. Marker assisted selection (MAS) is a novel and much more effective method for identifying genomic areas of crops experiencing stress that could not be identified earlier. The development of complete molecular linkage maps allows us to pyramid favorable characteristics to increase submergence tolerance via MAS. Nevertheless, due to genetic and environmental combination, having too many genes encoding a characteristic, and utilizing unsuitable populations, QTL identification was impeded to maintain adequate growth and production under waterlogged circumstances. Waterlogging responses of soybean and key cereal crops such as rice, barley, maize, and sesame identification of QTL linked to waterlogging tolerance, creation of resistant varieties, and long-term prospects. In environment, such stresses play a significant role in determining the species makeup of an ecosystem. They create economic harm on agricultural land, as well as long-term societal effects. The analysis of various aspects of the stress, especially low stress, has aided knowledge of the plant's biochemical responses to these two stresses. Field crops growth and productivity have slowed due to the harsh weather conditions. Therefore, crop damage and productivity losses are considerable. The morpho-physiological responses of crops during waterlogging, such as the formation of aerenchyma, shoot elongation, adventitious roots, and radial oxygen loss barrier in roots, antioxidant defense system, Anaerobic respiration, role of ethylene, are reviewed in this study.

**Keywords:** Marker assisted selection, waterlogging, submergence, anoxia, ethylene, hypoxia

### 1. Introduction

Sesame (*Sesamum indicum*) is an oil seed crop which is mainly grown for the edible oil purpose. It belongs to family Pedaliaceae and usually called sesamum. It is known for its good quality oil and is familiarly recognized as the "Queen of oilseeds" due to its antibacterial, antiviral properties and many health benefits for mankind. Sesame seeds contains high amount of oil 44-57% and rich source of protein 18-25%, contains carbs 13-14% (Borchani *et al.*, 2010) [10]. The sesame seeds are also known as "the seeds of immortality" because of presence of sesamin, sesamol, sesamol antioxidants. Based on the hull color sesame is two types; White and Black (Verma *et al.*, 2021) [64]. It contains an amino acid Tyrosine which helps in preventing anxiety and depression. To obtain the maximum yield the sesame required 25°C-37°C for its entire growing season (Terefe *et al.*, 2012) [60]. Northeastern India climate is well suited for the production of the sesame crop. According to Directorate of oil seed Development data West Bengal is leading in production and productivity with 2 Lakh MT and 951 kg/ha followed by Madhya Pradesh and Rajasthan (<http://oilseeds.dac.gov.in>). In the world Sesame seed production is high in Sudan with 981K MT. The top producers of sesame in world trend follows Sudan, Myanmar, India, Nigeria, and Tanzania. Major producers of sesame from 1991-2018 are India, Myanmar, and China. Over recent years the African countries becoming more conspicuous in sesame production because of different government initiatives (TREDGE, 2020) [62]. We are in the world where the food demand had been growing day by day and the production or farmland is going on shrinking. So, the researchers are trying new methods and technologies to increase the food production for the coming future generations. Due to ongoing COVID-19 pandemic situation, it made the consumers to reconsider the consumption habits for the healthy lifestyle which have led to the increase in demand for the nutrient rich ingredients. The global sesame seed market is expected grow at

compound annual growth rate (CAGR) of 2.2% around the period of 2021-2026 (Mordor Intelligence). Sesame flourishes well in tough environmental circumstances and requires less amount of water, fertilizers and does not require any pesticides and fungicides because of its elevated tolerance to Pests and Diseases (Myint *et al.*, 2020) [45]. It is greatly tolerant to drought and deficient soil conditions, even though it is readily affected by waterlogging and other stress that leads to the reduced yield and quality of the seeds (Kole, 2019) [33].

Waterlogging is the common problem that occurs in the areas where the annual rainfall is high. Waterlogging depending upon the level of moisture content present in the Field soil, it is also familiar with various designations like Submergence, Soil Humidity, Floods, Hypoxia and Anoxia. Sudan the leading producer of sesame in the world, The losses caused due to the floods in Sudan in 2020 is estimated at 1,00,44,942 tonnes in which sorghum accounts for 50% and sesame accounts for 25% (FAO & GoS, 2020) [22].

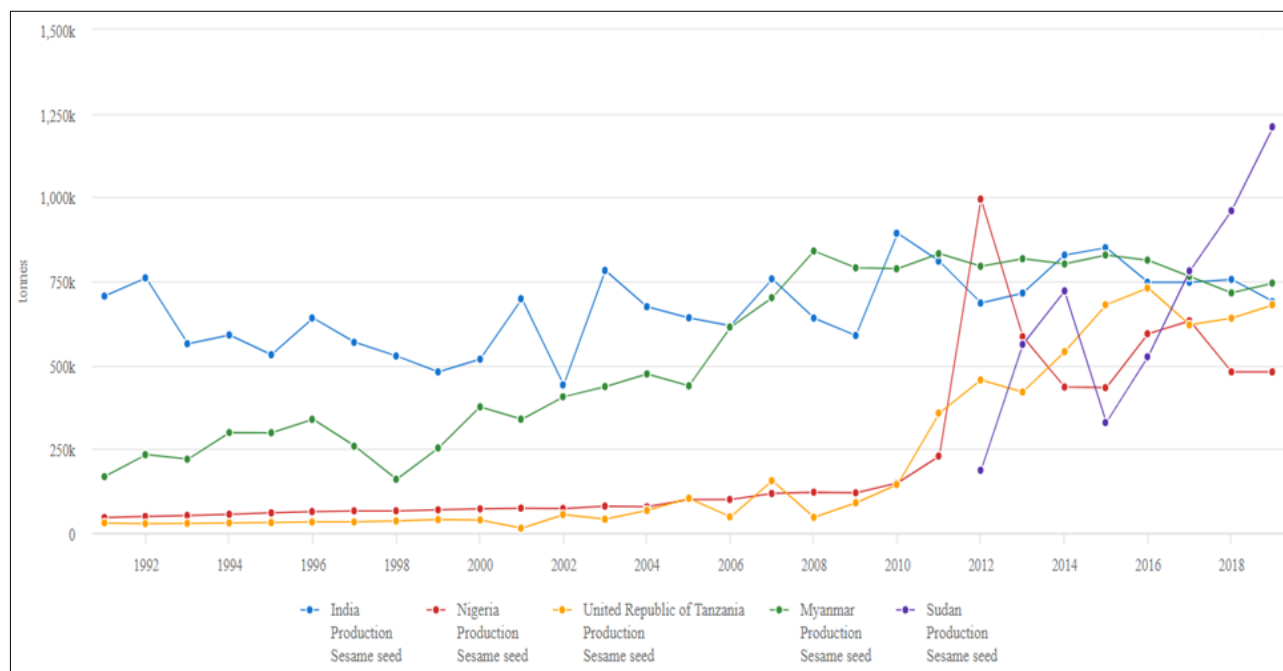


Fig 1: Production tendency of major sesame seed producing countries from 1991-2018 (Source: FAOSTAT Jul 18, 2021) [58]

## 2. What is waterlogging

Waterlogging is the condition that arises due to the stagnation of water in the field. This condition creates diminution of oxygen present in the rhizosphere and reduce the availability of oxygen to the roots of plants. Because, in waterlogged or submerged soils the rate of diffusion of gases is almost 100 times lower in the flooded soil which leads to reduced gaseous exchanges between the root tissue and the atmosphere (Kennedy *et al.*, 1992) [32]. Due to decrease in the oxygen supply the root, shoot growth and the nutrient uptake is reduced (Meyer *et al.*, 1985) [40]. Waterlogging-induced yield losses are aided by a variety of reasons, including changes in soil chemistry and the elements profile, which quickly reduces the redox potential of the soil. In wet soil, denitrification of both inorganic and organic N occurs, severely impeding plant growth and development (Muirhead, 1991) [44]. Hypoxia stress caused by waterlogging encourages soil microorganisms to produce more N<sub>2</sub>O. In respiration, these bacteria utilise nitrate as an alternate electron acceptor. The next electron acceptors are manganese oxides, followed by iron and sulphate. This raises the concentration of soluble Fe<sup>2+</sup> and Mn<sup>2+</sup> in the wet soil, which frequently exceeds the hazardous limit (Marschner, 1991) [38].

### 2.1 Hypoxia and Anoxia

Mesophytes normally cannot tolerate the excess soil moisture for the longer period of time only few mesophytes have the ability to do metabolic function normally under the

waterlogged condition. Hypoxia, the most prevalent form of stress triggered owing to decline of oxygen beneath the optimal concentration in partial submergence for short-term. Anoxia, similar form of stress triggered owing to the complete lack of oxygen in accordance with complete submergence for longer period of time. Prolonged exposure of the plant to hypoxia condition causes wilting and death of the plant (Fukao *et al.*, 2019) [23]. Thus, it is important for the crop to maintain its performance when roots of the crop are exposed to waterlogging and function normally after the recovery from the hypoxia stress period in the waterlogged soil prone regions. It is familiar for waterlogging to have a long-term influence on root development, even after the stress has been removed. (Dickin & Wright, 2008) [17].

## 3. Mechanisms of waterlogging tolerance

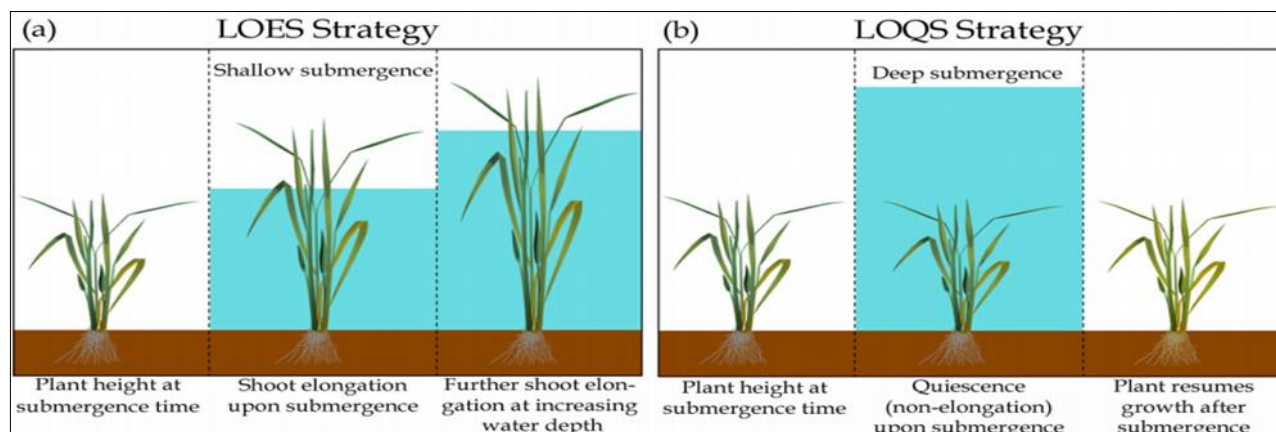
**3.1 Morphological and anatomical alterations:** These are the structural adaptations that are made by the plant during waterlogging condition. Morphological adaptations are helpful only when the plant is in the partial submerged condition.

### 3.1.1 Rapid shoot elongation

Plants belong to some plant species showed a mechanism called low oxygen escape syndrome (LOES) and enables the very survival of the submerged tissues of the plant. Most of the plant species that are grown in waterlogged prone areas have the ability to stimulate elongation of stem or leaves. The

stimulation of fast elongation can help the plant to reach in contact with atmosphere in the partial submerged condition, Due to fast elongation the plant may end up by dying if the energy already store in the plant got depleted before the plant emergence. In complete submergence condition the plant shows Low oxygen quiescence syndrome (LOQS) it uses the carbohydrates to generate energy in the plant for its expenses without shoot elongation. The plant starts growing only after the de-submergence. (Bailey-Serres & Voeselek, 2008) [8].

The submergence tolerant genotype of rice FR13 contain submerged tolerance gene SUB 1A does not elongate the stem in submerged condition, but it starts growing after the de-submergence. Some varieties of indica and japonica showed the elongation of stem under submerged condition to escape the submergence stress (Xu *et al.*, 2006) [70]. Resistance in *Rumex palustris* to submergence is due to swift stem elongation supported by ethylene (Herzog & Pedersen, 2014) [28].



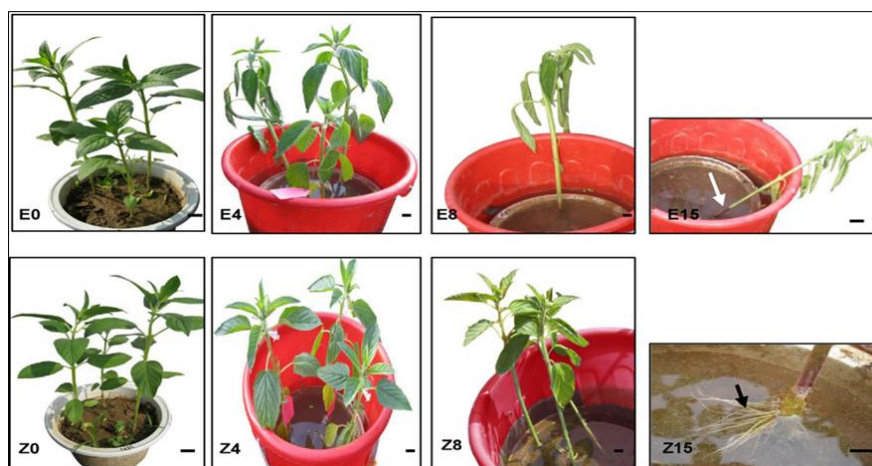
Source: (Striker, 2012) [59]

**Fig 2:** Shallow submergence plant with low oxygen escape syndrome (LOES) and Deep submergence plant with low oxygen quiescence syndrome (LOQS).

### 3.1.2 Adventitious root growth

Adventitious roots are arisen from the stimulus of outgrowth of primordia that are pre existed in the base of the stem (Jackson *et al.*, 1981) [30]. Wei *et al.*, (2013) [68] done experiment in sesame genotypes Ezhi-2 and ZMZ2541, In

Ezhi-2 the older leaves became pale and wilted with in 4 days and the at 15 days the Ezhi-2 endured to 100% death rate but ZMZ2541 leaves were normal until 8 days & by 15 days it developed dynamic AR from the submerged shoot.



**Fig 3:** Morphological responses of both tolerant and intolerant genotypes in Sesame ZMZ2541 (Z) and Ezhi-2 (E). where Z0, Z4, Z8, Z15 & E0, E4, E8, E15 are the day of waterlogging 0, 4, 8, 15. The black arrow in Z15 is indicating the adventitious that are formed by 15 days in ZMZ2541.

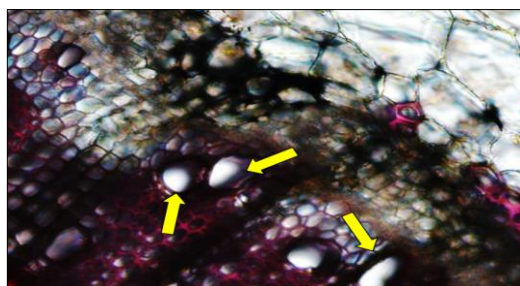
### 3.1.3 Aerenchyma formation

Aerenchyma is a specialized tissue which is formed in many plants in the waterlogged condition it contains constant gas filled channels or spaces. The transfer of O<sub>2</sub> from the shoots to the roots is facilitated by the aerenchyma, which provides a low-resistance internal channel (Drew *et al.*, 1985) [20]. These formed aerenchyma allows the plants survive aerobically to maintain the growth and development of the plant under the hypoxia condition (partially availability of oxygen). However,

some amount of oxygen that is delivered to roots through the aerenchyma gets defused into the rhizosphere that provides oxygen rich environment in the rhizosphere for the growth of microorganisms which in accordance with the conditions prevent the formations of toxic products N<sub>2</sub>O, Fe<sup>2+</sup> and Mn<sup>2+</sup> (Colmer, 2003) [12]. According to M. B. Jackson & Armstrong, (1999) [29], there are two forms of aerenchyma in plants on basis of procedure of development: Schizogenous and Lysigenous. Schizogenous aerenchyma develops species-



specific cell arrangements and occurs regularly in the roots of wetland species without cell demise, while the lysigenous aerenchyma have developed from programmed cell demise and cellwall autolysis to the abiotic stress response. (Evans, 2004) [21]. Aerenchyma is normally formed within 5 to 7 days of waterlogging or hypoxia (Thomson *et al.*, 1990) [61]. In the study conducted by Wei *et al.*, (2013) [68] ZMZ2541 (waterlogging tolerant genotype of sesame) under waterlogging situation roots synthesised the vast lysigenous aerenchyma. Further, they discovered that presence of aerenchymatous cells in the leaf vein epidermis in the ZMZ2541 that are stressed. But, they have witnessed the formed disorganized aerenchyma by the collapse of parenchymatous cells in Ezhi-2 (susceptible genotype of sesame to waterlogging).

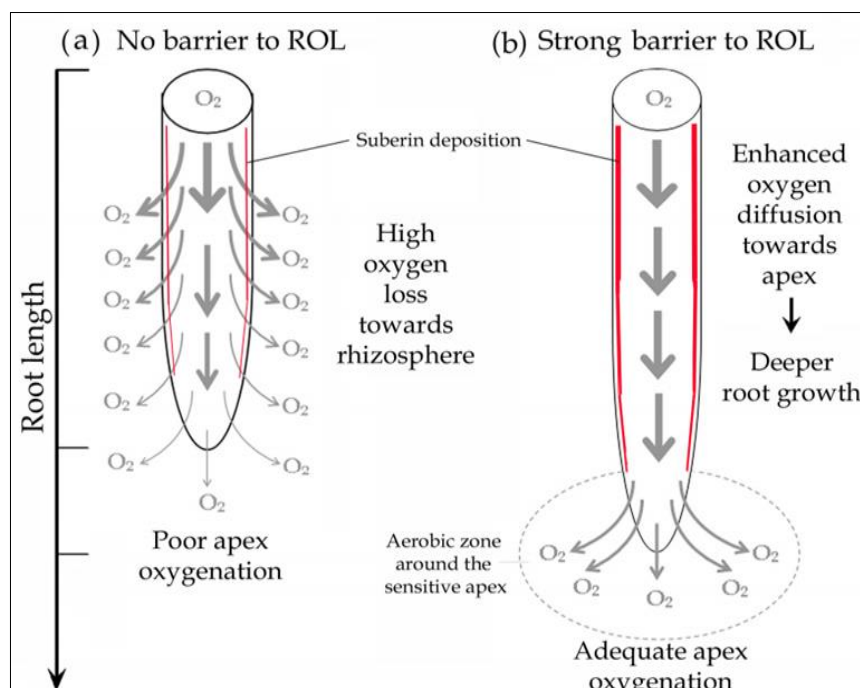


**Fig 4:** Transverse section of sesame plant stem showing formation of aerenchyma. Yellow arrow indicating aerenchyma. In the experiment conducted by (ANEE, 2016) [4] A thin transverse section of sesame stem is made at the waterlogging level and observed under digital microscope had seen the formation of different aerenchyma due to waterlogging.

### 3.1.4 ROL (Radial Oxygen Loss) barrier

ROL is the flux of oxygen from the roots to rhizosphere. In

partial submergence condition some portion of the leaves and shoot are in contact with the atmosphere which makes it easier for the uptake of oxygen. The Oxygen that is absorbed is transported to the roots in waterlogged or hypoxia condition to avoid root anoxia. It is vital for the transport of oxygen to root apex to continue its normal function under submerged condition (Armstrong, 1980) [6]. The  $O_2$  that is transported through the aerenchymas roots to the root tip may be used for metabolic purpose or lost into the rhizosphere is approximately 30-40% through the process of radial diffusion from roots (Yamauchi *et al.*, 2018) [71]. ROL leads to the decrease in the supply of oxygen to the plant root apex and reduce the root growth in the hypoxia or partial submerged condition, But it oxygenate the soil surrounding the roots in the waterlogged condition and helps in the growth of beneficial microorganisms (M. B. Jackson, 1984). Some plants are able to reduce the loss of oxygen to rhizosphere by the formation barrier to ROL (Pedersen *et al.*, 2021) [46]. Colmer, (2003) [12] conducted study on spatial motif of ROL through roots uncovered the barrier is located at the basal region of roots, but no barrier at the root apex. So, even then many wetland plants have strong barrier to ROL, some amount of  $O_2$  is released through the root apex. Release of oxygen into the rhizosphere creates  $O_2$  rich surroundings for roots and it reduces the formation of toxic ions  $Fe^{2+}$  and  $Mn^{2+}$  and sulphides (Armstrong & Armstrong, 2005) [5]. Suberin is the main contender functioning as barrier to  $O_2$  outflow (Shiono *et al.*, 2011; Ranatunga *et al.*, 2011) [57, 48]. Shiono *et al.*, (2011) [57] Based on the histochemical work done by him, suberin deposition is considered more important than the lignin because suberin increased before the changes in lignin. Suberin deposition on the cell wall of outer cortex in submerged condition showed the importance in ROL reduction.



Source: (Striker, 2012) [59].

**Fig 5:** Showing different models of ROL (Radial oxygen loss) from the roots. (a) These are the roots without barrier to ROL in outer cortex. This, results in the loss of oxygen from all positions of the roots caused poor apex  $O_2$ , and short roots under waterlogging condition. (b) These are the roots of species with strong barrier to ROL. Which results in the supply of oxygen to the root apex efficiently and oxygenates the Rhizosphere. Physical barrier to Radial loss of oxygen is due to presence of suberin deposition in the cell wall of exodermis. Suberin is indicated by red line in the (Figure 5) thickness showing the amount of it present and thickness of grey arrows showing the quantity of  $O_2$  available.

Abiko & Miyasaka, (2020) <sup>[1]</sup> had strained adventitious roots of *Colocasia esculenta* with methylene blue they observed that there is no formation of blue coloured area in the middle of roots and blue colour is found in the root tips only. This indicate that there is leakage of oxygen in the root tips and there is no leakage in the other portion of the roots.

### 3.1.5 Metabolic Adaptations

The first metabolic adaptation of the plant to injury and anaerobic conditions is improved by (Crawford, 1978). Plant tissues suffer from the lack of energy due to decrease in the respiration under the hypoxia or anoxia condition in waterlogging intolerant and waterlogging tolerant genotypes (Gibbs and Greenway, 2003). The tolerant plant genotypes can overcome the energy crunch by different metabolic adaptations such as anaerobic respiration, preventing cytoplasmic acidification, Continuous energy metabolism and sugar supply, Development of antioxidants for the survival under anaerobic conditions (A Setter *et al.*, 1997) <sup>[54]</sup>.

### 3.1.6 Anaerobic respiration

In the presence of oxygen, plant cells produce energy by aerobic respiration, which comprises glycolysis, the TCA or Krebs cycle, and oxidative phosphorylation. In the absence of oxygen (anoxia), the Krebs cycle and oxidative phosphorylation are inhibited, and cells must resort to anaerobic respiration to meet their energy needs (Davis, 1980) <sup>[15]</sup>. Glycolysis and fermentation are types of anaerobic respiration. Glycolysis is the main source of energy generation under anaerobic conditions. The regeneration of NAD<sup>+</sup>, a cofactor derived from NADH, is required for the glycolytic process to continue to function (Drew, 1997) <sup>[19]</sup>. To recycle NADH to NAD<sup>+</sup>, large amounts of pyruvate produced as a by-product of glycolysis must be transformed to alternate products. The most significant mechanism for

recycling NADH to NAD<sup>+</sup> under oxygen deficiency is ethanol fermentation or lactate fermentation (Kennedy *et al.*, 1992; Ricard *et al.*, 1994) <sup>[32, 49]</sup>. Pyruvate is the substrate of pyruvate decarboxylase (PDC), which produces CO<sub>2</sub> and acetaldehyde, which is reduced to ethanol by alcohol dehydrogenase oxidising NADH to NAD<sup>+</sup>. In lactic fermentation, pyruvate is the substrate of lactate dehydrogenase (LDH), which converts NADH to NAD<sup>+</sup> to produce lactate. The efficiency of glycolysis and fermentation in producing energy is substantially lower than that of aerobic respiration. Furthermore, the cell is exposed to the end-products of the glycolytic and fermentative pathways, such as ethanol, lactic acid, and CO<sub>2</sub>. The persistence of active glycolysis and the development of fermentative metabolism have been well documented as adaptive processes for plant tolerance to anoxia (Kennedy *et al.*, 1992; Sairam *et al.*, 2008) <sup>[32, 51]</sup>. Anoxia is always preceded by hypoxia in a wet environment (Setter and Waters, 2003), and hypoxia is considered hypoxic pre-treatment prior to exposing the plants to anoxia (Waters *et al.*, 1991). Hypoxia speeds up the induction of glycolytic and fermentative enzymes such as aldolase and enolase, as well as ADH and PDC (Albrecht *et al.*, 2004). This induction can help anoxic plants increase their tolerance to anoxia by improving or at least maintaining their glycolytic rate. On the set of anoxia, anoxic cells may perform lactic fermentation rather than ethanolic fermentation, but ethanol is the less degrading product of fermentation (Davies, 1980) <sup>[15]</sup>. Menegus *et al.*, (1991) <sup>[39]</sup> The increased lactate transport out of the roots and into the surrounding medium, on the other hand, may help to prevent cytoplasmic acidification (Xia and Saglio 1992) <sup>[69]</sup>. Furthermore, decreased cytoplasmic pH causes PDC activation and LDH inhibition (Davis, 1980) <sup>[15]</sup>, resulting in a shift from lactate to ethanolic fermentation.

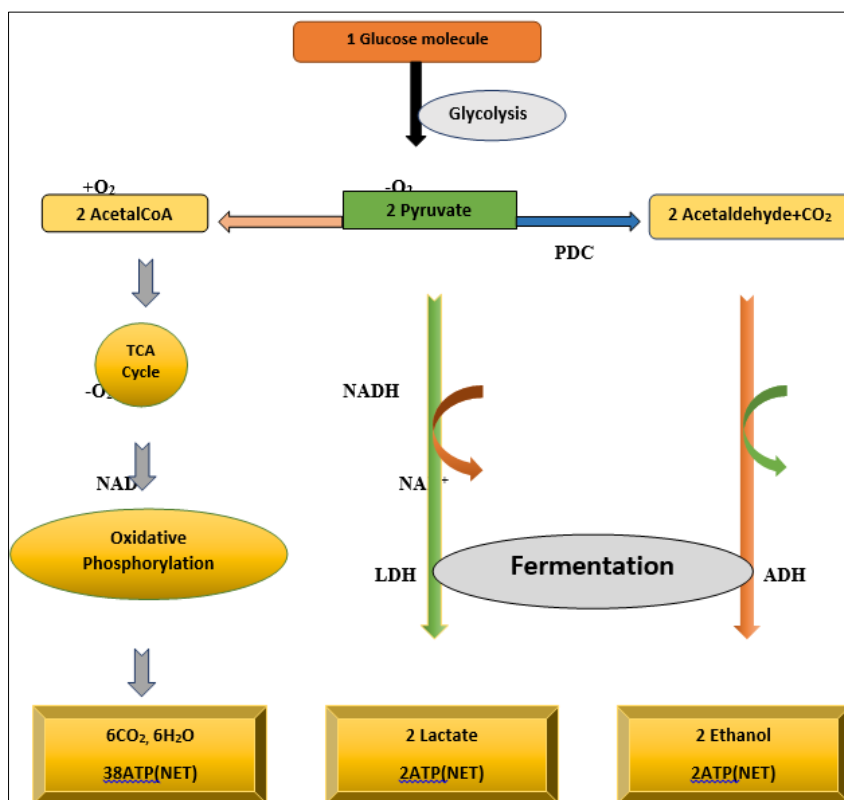


Fig 6: Flow chart of aerobic and anaerobic respiration

### 3.2 Oxidative stress & Antioxidant defence system

Plants are subjected to oxidative stress when they are exposed to most unfavourable situations, such as hypoxia or anoxia, which results in the creation of reactive oxygen species (ROS) such as superoxide radicals, hydroxyl radicals, and hydrogen peroxide, which inhibit plant growth (Mittler *et al.*, 2004) [43]. High concentration of ROS can cause leaf membrane to lipid peroxidation and delipidating, damage to DNA, proteins, which leads to damage of cell membranes and cell organelles (Baxter *et al.*, 2014). Exposure of the plant to normal O<sub>2</sub> condition after a period of O<sub>2</sub> stress can cause serious injury by ROS (Crawford, 1982) [13]. The inability of the scavenging system to metabolise harmful active oxygen due to increased ROS production or decreased scavenging enzyme activity is the most common cause of oxidative stress (Yordanova *et al.*, 2004) [72]. Under waterlogged condition plants had developed certain endogenous antioxidant systems. They are enzymatic and non-enzymatic antioxidants components. Enzymatic components are catalase (CAT), Superoxide dismutase (SOD), Monodehydroascorbate reductase (MDHAR), Peroxidases (POX), Glutathione reductase (GR), Glutathione peroxidase (GPX), Dehydroascorbate reductase (DHAR), Glutathione s-transferase (GST). Non-Enzymatic are Ascorbate (AsA), Glutathione (GSH), Tocopherols, Carotenoids & Pro (Hasanuzzaman *et al.*, 2012) [27]. *Cajanuscajan* when exposed to waterlogging for 6 days showed increases in SOD, GR, APX, CAT activity (Sairam *et al.*, 2009) [53]. *Oryzasativa* exposed to waterlogging for 12 days they observed increase in LDH, ADH, PDC, APX, SOD, CAT & Ethylene (Damanik *et al.*, 2010) [14]. *Zea mays* waterlogged for 10 days showed increase in SOD, POD, APX, CAT, GR activity (Bin *et al.*, 2010) [10]. *Vignaradiata* exposed to waterlogging for 8 days showed an increase in SOD, GR, APX activity (Sairam *et al.*, 2011) [52]. During waterlogging experiments on *Brassica juncea* (L.) seedlings, Lee *et al.* (2014) [35] discovered that a CAT-encoding gene was downregulated, while SOD and POD genes were upregulated. By converting H<sub>2</sub>O<sub>2</sub> to O<sub>2</sub>, CAT may play a role in regulating H<sub>2</sub>O<sub>2</sub> levels. Down-regulation of this gene would result in an increase in H<sub>2</sub>O<sub>2</sub> levels in the leaves of rape seedlings, causing photosynthetic organ damage and premature ageing. Wei *et al.* (2013) [68] conducted studies on *Sesamum indicum* waterlogged sesame crop for 8 days and observed an increase activity of ADH, LDH, PDC, APX, SOD, MDA up to 6 days and then decreased. The information given by different researchers showed that ROS scavenging capacity of different plants can be augmented by increase in antioxidant chemicals, Oxidative damage caused due to waterlogging in the resistance genotypes can be resisted by greater antioxidant activity.

#### 3.2.1 Enhanced accessibility of solvable carbohydrates

The amount of solvable carbohydrates presents in the roots of the plants and the enzymes that can hydrolyse carbohydrates also determines the waterlogging tolerance of a crop (Sairam *et al.*, 2009) [53]. During the waterlogged condition the change of energy use to anaerobic from aerobic under hypoxia or anoxia condition the amount of energy required for the plant tissue is drastically reduced to very little number of ATPs produced from one molecule of glucose. It is consequently critical to maintain a high level of anaerobic metabolic rate in hypoxic or anoxic roots to provide the energy charge required to sustain metabolism in roots for plant survival (Jackson and

Drew, 1984). Zeng *et al.* (1999) [73] observed the two enzymes responsible for the sucrose hydrolysis, the enzyme activity of sucrose invertase (SI) is downregulated and the sucrose synthase is upregulated under hypoxia in maize seedlings. The roots of tolerant varieties showed higher amount of solvable carbohydrates content as compared to susceptible genotypes in pigeon pea. Further, in waterlogged tolerant genotypes it showed an increased activity of sucrose synthase by increase in the expression of mRNA for sucrose synthase, but susceptible varieties showed very low expression under hypoxia (Sairam *et al.*, 2009) [53].

#### 3.2.2 Role of ethylene production under waterlogged condition

Rapid build-up of ethylene in the roots is the most important adaptation under waterlogged condition shown by the tolerant plants. Ethylene is the gaseous hormone and the rate of diffusion of ET under waterlogging is very slow (Hartman *et al.*, 2019) [26].

ACC (1-amino cyclopropane 1-carboxylic acid) is the precursor of the ethylene which increases the production of ethylene in the roots of plants under waterlogging (Bradford K. J. N. D and Yang S. F, 1980) [31]. ACC formation can occur under anaerobic condition, but the formation of ethylene is not possible under anaerobic condition, so, it requires oxygen in the roots for the ethylene production. In the absence of oxygen, the accumulation of ACC increases to the shoot (Alamgir. H *et al.*, 2011) [24]. Ethylene production and the perception are essential for the formation of Adventitious roots. Vidoz *et al.* (2010) [65] conducted an experiment on 4 weeks old tomato seedlings by treating with the waterlogging and above-ground portions were sprayed daily with 500 M aminoethoxy vinyl glycine (AVG), an inhibitor of ET production. As soon as plants waterlogged, plants that were not treated with AVG were submerged for 72 hours, Adventitious root primordia appeared at the base of the stem. These primordia lengthened and produced a high number of ARs within 7 days. Significantly fewer ARs were detected in tomato plants treated with AVG compared to untreated plants. Formation of aerenchyma is normally initiated by the ethylene production (M. B. Jackson & Armstrong, 1999) [29].

### 4. Breeding for water logging tolerance

Breeding methods rely heavily on the presence of wild relatives, elite cultivars, and landraces to allow for the selection of superior lines for improvement (Akbar *et al.*, 2011) [2]. Due to lack of oxygen during and after waterlogging the plants may get affected with root rot and there is decrease in growth and yield. Waterlogging or submerged tolerance is controlled by one or few genes with little effect (Mohanty H. K & Khush G. S, 1985) [25]. Waterlogging tolerant cultivar like Zhongzhi No. 13 are crossed with the distinct genotype for the product of superior varieties with the desired traits (Wang *et al.*, 2016) [66].

#### 4.1 Selection of parents for breeding for water logging tolerance

Sesame genotypes that are more resistant to waterlogging during the vegetative stage were selected by an experiment in pot culture, which examined physiological responses as well as antioxidative enzyme activities. This study compared flooded (during the vegetative stage) and control (no waterlogged) conditions for four sesame genotypes: BD-6980,



BD6985, BD-6992, and BD-70112. Plant height, root volume, root dry, root length weight, and leaf area per plant reduced substantially in all four sesame genotypes under waterlogged conditions as compared to the control condition during vegetative stage. It was shown that waterlogged plants had a higher SPAD (Soil and Plant Analyzer Development) score as well as a higher specific leaf mass compared to controls during the waterlogging phase. All the genotypes exhibited improved root, stem, leaf, and petiole performance when it came to waterlogging tolerance. For all components, BD 6980 demonstrated greater resistance to waterlogging than any other genotype. However, most antioxidant enzyme activities such as Peroxidase (POD), Catalase (CAT), Glutathione peroxidase (GPX), Ascorbate peroxidase (APX) and Superoxide dismutase (SOD) exhibited an increasing tendency in waterlogged plants than that of control plants in all genotypes. It is believed that sesame genotype BD 6980 is extremely resistant to waterlogging because to its low MDA concentration and strong antioxidant activity, whereas the other three genotypes are somewhat tolerant (Saha *et al.*, 2017)<sup>[50]</sup>.

Sesame genotypes for waterlogging tolerance were evaluated by Athul *et al.*, (2017) in Kerala, India. At the seedling stage (20 days after planting), thirty genotypes were screened for waterlogging tolerance by flooding them for 24 hours, 48 hours, and 72 hours. All genotypes survived 24 and 48 hours of waterlogging, however only 17 genotypes survived 72 hours. Observations of genotypes that survived 72 hours of flooding were recorded, and the findings revealed that the genotypes differed considerably for the characteristics under investigation. The ten genotypes with the best survival percentages were chosen for the field experiment. Ayali, Sesamummalabaricum, SC 207, Thilarani, Thilak, GT 10, SV 2, TKG 308, TKG 22 and Rama. Flooding was forced in the field experiment for 72 hours, and biometrical characteristics were collected and statistically analysed. The genotypes indicated substantial differences in all the characteristics. Ayali, a native variety, had the largest yield per plant (7.46g) while Sesamummalabaricum had the lowest (2.92g). Thilak (48.6 percent) had the greatest oil content, while Sesamummalabaricum, a wild species, had the lowest (32.5 percent).

#### 4.2 MAS (marker assisted selection) breeding for waterlogging tolerance

Since the mid-1990s, key genes important for waterlogging tolerance have been identified, making it easier for researchers to focus on modification or use. Use of such genes to create a new waterlogged-tolerant crop Rice, maize, wheat, barley, soybeans and other grains are examples. In the instance of rice, the Sub1 gene was introgressed to a specific genotype, markers aided backcrossing (MAB) types for various Land kinds, farmer preferences, and the addition of new genotypes it became feasible to create new kinds through genetic engineering.

For genetic diversity research, molecular marker methods such as random amplified polymorphic DNA (RAPD), simple sequence repeats (SSR), amplified fragment length polymorphism (AFLP), and inter simple sequence repeats (ISSR) are often utilised (Laurentin & Karlovsky, 2006)<sup>[34]</sup>. These are the QTL for waterlogging that are identified in the mid-1990s. In Rice (*Oryzasativa* L.) the Sub1 in chromosome number (Ch. 9) is identified by using the SSR marker by

marker assisted selection method (Xu *et al.*, 2006)<sup>[70]</sup>. In Barley (*Hordeumvulgare* L.) tfy2.1-1, tfy1.1-2, tfy1.2-1 are identified by using RFLP marker by MAS (H. Li *et al.*, 2008). In Maize (L.) the gene for waterlogging is identified on Ch. 4, 9 by using SSR marker by Composite interval mapping (CIM) method (Qiu *et al.*, 2007)<sup>[47]</sup>. Sesame is quite sensitive to waterlogging stress. After 2–3 days of waterlogging, the crop's development and yield are reduced, which is common when they are cultivated on poorly drained soils (Ucan *et al.*, 2007)<sup>[63]</sup>. Wang *et al.*, (2012)<sup>[67]</sup> discovered 13,307 DEGs (differentially expressed genes) for sesame during waterlogging stress. A larger investigation discovered a total of 1,379 genes as the main genes that acts in relation to waterlogging. They found 66 genes that might be candidates for increasing sesame resistance to waterlogging (Wang *et al.*, 2016)<sup>[66]</sup>. Simultaneously, six QTLs (qEZ09ZCL13, qEZ10ZCL07, qWH09CHL15, qEZ10CHL07, and qWH10CHL09, qWH10ZCL09) which were related to waterlogging characteristics, and an SSR marker (ZM428) strongly linked to qWH10CHL09 was revealed as an efficacious marker for marker-assisted selection (MAS) against waterlogging tolerance (Zhang *et al.*, 2014)<sup>[75]</sup>.

#### 4.3 Waterlogging tolerance through genetic engineering

Manipulation of a specific gene in plant for increasing the production or tolerance to a particular stress is becoming necessary day by day (Lemay & Moineau, 2020)<sup>[36]</sup>. The production of agronomical and horticulture crops against the climatic stress is becoming difficult, genome editing can help in decreasing the environmental effect in the production (Lemay & Moineau, 2020)<sup>[36]</sup>. Through genetic engineering we can increase or decrease the expression of endogenous genes. The development of editing technology has ushered in a new age of discoveries in a variety of biotechnology sectors, including agricultural investigation (Doudna and Charpentier, 2014)<sup>[18]</sup>. Nevertheless, there is one exception. There has been one documented case of gene manipulation in Stress caused by submergence/waterlogging. Yamauchi *et al.* (2017) modified truncated rice respiratory burst oxidase homolog (RBOHH) genes to highlight the critical function that peroxide plays. This enzyme's output influences waterlogging signalling. and the development of aerenchyma. Unfortunately, homozygous plants found to be infertile, preventing further phenotyping. A new study presents an intriguing paradigm for genome editing for plant biotechnology applications. Miao *et al.* (2018)<sup>[41]</sup> used Cas9 to generate mutants for all Pyrabactin resistance 1 (PYL) ABA receptors in rice and tested all combos for biotechnologically interesting genotypes. Among all conceivable combinations, pyl1/4/6 boosted growth and productivity by being free of ABA-related actual growth limitations while retaining ABA-controlled positive traits such as undesired seed sprouting, which are found in several other mutant combos. They believe that the gathered understanding of submergence/flooding stress is advanced enough to apply these types of strong gene and genome editing methods with many of its regulating branches, particularly the complicated network of TF groups. These potential innovation methods can generate variants ranging from single base editing to gene fusions and replacements and maybe even transgene-free genomes (Miki *et al.*, 2018; Chen *et al.*, 2018; Zhang Y. *et al.*, 2016)<sup>[42, 11, 74]</sup>. Great number of studies have shown that WRKY gene families are expressed

sturdily with the abiotic stresses, drought, salt stress and waterlogging (Diao *et al.*, 2016) [16]. To discover sesame WRKY proteins, all Arabidopsis WRKY candidate genes were utilised as query in the Basic Local Alignment Search Tool (BLAST). There were 61 putative WRKY genes discovered in all, and anticipated protein sequences lacking a WRKY domain were excluded. Twelve differential expression SiWRKY genes were chosen, and their expression levels measured by qRT-PCR at different intervals following the beginning of each abiotic stress to validate the identification of some of the genes crucial for waterlogging and drought resistance. For both the waterlogging-tolerant and -sensitive cultivars, six SiWRKY genes were transcribed at various periods after the commencement of the waterlogging treatment ( $P < 0.05$ ). During the waterlogging treatment, the expression levels of SiWRKY13, SiWRKY35, and SiWRKY43 rose, albeit every gene's expression spiked at a different period. SiWRKY35 reached its maximal expression before SiWRKY13 and SiWRKY43. Waterlogging, on the other hand, reduced the expression of SiWRKY17, SiWRKY59, and SiWRKY63. In the study conducted by them the variable expression patterns of SiWRKY genes in various cultivar tissues revealed that these genes play distinct functions in sesame growth and many display tissue-specific transcriptional pattern. Furthermore, SiWRKY gene expression studies indicated that some were significantly elevated or negatively regulated in relation to waterlogging and drought stress. These findings will aid future research into the roles of WRKY genes involved in abiotic stress responses, as well as the creation of molecular breeding strategies to improve abiotic stress tolerance in sesame (D. Li *et al.* 2017) [37].

## 5. Conclusion

In this review we had understood completely about the waterlogging and how it is affecting the crop production especially in the sesame. By this we can conclude that the waterlogging is the major problem that occurring in the sesame crop. Waterlogging tolerance is every important for the sustainability of the crop in the frequently waterlogged areas. Phenotypic based Plant breeding techniques is time consuming, and it have lot of other problems as we seen, and it is not enough for developing a plant that are tolerant to waterlogging in a short period of time. Mapping of QTLs for waterlogging tolerance is very much necessary for the identification of small effect contributed by many genes and should use techniques like MAS which increases the precision for the selection for the screening of plants in a population which is simple and precise as compared to the phenotypic selection.

## 6. References

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