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## Effect of plant growth regulators on growth and physiological parameters of wheat (*Triticum aestivum* L.) under late sown condition

Mayanker Singh, Kushwaha SP, Shikha Shahi and Abhishek Pati Tiwari

### Abstract

The present study entitled “Effect of plant growth regulators on growth and physiological parameters of wheat (*Triticum aestivum* L.) Under late sown condition” has been taken to focus the involvement and potential role of plant growth regulator and was carried out in during 2019-20 and 2020-21 at Agronomy Research Farm of C.S.A. University of Agriculture and Technology, Kanpur (U.P.). The experiment was done during Rabi (2019-20 and 2020-21) using wheat variety K-9423 using randomized block design having eleven treatments and three replications having thirty-three plots different plant growth regulators are used at various growth stages for overall development of crop. The details of the following treatments used for two years of experimentation is as follows T<sub>1</sub>: Control, T<sub>2</sub>: IAA, 25 ppm T<sub>3</sub>: IAA, 50 ppm T<sub>4</sub>: Abscisic acid 10 ppm, T<sub>5</sub>: Abscisic acid 20 ppm, T<sub>6</sub>: TRIA 5 ppm, T<sub>7</sub>: TRIA 10 ppm, T<sub>8</sub>: Brassinosteroid 5 ppm, T<sub>9</sub>: Brassinosteroid 10 ppm, T<sub>10</sub>: Cytokinin 5 ppm, T<sub>11</sub>: Cytokinin 10 ppm. Foliar spraying at two stages - first at 30 DAS (Vegetative stage) and second at anthesis in uniform dose of all treatments. The maximum Relative water content (RWC) (%) at heading stage in wheat was noted in T<sub>3</sub> with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> and was statistically at par with T<sub>2</sub> IAA 25 ppm and T<sub>10</sub> Cytokinin 5 ppm. Minimum Relative water content (RWC) (%) at heading stage was observed under T<sub>5</sub> Abscisic acid 20 ppm during both the years of investigation. The maximum Protein content in grain (%) in wheat was found in T<sub>3</sub> with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> and was statistically at par with T<sub>2</sub> IAA 25 ppm and T<sub>10</sub> Cytokinin 5 ppm. Least Protein content in grain (%) was observed under T<sub>5</sub> Abscisic acid 20 ppm during both the years of investigation.

**Keywords:** RWC, IAA, *Triticum aestivum* L., physiological parameters, treatments

### Introduction

Late sowing of wheat is a major problem in the rice-wheat (Hobbs and Gupta, 2002)<sup>[58]</sup> and cotton-wheat (Khan *et al.*, 2010)<sup>[48]</sup> areas of Asia. Regmi *et al.* (2002)<sup>[52]</sup> also reported a yield decline in wheat when it was sown after the third week of November. A major reason for late sowing is the late harvest of the preceding crops. The inputs applied to the wheat crop were not efficiently utilized and resulted in reduced yield under late sowing (Hobbs and Gupta, 2002)<sup>[58]</sup>. In late sown wheat, all the growth stages, such as tillering, flowering and grain filling are adversely affected by the shortened growing period. The reduction in the optimum growth period caused by a rise in temperature leads to leaf senescence resulting in a photosynthetic rate that is too low to meet plant C economy (Hensel, *et al.* 1993; Sharma-Natu, *et al.* 2006)<sup>[61, 59]</sup>. As a result, it affects two important yield parameters, i.e., the number of grains per spike and grain weight (Ugarte, *et al.* 2007)<sup>[60]</sup>. Each day delay in sowing of wheat after 20th November onward in decreases grain yield at the rate of 36 kg ha<sup>-1</sup> day<sup>-1</sup> (Hussain, *et al.* 1998)<sup>[63]</sup>. To meet the increased demand for food grain of rapidly growing population, there are many yields boosting agronomic techniques like application of certain plant growth regulators which needs due attention. Although plant growth regulators have been used in agriculture for as long as crop cultivation their impact up to now has been relatively little detected and their application is limited to some specific objectives for example quality and quantity improvement (Pandey *et al.* 2011)<sup>[62]</sup>.

Plant growth regulators (PGRs) are known to influence a wide array of physiological processes like photosynthesis (Jia and Lu 2003)<sup>[53]</sup> and stomata conductance (Mori *et al.*, 2006)<sup>[56]</sup>. PGRs are reported to modify growth and development in various ways under different growth conditions (Pan *et al.*, 2013)<sup>[65]</sup> and influence the crop yield (Yan *et al.* 2011)<sup>[55]</sup>.

The importance of biologically active substances in plants is well documented. Plant growth regulators are natural compounds that have shown far reaching effects on the growth and development of plants even at low concentration (Arshad and Franken Berger, 1998) [57]. Plant growth regulators are known to affect growth, flowering and assimilate translocation in plants (Hayat *et al.*, 2007 [24]. Studies have shown that plant hormones could regulate the partitioning and translocation of photo assimilates during grain filling (Ahmadi and Baker, 1999) [34]. Both 3-indoleacetic acid (IAA) and cytokinins have been shown to play an important role in the transportation of assimilates to wheat spikes (Darussalam *et al.*, 1998 and Lejeune *et al.*, 1998) [45, 46]. As well, cytokinins are required for cell division during the early phase of grain filling (Yang *et al.*, 2000) [33]. Plant growth regulators impress production of plant hormones and are thus able to modify plant growth and development, this being a regulatory rather than a nutritional role. They are generally only applied at low rates without showing phytotoxic effects (Rademacher and Brahm, 2010) [3]. Foliar application of plant growth regulators, both natural and synthetic, has proven worthwhile for improving crop growth against a variety of abiotic stresses. Remarkable accomplishments of plant growth regulators, such as manipulating plant developments, enhancing yield and quality have been actualized in recent years using new emerging and efficient plant growth regulators. It has long been ascertained that plant hormones including auxins, gibberellins, cytokinin and ethylene etc., in some studies with wheat, characteristics associated with stem strength have increased with application of PGRs, in particular stem diameter stem wall thickness (Tripathi *et al.*, 2003) [64]. Given the potential influence on yield components from productive tillers to grains/spike and then grain weight, timing of the PGR should also progressively affect these yield components. Disparity in the chemical nature and mode of action of PGRs result in variation in response to application. Uptake sites vary; while most PGRs are mostly absorbed through leaves and stems (Kim *et al.*, 2010) [10]. Reduction in plant biomass due to higher PGRs may also have negative consequences for grain yield through reduced photosynthetic area and lower levels of stored reserves for re-translocation at grain filling time of wheat (Espindula *et al.*, 2009) [47]. Other crop management inputs such as irrigation, nitrogen status and earlier time of sowing have also influence in greater yield response to PGR treatment.

## Materials and Methods

In order to accomplish the objectives of the present study entitled “Effect of plant growth regulators on morpho-physiological parameters and yield of wheat (*Triticum aestivum* L.) Under late sown condition” was carried out during Rabi seasons of 2018-19 and 2019-20.

The present investigation entitled “Effect of plant growth regulators on morpho-physiological parameters and yield of wheat (*Triticum aestivum* L.) Under late sown condition” has been taken to focus the involvement and potential role of plant growth regulator and was carried out in during 2019-20 and 2020-21 at Agronomy Research Farm of C.S.A. University of Agriculture and Technology, Kanpur (U.P.).

## Results and Discussion

### Number of tillers per plant

The data pertaining to number of tillers per plant presented in

the (Table-4.2) revealed that the maximum number of tillers per plant at 30 days in wheat was observed in T<sub>3</sub> (2.99) and (3.11) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (2.88), (2.89) which was statistically at par with T<sub>2</sub> (2.80), (2.86), T<sub>10</sub> (2.66), (2.71), T<sub>9</sub> (2.60), (2.66), T<sub>8</sub> (2.61), (2.64). Least number of tillers per plant was Observed under T<sub>5</sub> (2.36) and (2.41) during both the years of investigation.

The maximum number of tillers per plant at 60 days in wheat was observed in T<sub>3</sub> (6.22), (6.32) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (5.66) and (5.80) which was statistically at par with T<sub>2</sub> (5.66), (5.70), T<sub>10</sub> (5.60), (5.64), T<sub>9</sub> (5.40), (5.44), T<sub>8</sub> (5.11), (5.20). Least number of tillers per plant was Observed under T<sub>5</sub> (4.22) and (4.33) during both the years of investigation.

The maximum number of tillers per plant at 90 days in wheat was observed in T<sub>3</sub> (6.99), (7.11) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (6.89), (7.00) which was statistically at par with T<sub>2</sub> (6.33), (6.40), T<sub>10</sub> (6.35), (6.39), T<sub>9</sub> (6.05), (6.10), T<sub>8</sub> (5.90), (5.98). Least number of tillers per plant was Observed under T<sub>5</sub> (5.38) and (5.43) during both the years of investigation.

The maximum number of tillers per plant at harvest days in wheat was observed in T<sub>3</sub> (7.77) and (7.89) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (7.44), (7.56) which was statistically at par with T<sub>2</sub> (7.22) and (7.33), T<sub>10</sub> (6.44), (6.56), T<sub>9</sub> (6.34), (6.45), T<sub>8</sub> (6.00), (6.11). Least number of tillers per plant was Observed under T<sub>5</sub> (5.51) and (5.55) during both the years of investigation. Kumar and Sharma (2003) [32] reported that sowing time significantly affects the number of tillers per meter row length. The crop sown on 30th November (105.0) produced significantly higher number of tillers per meter row length than the crop sown on 16 December (94.1) and 31 December (94.1); Later dates of sowing did not differ significantly. Negi *et al.* (2003) [35] reported that sowing dates significantly affects the effective tillers. Significantly higher number of effective tillers m<sup>-2</sup> was recorded in crop sown on 28 November (245.6) than the crop sown on 28 October (186.8). Dhaka *et al.* (2006) [38] found in pooled analysis of two years of study, that the numbers of tillers per plant were reduced by 4.8% when wheat was sown on 25 December (3.9 tillers/plant) instead of 20 November (4.1 tillers/plant), but this difference was not statistically significant. Shah *et al.* (2006) [37] carried out a field experiment at Peshawar (Pakistan) and reported that sowing on 1st November resulted in higher seedling emergence (179.29 seedling per meter square). Shahzad *et al.* (2007) [39] from Pakistan reported that the crop sown on December 15 produced significantly a greater number of fertile tillers (m<sup>-2</sup>) (499.77) than the crop sown on November, 15 (475.33) and 30 (462.88). Singh *et al.* (2011) [40] found that crop sown on 15th October produced higher grain yield as compared to crop sown on 25th October, 4th and 14th November. The increase in yield was due to increase in effective tillers and 1000-grain weight of early sown crop. Hussain and Leitch (2007) [49] studied that early application of PGRs has resulted in increased tiller population compared with later applications.

### Number of leaves per plant

The data pertaining to number of leaves per plant presented in the (Table-4.3) revealed that the maximum number of leaves per plant at 30 days in wheat was observed in T<sub>3</sub> (7.11) and

(7.25) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (6.88), (6.91) which was statistically at par with T<sub>2</sub> (6.77) and (6.76), T<sub>10</sub> (6.61) and (6.69), T<sub>9</sub> (6.55), (6.60), T<sub>8</sub> (6.49) and (6.54). Least number of leaves per plant was Observed under T<sub>5</sub> (6.10) and (6.22) during both the years of investigation.

The maximum number of leaves per plant at 60 days in wheat was observed in T<sub>3</sub> (23.50) and (23.80) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (23.10) and (23.18) which was statistically at par with T<sub>2</sub> (22.65), (22.87), T<sub>10</sub> (22.45), (22.73), T<sub>9</sub> (22.33), (22.67), T<sub>8</sub> (22.05) and (22.41). Least number of leaves per plant was Observed under T<sub>5</sub> (20.11) and (20.38) during both the years of investigation.

The maximum number of leaves per plant at 90 days in wheat was observed in T<sub>3</sub> (24.51) and (24.94) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (24.11) and (24.40) which was statistically at par with T<sub>2</sub> (23.00), (23.98), T<sub>10</sub> (23.51), (23.90), T<sub>9</sub> (22.96), (23.33), T<sub>8</sub> (22.70) and (22.91). Least number of leaves per plant was Observed under T<sub>5</sub> (5.38) and (5.43) during both the years of investigation. Patil and Subbanna (1988) [51] conducted field experiment at UAS, Bangalore reported that triacontanol sprayed in varying concentrations significantly increased the number of leaves per plant, leaf area, plant spread, fresh weight of untrimmed lettuce heads wherein triacontanol at 4 ppm twice recorded the highest number of leaves per plant (22.5), leaf area (3698.8 cm<sup>2</sup>), plant spread (1112.2 cm<sup>2</sup>) and fresh weight of untrimmed heads (341 g) while the least values were obtained in control treatment.

#### Leaf area per plant

The data pertaining to leaf area per plant presented in the (Table-4.4) revealed that the maximum leaf area per plant at 30 days in wheat was observed in T<sub>3</sub> (50.50) and (50.72) with application of IAA 50 Ppm which was significantly superior over T<sub>11</sub> (50.20) and (50.35) which was statistically at par with T<sub>2</sub> (50.05), (50.27), T<sub>10</sub> (49.98), (50.20), T<sub>9</sub> (49.91), (50.05), T<sub>8</sub> (49.76), (49.91). Least leaf area per plant was Observed under T<sub>5</sub> (49.43) and (49.74) during both the years of investigation.

The maximum leaf area per plant at 60 days in wheat was observed in T<sub>3</sub> (513.65) and (514.31) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (496.85) and (497.74) which was statistically at par with T<sub>2</sub> (502.82) and (503.92), T<sub>10</sub> (488.90), (489.78), T<sub>9</sub> (486.02), (488.01), T<sub>8</sub> (482.27) and (483.37). Least leaf area per plant was Observed under T<sub>5</sub> (465.91) and (466.57) during both the years of investigation.

The maximum leaf area per plant at 90 days in wheat was observed in T<sub>3</sub> (748.14) and (750.10) with application of IAA 50 PPM which was significantly superior over T<sub>11</sub> (742.00) and (742.30) which was statistically at par with T<sub>2</sub> (732.22), (745.10), T<sub>10</sub> (738.20), (738.60), T<sub>9</sub> (736.70), (737.20), T<sub>8</sub> (732.80) and (733.30). Least leaf area per plant was Observed under T<sub>5</sub> (725.10) and (726.50) during both the years of investigation. Meyer and Green (1980) found that the amount of photosynthate available for biomass production is related to the current leaf area and the photosynthetic rate of the crop in consideration.

#### Days to Heading

The data pertaining to days to heading presented in the (Table-4.4) revealed that the maximum days to heading in

wheat was observed in T<sub>3</sub> (77.11) and (78.34) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (71.66) and (72.92) which was statistically at par with T<sub>2</sub> (77.33), (77.67), T<sub>10</sub> (73.33), (74.81), T<sub>9</sub> (72.66), (75.38) and T<sub>8</sub> (72.16) and (72.46). Least days to heading was Observed under T<sub>5</sub> (76.33) and (76.55) during both the years of investigation. Meena (2009) [50] observed that number of days taken to physiological maturity and days to heading was significantly affected by sowing dates. The November 14 sown crop took significantly higher number of days to attain maturity and heading as compared to December 4 and December 24 sown crops.

#### Days to Maturity

The data pertaining to days to maturity presented in the (Table-4.4) revealed that the maximum days to maturity in wheat was observed in T<sub>3</sub> (126.00) and (127.00) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (119.00) and (119.15) which was statistically at par with T<sub>2</sub> (125.00), (126.15), T<sub>10</sub> (120.00), (120.30), T<sub>9</sub> (120.60), (121.00), T<sub>8</sub> (121.33) and (121.66). Least days to maturity was Observed under T<sub>5</sub> (121.33) and (121.66) during both the years of investigation. Meena (2009) [50] observed that number of days taken to physiological maturity and days to heading was significantly affected by sowing dates. The November 14 sown crop took significantly higher number of days to attain maturity and heading as compared to December 4 and December 24 sown crops.

#### Dry matter accumulation

The data pertaining to dry matter accumulation presented in the (Table-4.5) revealed that the maximum dry matter accumulation at 30 days in wheat was observed in T<sub>3</sub> (2.41) and (2.43) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (2.41) and (2.42) which was statistically at par with T<sub>2</sub> (2.40), (2.41), T<sub>10</sub> (2.40), (2.41), T<sub>9</sub> (2.38), (2.39), T<sub>8</sub> (2.36), (2.37). Least dry matter accumulation was Observed under T<sub>5</sub> (2.39) and (2.41) during both the years of investigation.

The maximum dry matter accumulation at 60 days in wheat was observed in T<sub>3</sub> (10.79) and (10.88) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (10.58), (10.76) which was statistically at par with T<sub>2</sub> (10.44) and (10.53), T<sub>10</sub> (10.32), (10.40), T<sub>9</sub> (9.91) and (9.99) and T<sub>8</sub> (9.79) and (9.87). Least dry matter accumulation was Observed under T<sub>5</sub> (10.23) and (10.35) during both the years of investigation.

The maximum dry matter accumulation at 90 days in wheat was observed in T<sub>3</sub> (21.32) and (21.55) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (20.90) and (21.30) which was statistically at par with T<sub>2</sub> (20.62) and (20.84), T<sub>10</sub> (20.37) and (20.57), T<sub>9</sub> (19.42) and (19.71) and T<sub>8</sub> (19.17) and (19.36). Least dry matter accumulation was Observed under T<sub>5</sub> (20.17) and (20.40) during both the years of investigation.

The maximum dry matter accumulation at harvest in wheat was observed in T<sub>3</sub> (29.10) and (29.45) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (28.50) and (29.10) which was statistically at par with T<sub>2</sub> (27.76), (28.12), T<sub>10</sub> (27.32), (27.35), T<sub>9</sub> (25.90), (26.35), T<sub>8</sub> (25.55) and (25.86). Least dry matter accumulation was Observed under T<sub>5</sub> (26.97) and (27.36) during both the years of investigation SC and Luan S (2012) reported that



application of ABA enhanced GB accumulation, RWC and dry matter production in maize plants.

### Leaf Area Index (LAI)

The data pertaining to leaf area index presented in the (Table-4.6) revealed that the maximum leaf area index at 30 days in wheat was observed in T<sub>3</sub> (0.26) and (0.28) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (0.25) and (0.27) which was statistically at par with T<sub>2</sub> (0.25) and (0.26), T<sub>10</sub> (0.25) and (0.26), T<sub>9</sub> (0.25) and (0.26) and T<sub>8</sub> (0.25) and (0.26). Least leaf area index was Observed under T<sub>5</sub> (0.24) and (0.25) during both the years of investigation.

The maximum leaf area index at 60 days in wheat was observed in T<sub>3</sub> (2.38) and (2.40) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (2.33) and (2.35) which was statistically at par with T<sub>2</sub> (2.30) and (2.32), T<sub>10</sub> (2.26) and (2.28), T<sub>9</sub> (2.25) and (2.27) and T<sub>8</sub> (2.23) and (2.25). Least leaf area index was Observed under T<sub>5</sub> (2.05) and (2.13) during both the years of investigation.

The maximum leaf area index at 90 days in wheat was observed in T<sub>3</sub> (3.46) and (3.48) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (3.39) and (3.42) which was statistically at par with T<sub>2</sub> (3.33) and (3.36), T<sub>10</sub> (3.27) and (3.30), T<sub>9</sub> (3.26) and (3.29) and T<sub>8</sub> (3.24) and (2.28). Least leaf area index was Observed under T<sub>5</sub> (3.11) and (3.17) during both the years of investigation.

The maximum leaf area index at harvest in wheat was observed in T<sub>3</sub> (7.89) and (7.11) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (7.56) and (6.88) which was statistically at par with T<sub>2</sub> (7.33) and (6.77), T<sub>10</sub> (6.56) and (6.61), T<sub>9</sub> (6.45) and (6.55) and T<sub>8</sub> (6.11) and (6.49). Least leaf area index was Observed under T<sub>5</sub> (5.55) and (6.10) during both the years of investigation. Aldesuquy (2000) reported that the significance of leaf area increments in higher grain production have been discussed and emphasized from time to time where the higher leaf area index was found to be positively correlated with higher grain yield. In the discussions, it can be said that higher leaf area implies more because the sink source relationship increases with increase in photosynthates and hence is reflected to grains hence more grain yield recorded.

### Physiological Parameters

#### Relative Growth Rate (RGR)

The data pertaining to relative growth rate presented in the (Table-4.7) revealed that the maximum relative growth rate at 30 days in wheat was observed in T<sub>3</sub> (49.96) and (51.80) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (49.31) and (49.46) which was statistically at par with T<sub>2</sub> (49.00), (49.15), T<sub>10</sub> (48.62), (48.71), T<sub>9</sub>(47.54) and (47.67) and T<sub>8</sub> (47.42) and (47.55). Least relative growth rate was Observed under T<sub>5</sub> (48.47) and (48.57) during both the years of investigation.

The maximum relative growth rate at 60 days in wheat was observed in T<sub>3</sub> (22.70) and (22.78) with application of IAA 50 PPM which was significantly superior over T<sub>11</sub> (22.69) and (22.76) which was statistically at par with T<sub>2</sub> (22.68) and (22.75), T<sub>10</sub> (22.66) and (22.73), T<sub>9</sub> (22.42) and (22.48) and T<sub>8</sub> (22.40) and (22.46). Least relative growth rate was Observed under T<sub>5</sub> (22.62) and (22.71) during both the years of investigation.

The maximum relative growth rate at 90 days in wheat was observed in T<sub>3</sub> (10.37) and (10.44) with application of IAA 50

ppm which was significantly superior over T<sub>11</sub> (10.34) and (10.40) which was statistically at par with T<sub>2</sub> (9.91) and (10.01), T<sub>10</sub> (9.80) and (9.98), T<sub>9</sub> (9.58) and (9.67) and T<sub>8</sub> (9.57) and (9.64). Least relative growth rate was Observed under T<sub>5</sub> (9.68) and (9.78) during both the years of investigation. Wahid *et al.* (2007) <sup>[37]</sup> observed in pearl millet that net assimilation rate, plants relative growth rate as well as shoot dry mass get considerably reduced due to elevated temperatures. Elevated temperature hastens phenological development in wheat and influence grain growth and yield. Increase in atmospheric temperature can disturb plant productivity since duration and rate of grain filling together are controlled by change in temperature. Enhancement in rate of grain development and shortening of grain filling period has been observed in several crops

#### Canopy Temperature depression (CTD) Pre and post anthesis

The maximum Canopy Temperature depression (CTD) Pre anthesis in wheat was observed in T<sub>3</sub> (5.05) and (5.16) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (4.76) and (4.81) which was statistically at par with T<sub>2</sub> (4.61) and (4.68), T<sub>10</sub> (4.55) and (4.63), T<sub>9</sub> (4.33) and (4.40) and T<sub>8</sub> (4.24) and (4.32). Least Canopy Temperature depression (CTD) Pre anthesis was Observed under T<sub>5</sub> (4.05) and (4.11) during both the years of investigation.

The maximum Canopy Temperature depression (CTD) Post anthesis in wheat was observed in T<sub>3</sub> (6.75) and (6.83) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (6.15) and (6.22) which was statistically at par with T<sub>2</sub> (5.93), (6.01), T<sub>10</sub> (5.76), (5.82), T<sub>9</sub> (5.62), (5.70), T<sub>8</sub> (5.51) and (5.64). Least Canopy Temperature depression (CTD) was Observed under T<sub>5</sub> (5.11) and (5.19) during both the years of investigation. Sharma *et al.* (2003) <sup>[32]</sup> reported that canopy temperature depression (CTD 0C), relative water content (%) and water potential decreased significantly with increases water deficit in soil.

#### Chlorophyll Intensity (SPAD) (%) Pre and post anthesis

The maximum Chlorophyll Intensity (SPAD) (%) Pre anthesis in wheat was observed in T<sub>3</sub> (44.25) and (44.78) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (44.10) and (44.66) which was statistically at par with T<sub>2</sub> (43.80) and (43.92), T<sub>10</sub> (43.71) and (43.79), T<sub>9</sub> (43.55) and (43.60) and T<sub>8</sub> (43.30) and (43.56). Least Chlorophyll Intensity (SPAD) (%) Pre anthesis was Observed under T<sub>5</sub> (42.11) and (42.40) during both the years of investigation.

The maximum Chlorophyll Intensity (SPAD) (%) Post anthesis in wheat was observed in T<sub>3</sub> (42.55) and (42.66) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (42.13) and (42.22) which was statistically at par with T<sub>2</sub> (41.81) and (41.88), T<sub>10</sub> (41.43) and (41.51), T<sub>9</sub> (41.31) and (41.38) and T<sub>8</sub> (41.05) and (41.13). Least Chlorophyll Intensity (SPAD) (%) Post anthesis was Observed under T<sub>5</sub> (40.21) and (40.27) during both the years of investigation. Patil *et al.* (2002) <sup>[41]</sup> conducted an experiment at Akola (Maharashtra) and investigated that the effect of five sowing dates (initial sowing was performed on 15th November, whereas, successive sowing was conducted at 15-day intervals) on wheat cultivars *viz.*, AKW-1071, AKW-381, AKW-3294, NIAW34, HI-977, and HD-2501. It was observed that the sowing on 15th November resulted in

the highest chlorophyll content. Singh *et al.* (2002)<sup>[42]</sup> reported that chlorophyll content was higher in 10th and 30th October sowing as compared to 20th November and 10th December). Fattah *et al.* (1998)<sup>[43]</sup> observed that foliar spray of 2 ppm and 4 ppm triacontanol significantly increased the plant height, leaf area index, dry weight per plant and chlorophyll content in leaves of niger as compared to control.

#### Relative Water Content (RWC) (%) Pre anthesis

The maximum Relative Water Content (RWC) (%) Pre anthesis in wheat was observed in T<sub>3</sub> (72.42) and (72.65) with application of IAA 50 PPM which was significantly superior over T<sub>11</sub> (70.97) and (71.19) which was statistically at par with T<sub>2</sub> (70.07), (70.29), T<sub>10</sub> (68.99), (69.21), T<sub>9</sub> (68.63), (68.84), T<sub>8</sub> (68.09) and (68.30). Least Relative Water Content (RWC) (%) Pre anthesis was Observed under T<sub>5</sub> (65.66) and (65.87) during both the years of investigation. Bano *et al.* (2012) demonstrated that exogenous application of ABA protected wheat from drought-induced oxidative damage by improving the antioxidant system and relative water content.

#### Protein Content (%) Pre anthesis

The maximum Protein Content (%) Pre anthesis in wheat was observed in T (11.45) and (11.48) with application of IAA 50

ppm which was significantly superior over T<sub>11</sub> (11.40) and (11.43) which was statistically at par with T<sub>2</sub> (11.37) and (11.40), T<sub>10</sub> (11.37) and (11.40), T<sub>9</sub> (11.34) and (11.37) and T<sub>8</sub> (11.33) and (11.36). Least Protein Content (%) Pre anthesis was Observed under T<sub>5</sub> (11.18) and (11.21) during both the years of investigation. Concluded that the foliar application of TRIA has been reported to enhance chlorophyll contents, total soluble sugars, protein, nucleic acids, photosynthetic rate and chlorophyll fluorescence in wheat. Hayat *et al.* (2007)<sup>[24]</sup> reported that activity of synthesis of protein in plants was enhanced greatly under stressful condition.

#### Main spike length

The maximum main spike length in wheat was observed in T<sub>3</sub> (9.10) and (9.12) with application of IAA 50 ppm which was significantly superior over T<sub>11</sub> (8.99) and (9.05) which was statistically at par with T<sub>2</sub> (8.85) and (8.86), T<sub>10</sub> (8.73) and (8.75), T<sub>9</sub> (8.70), (8.73), T<sub>8</sub> (8.68), (8.70). Least main spike length was Observed under T<sub>5</sub> (8.60) and (8.62) during both the years of investigation. Emam, and Cartwright, (1998)<sup>[44]</sup> reported that hormonal treatments stimulated significant increase in number of tillers, spike length, biomass and grain yield of the plants when composition was sprayed.

**Table 1:** Effect of plant growth regulators on Number of tiller Plant at different growth stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	Number of tiller Plant 30 I year	Number of tiller Plant 30 II year	Number of tiller Plant 60 I year	Number of tiller Plant 60 II year	Number of tiller Plant 90 I year	Number of tiller Plant 90 II year	Number of tiller Plant at harvest I year	Number of tiller Plant at harvest II year
T <sub>0</sub> Control	2.00	2.11	3.41	3.52	4.22	4.44	4.66	4.77
T <sub>2</sub> IAA 25 ppm	2.80	2.86	5.66	5.70	6.33	6.40	7.22	7.33
T <sub>3</sub> IAA 50 ppm	2.99	3.11	6.22	6.32	6.99	7.11	7.77	7.89
T <sub>4</sub> Absciscic acid 10 ppm	2.41	2.47	4.44	4.50	5.65	5.72	5.44	5.51
T <sub>5</sub> Absciscic acid 20 ppm	2.36	2.41	4.22	4.33	5.38	5.43	5.51	5.55
T <sub>6</sub> TRIA 5ppm	2.45	2.50	4.70	4.88	5.70	5.76	5.76	5.88
T <sub>7</sub> TRIA 10 ppm	2.55	2.61	4.99	5.11	5.74	5.81	5.99	5.75
T <sub>8</sub> Brassinosteroid 5 ppm	2.61	2.64	5.11	5.20	5.90	5.98	6.00	6.11
T <sub>9</sub> Brassinosteroid 10 ppm	2.60	2.66	5.40	5.44	6.05	6.10	6.34	6.45
T <sub>10</sub> Cyto kinin 5 ppm	2.66	2.71	5.60	5.64	6.35	6.39	6.44	6.56
T <sub>11</sub> Cyto kinin 10 ppm	2.88	2.89	5.66	5.80	6.89	7.00	7.44	7.56
S.Em±	0.056	0.072	0.115	0.092	0.156	0.164	0.140	0.171
CD at 5%	0.170	0.219	0.349	0.279	0.472	0.496	0.425	0.520

**Table 2:** Effect of plant growth regulators on Number of leaves per plant at different growth stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	Number of leaves per plant 30 I Year	Number of leaves per plant 30 II Year	Number of leaves per plant 60 I Year	Number of leaves per plant 60 II Year	Number of leaves per plant 90 I Year	Number of leaves per plant 90 II Year
T <sub>0</sub> Control	5.91	6.11	12.17	19.37	20.15	20.40
T <sub>2</sub> IAA 25 ppm	6.77	6.76	12.78	22.87	23.00	23.98
T <sub>3</sub> IAA 50 ppm	7.11	7.25	13.35	23.80	24.51	24.94
T <sub>4</sub> Absciscic acid 10 ppm	6.00	6.32	13.21	20.61	21.23	21.75
T <sub>5</sub> Absciscic acid 20 ppm	6.10	6.22	13.03	20.38	21.10	21.33
T <sub>6</sub> TRIA 5 ppm	6.31	6.36	13.45	21.57	22.33	22.66
T <sub>7</sub> TRIA 10 ppm	6.37	6.41	12.54	21.79	22.55	22.88
T <sub>8</sub> Brassinosteroid 5 ppm	6.49	6.54	13.62	22.41	22.70	22.91
T <sub>9</sub> Brassinosteroid 10 ppm	6.55	6.60	13.74	22.67	22.96	23.33
T <sub>10</sub> Cytokinin 5 ppm	6.61	6.69	13.77	22.73	23.51	23.90
T <sub>11</sub> Cytokinin 10 ppm	6.88	6.91	13.91	23.18	24.11	24.40
S.Em±	0.140	0.130	0.156	0.434	0.605	0.491
CD at 5%	0.426	0.395	0.468	1.318	1.834	1.490

**Table 3:** Effect of plant growth regulators on leaf area per plant at different growth stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatment s (T)	leaf area per plant 30 I Year	leaf area per plant 30 II Year	leaf area per plant 60 I Year	leaf area per plant 60 II Year	leaf area per plant 90 I Year	leaf area per plant 90 II Year
T <sub>0</sub> Control	47.95	48.17	451.65	452.76	658.15	660.60
T <sub>2</sub> IAA 25 ppm	50.05	50.27	502.82	503.92	732.22	745.10
T <sub>3</sub> IAA 50 ppm	50.50	50.72	513.65	514.31	748.14	750.10
T <sub>4</sub> Abscisic acid 10 ppm	49.58	49.80	470.77	471.66	727.35	728.40
T <sub>5</sub> Abscisic acid 20 ppm	49.43	49.74	465.91	466.57	725.10	726.50
T <sub>6</sub> TRIA 5 ppm	49.61	49.83	475.19	476.30	729.70	730.30
T <sub>7</sub> TRIA 10 ppm	49.73	49.88	477.63	478.51	731.85	732.30
T <sub>8</sub> Brassinosteroid 5 ppm	49.76	49.91	482.27	483.37	732.80	733.30
T <sub>9</sub> Brassinosteroid 10 ppm	49.91	50.05	486.02	488.01	736.70	737.20
T <sub>10</sub> Cytokinin 5 ppm	49.98	50.20	488.90	489.78	738.20	738.60
T <sub>11</sub> Cytokinin 10 ppm	50.20	50.35	496.85	497.74	742.00	742.30
S.Em±	1.322	1.064	12.915	10.426	14.120	19.824
CD at 5%	4.009	3.228	39.177	31.626	42.834	60.136

**Table 4:** Effect of plant growth regulators on Dry matter production per plant (g) at different growth stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	Dry matter production plant <sup>-1</sup> (g) 30 I Year	Dry matter production plant <sup>-1</sup> (g) 30 II Year	Dry matter production plant <sup>-1</sup> (g) 60 I Year	Dry matter production plant <sup>-1</sup> (g) 60 II Year	Dry matter production plant <sup>-1</sup> (g) 90 I Year	Dry matter production plant <sup>-1</sup> (g) 90 II Year	Dry matter production plant <sup>-1</sup> (g) Harvest I Year	Dry matter production plant <sup>-1</sup> (g) Harvest II Year
T <sub>0</sub> Control	2.34	2.36	9.53	9.66	18.50	18.89	24.52	25.10
T <sub>2</sub> IAA 25 ppm	2.40	2.41	10.44	10.53	20.62	20.84	27.76	28.14
T <sub>3</sub> IAA 50 ppm	2.41	2.43	10.79	10.88	21.32	21.55	29.10	29.45
T <sub>4</sub> Abscisic acid 10 ppm	2.39	2.40	10.11	10.24	19.85	20.12	26.53	26.95
T <sub>5</sub> Abscisic acid 20 ppm	2.39	2.41	10.23	10.35	20.17	20.40	26.97	27.36
T <sub>6</sub> TRIA 5 ppm	2.38	2.39	9.99	10.11	19.60	19.85	26.15	26.56
T <sub>7</sub> TRIA 10 ppm	2.38	2.40	10.22	10.31	20.15	20.38	26.99	27.30
T <sub>8</sub> Brassinosteroid 5 ppm	2.36	2.37	9.79	9.87	19.17	19.36	25.55	25.86
T <sub>9</sub> Brassinosteroid 10 ppm	2.38	3.39	9.91	9.99	19.42	19.71	25.90	26.35
T <sub>10</sub> Cytokinin 5 ppm	2.40	2.41	10.32	10.40	20.37	20.57	27.35	27.75
T <sub>11</sub> Cytokinin 10 ppm	2.41	2.44	10.58	10.76	20.90	21.30	28.50	29.10
S.Em±	0.063	0.051	0.273	0.219	0.394	0.548	0.538	0.734
CD at 5%	0.192	0.156	0.829	0.663	1.194	1.662	1.631	2.226

**Table 5:** Effect of plant growth regulators on Days to heading and Days to maturity in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	Days of heading I Year	Days of heading II Year	Days of maturity I Year	Days of maturity II Year
T <sub>0</sub> Control	75.10	76.23	122.33	123.66
T <sub>2</sub> IAA 25 ppm	77.33	77.67	125.00	126.15
T <sub>3</sub> IAA 50 ppm	77.11	78.34	126.00	127.00
T <sub>4</sub> Abscisic acid 10 ppm	76.30	77.21	124.66	125.33
T <sub>5</sub> Abscisic acid 20 ppm	76.33	76.55	126.00	126.30
T <sub>6</sub> TRIA 5 ppm	75.00	75.27	122.66	122.66
T <sub>7</sub> TRIA 10 ppm	75.33	76.34	122.00	122.33
T <sub>8</sub> Brassinosteroid 5 ppm	72.16	72.46	121.33	121.66
T <sub>9</sub> Brassinosteroid 10 ppm	72.66	73.58	120.60	121.00
T <sub>10</sub> Cytokinin 5 ppm	73.33	74.81	120.00	120.30
T <sub>11</sub> Cytokinin 10 ppm	71.66	72.92	119.00	119.15
S.Em±	1.562	2.029	2.541	3.289
CD at 5%	4.737	6.154	7.709	9.978

**Table 6:** Effect of plant growth regulators on Leaf Area Index at different growth stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	Lai 30 <sup>1</sup> Year	Lai 30 <sup>2</sup> Year	Lai 60 <sup>1</sup> Year	Lai 60 <sup>2</sup> Year	Lai 90 <sup>1</sup> Year	Lai 90 <sup>2</sup> Year	Lai Harvest <sup>1</sup> Year	Lai Harvest <sup>2</sup> Year
T <sub>0</sub> Control	0.23	0.24	2.00	2.03	2.91	3.11	4.77	5.91
T <sub>2</sub> IAA 25ppm	0.25	0.26	2.30	2.32	3.33	3.36	7.33	6.77
T <sub>3</sub> IAA 50 ppm	0.26	0.28	2.38	2.40	3.46	3.48	7.89	7.11
T <sub>4</sub> Abscisic acid 10 ppm	0.24	0.25	2.19	2.21	3.16	3.19	5.51	6.00
T <sub>5</sub> Abscisic acid 20 ppm	0.24	0.25	2.05	2.13	3.11	3.17	5.55	6.10
T <sub>6</sub> TRIA 5 ppm	0.25	0.26	2.20	2.22	3.20	3.22	5.88	6.31
T <sub>7</sub> TRIA 10 ppm	0.25	0.26	2.22	2.23	3.21	3.24	5.75	6.37
T <sub>8</sub> Brassinosteroid 5 ppm	0.25	0.26	2.23	2.25	3.24	3.28	6.11	6.49
T <sub>9</sub> Brassinosteroid 10 ppm	0.25	0.26	2.25	2.27	3.26	3.29	6.45	6.55

T <sub>10</sub> Cytokinin 5 ppm	0.25	0.26	2.26	2.28	3.27	3.30	6.56	6.61
T <sub>11</sub> Cytokinin 10 ppm	0.25	0.27	2.33	2.35	3.39	3.42	7.56	6.88
S.Em±	0.005	0.007	0.048	0.044	0.086	0.087	0.171	0.140
CD at 5%	0.016	0.021	0.146	0.133	0.262	0.265	0.520	0.426

**Table 7:** Effect of plant growth regulators on Relative growth rate at different growth stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	RGR 30 I year	RGR 30 II year	RGR 60 I year	RGR 60 II year	RGR 90 I year	RGR 90 II year
T <sub>0</sub> Control	46.81	46.97	22.11	22.35	9.39	9.47
T <sub>2</sub> IAA 25 ppm	49.00	49.15	22.68	22.75	9.91	10.01
T <sub>3</sub> IAA 50 ppm	49.96	51.80	22.70	22.78	10.37	10.44
T <sub>4</sub> Abscisic acid 10 ppm	48.07	48.36	22.49	22.51	9.66	9.74
T <sub>5</sub> Abscisic acid 20 ppm	48.47	48.57	22.62	22.71	9.68	9.78
T <sub>6</sub> TRIA 5 ppm	47.81	48.07	22.43	22.49	9.61	9.70
T <sub>7</sub> TRIA 10 ppm	48.57	48.59	22.63	22.71	9.74	9.79
T <sub>8</sub> Brassinosteroid 5 ppm	47.42	47.55	22.40	22.46	9.57	9.64
T <sub>9</sub> Brassinosteroid 10 ppm	47.54	47.67	22.42	22.48	9.58	9.67
T <sub>10</sub> Cytokinin 5 ppm	48.62	48.71	22.66	22.73	9.80	9.98
T <sub>11</sub> Cytokinin 10 ppm	49.31	49.46	22.69	22.76	10.34	10.40
S.Em±	1.034	1.303	0.481	0.458	0.265	0.202
CD at 5%	3.137	3.953	1.458	1.390	0.804	0.613

**Table 8:** Effect of plant growth regulators on Canopy temperature depression (CTD and Chlorophyll intensity (SPAD) (%) at Pre and post anthesis stages in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	Canopy temperature depression (CTD) Pre-anthesis I Year	Canopy temperature depression (CTD) Pre-anthesis II Year	Canopy temperature depression (CTD) Post-anthesis I Year	Canopy temperature depression (CTD) Post-anthesis II Year	Chlorophyll intensity (SPAD) (%) Pre-anthesis I Year	Chlorophyll intensity (SPAD) (%) Pre-anthesis II Year	Chlorophyll intensity (SPAD) (%) Post-anthesis I Year	Chlorophyll intensity (SPAD) (%) Post-anthesis II Year
T <sub>0</sub> Control	4.15	4.19	4.81	5.21	40.15	40.36	38.25	38.50
T <sub>2</sub> IAA 25 ppm	4.61	4.68	5.93	6.01	43.80	43.92	41.81	41.88
T <sub>3</sub> IAA 50 ppm	5.05	5.16	6.75	6.83	44.25	44.78	42.55	42.66
T <sub>4</sub> Abscisic acid 10 ppm	4.10	4.15	5.21	5.26	42.50	42.61	40.66	40.71
T <sub>5</sub> Abscisic acid 20 ppm	4.05	4.11	5.11	5.19	42.11	42.40	40.21	40.27
T <sub>6</sub> TRIA 5 ppm	4.16	4.22	5.30	5.38	42.85	43.05	40.71	40.78
T <sub>7</sub> TRIA 10 ppm	4.20	4.28	5.43	5.55	43.11	43.31	40.84	40.92
T <sub>8</sub> Brassinosteroid 5 ppm	4.24	4.32	5.51	5.64	43.30	43.56	41.05	41.13
T <sub>9</sub> Brassinosteroid 10 ppm	4.33	4.40	5.62	5.70	43.55	43.60	41.31	41.38
T <sub>10</sub> Cytokinin 5 ppm	4.55	4.63	5.76	5.82	43.71	43.79	41.43	41.51
T <sub>11</sub> Cytokinin 10 ppm	4.76	4.81	6.15	6.22	44.10	44.66	42.13	42.22
S.Em±	0.118	0.095	0.153	0.124	0.849	1.166	1.094	0.881
CD at 5%	0.357	0.287	0.463	0.375	2.575	3.537	3.318	2.673

**Table 9:** Effect of plant growth regulators on Relative water content (RWC) and Protein content (%) at Pre anthesis stage in wheat (*Triticum aestivum* L.) under late sown condition

Treatments (T)	RWC Pre anthesis I Year	RWC Pre anthesis II Year	Protein content (%) Pre anthesis I Year	Protein content (%) Pre anthesis II Year
T <sub>0</sub> Control	63.95	64.15	11.05	11.14
T <sub>2</sub> IAA 25 ppm	70.07	70.29	11.37	11.40
T <sub>3</sub> IAA 50 ppm	72.42	72.65	11.45	11.48
T <sub>4</sub> Abscisic acid 10 ppm	66.47	66.68	11.22	11.26
T <sub>5</sub> Abscisic acid 20 ppm	65.66	65.87	11.18	11.21
T <sub>6</sub> TRIA 5 ppm	67.05	67.26	11.28	11.31
T <sub>7</sub> TRIA 10 ppm	67.37	67.58	11.30	11.34
T <sub>8</sub> Brassinosteroid 5 ppm	68.09	68.30	11.33	11.36
T <sub>9</sub> Brassinosteroid 10 ppm	68.63	68.84	11.34	11.37
T <sub>10</sub> Cytokinin 5 ppm	68.99	69.21	11.37	11.40
T <sub>11</sub> Cytokinin 10 ppm	70.97	71.19	11.40	11.43
S.Em±	1.820	1.473	0.228	0.310
CD at 5%	5.521	4.468	0.691	0.941



## Conclusions

On the basis of results presented and discussed in preceding chapters concluded that maximum yield and productivity and build up in soil fertility might be achieved with the application of recommended doses of fertilizer along with IAA 50 PPM, IAA 25 ppm and Cytokinin 5 ppm as a growth enhancer. Hence it may be recommended for enhancing productivity of wheat.

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## References

1. Fischer RA. Statistical analysis. Oliver Boyed. London and Edinurg; c1937.
2. Gupta NK, Gupta S, Kumar A. Effect of water stress on physiological attributes and their relationship with growth and yield of wheat cultivars at different stages. J Agron. Crop Sci. 2001;186:55-62.
3. Rademacher W, Brahm L. Plant Growth Regulators, Ullmann's Encyclopedia of Industrial Chemistry, Wiley-Vch Verlag Gmbh and Co. Kгаа; c2010.
4. Rahman MS, Islam MATN, Karim MA. Influence of GA<sub>3</sub> and MH and their time of spray on dry matter accumulation and growth attributes of soybean. Pak. J of Biol. Sci. 2004;7(1):1851-1857.
5. Rai RK, Tripathi N, Gautam D, Singh P. Exogenous application of ethrel and Gibberellic acid stimulates physiological growth of late planted sugarcane with short growth period in sub-tropical India. J Plant Growth Regul. 2017;36:472-486.
6. Rai VK, Sharma SS, Sharma S. Reversal of ABA-induced stomatal closure by phenolic compounds. Journal of Experimental Biology.1986;37:129-134.
7. Rajjou L, Belghazi M, Huguot R, Robin C, Moreau A, Job C, *et al.* Proteomic investigation of the effect of salicylic acid on Arabidopsis seed germination and establishment of early defense mechanisms. Plant Physiology.2006;141:910-923.
8. Rakhmankulova ZF, Fedyayev VV, Rakhmatulina SR, Ivanov CP, Gilvanova IR, Usmanov IY. The effect of wheat seed presowing treatment with salicylic acid on its endogenous content, activities of respiratory pathways, and plant antioxidant status. Russia Journal of Plant Physiology.2010;57:778-783.
9. Roostaei M, Mohammadi SA, Amri A, Majidi E, Nachit M, Haghparast R. Chlorophyll fluorescence parameters and drought tolerance in a mapping population of winter bread wheat in the highlands of Iran. Russ. J Plant Physiol. 2011;58:351-358.
10. Rout S, Beura S, Khare N, Patra SS, Nayak S. Effect of seed pre-treatment with different concentrations of Gibberellic acid (GA<sub>3</sub>) on seed germination and seedling growth of *Cassia fistula* L. J Med. Plants Stud. 2017;5(6):135-138.
11. Rukasz I, Michalek W. Effect of foliar application of phytohormonal on barley yield. Annals Univ. Mariae Curie-Sklodowska. Sectio E. Agri. 2004;59(4):1543-1548.
12. Ruuska SA, Rebetzke GJ, Van Herwaarden AF, Richards RA, Fettell NA, *et al.* Genotypic variation in water-soluble carbohydrate accumulation in wheat. Functional Plant Biology. 2006;33:799-809.
13. Sahu GK, Sabat SC. Changes in growth, pigment content and antioxidants in the root and leaf tissues of wheat plants under the influence of exogenous salicylic acid. Brazil Journal of Plant Physiology. 2011;23:209-212.
14. Saini HS, Aspinall D. Abnormal sporogenesis in wheat (*Triticum aestivum* L.) induced by short periods of high temperature. Annual Review of Botany.1982;49:835-846.
15. Saint Pierre C, Trethowan R, Reynolds M. Stem solidness and its relationship to water-soluble carbohydrates: Association with wheat yield under water deficit. Functional Plant Biology. 2010;37:166-174.
16. Sarkar PK, Haque MDS, Karim MA. Growth Analysis of Soybean as Influenced by GA<sub>3</sub> and IAA and their Frequency of Application. J of Agron. 2002;1:123-126.
17. Savchenko T, Walley JW, Chehab EW, Xiao Y, Kaspi R, Pye MF, *et al.* Arachidonic acid: an evolutionarily conserved signaling molecule modulates plant stress signaling networks. Plant Cellular Biology. 2010;22:3193-3205.
18. Scharf KD, Heider H, Hohfeld I, Lyck R, Schmidt E, Nover L. The tomato HSF system: HsfA2 needs interaction with HsfA1 for efficient nuclear import and may be localized in cytoplasmic heat stress granules. Molecular Cell Biology.1998;18:2240-2251.
19. Scharf KD, Hohfeld I, Nover L. Heat stress response and heat stress transcription factors. Journal of Bioscience. 1998b;23:313-329.
20. Schwartz SH, Tan BC, Gage DA, Zeevaart JA, McCarty DR. Specific oxidative cleavage of carotenoids by VP14 of maize. Science.1997;276:1872-1874.
21. Scofield GN, Ruuska SA, Aoki N, Lewis DC, Tabe LM, Jenkins CL. Starch storage in the stems of wheat plants: localization and temporal changes. Annual Review of Botany.2009;103:859-868.
22. Seleiman M, Ibrahim M, Abdel-Aal S, Zahran G. Effect of sowing dates on productivity, technological and rheological characteristics of bread wheat. J Agron. Crop Sci. 2011;2(1):1-6.
23. Semenov MA, Halford NG. Identifying target traits and molecular mechanisms for wheat breeding under a changing climate. Journal of Experimental Botany. 2009;60:2791-2804.
24. Shakirova F. Role of hormonal system in the manifestation of growth promoting and anti-stress action of salicylic acid. In: Hayat S, Ahmad A (Eds.), Salicylic acid: a plant hormone. Springer, Netherlands; c2007. p. 69-89.
25. Shakirova FM. Role of hormonal system in the manifestation of growth promoting and antistress action of salicylic acid. In Salicylic acid: a plant hormone. Springer, Netherlands; c2007.
26. Shakirova FM, Sakhabutdinova AR, Bezrukova MV, Fatkhutdinova RA, Fatkhutdinova DR. Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. Plant Science. 2003;164:317-322.
27. Shakirova FM, Shakhutdinova AR, Bezrukova MV, Fatkhutdinova RA, Fatkhutdinova DR. Changes in the hormonal status of wheat seedling induced by salicylic acid and salinity. Plant Sci. 2003;164:317-22.
28. Shamsi K, Kobraee S, Rasekhi B. Variation of yield



- components and some morphological traits in bread wheat grown under drought stress. *Ann. Bio. Res.* 2011;2:372-377.
29. Shani E, Yanai O, Ori N. The role of hormones in shoot apical meristem function. *Plant Biology.* 2006;5(9):488-489.
  30. Sharafizad M, Naderi A, Ata Saidat S, Sakinezad T. Effect of salicylic acid on wheat yield and its components under drought stress. *Advances in environmental biology.* 2013;7(4):629-635.
  31. Sharkey TD. Effects of moderate heat stress on photosynthesis: Importance of thylakoid reactions, rubisco deactivation, reactive oxygen species and thermotolerance provided by isoprene. *Plant Cell Environment.* 2005;28:269-277.
  32. Sharma KD, Pannu RK, Tyagi PK, Chaudhary BD, Singh DP. Effect of moisture stress on plant water relations and yield of different wheat genotypes. *Indian J of Plant Physiol.* 2003;8:99-102.
  33. Yang J, Peng S, Visperas RM, Sanico AL, Zhu Q, Gu S. Grain filling pattern and cytokinin content in the grains and roots of rice plants. *Plant Growth Regulation.* 2000;30:261-270.
  34. Ahmadi A, Baker DA. Effects of abscisic acid (ABA) on grain filling processes in wheat. *Plant Growth Regulation.* 1998;28:187-197.
  35. Negi, *et al.* *Indian For.* 2003;129:757-769.
  36. Wahid, *et al.* Pretreatment of seed with H<sub>2</sub>O<sub>2</sub> improves salt tolerance of wheat seedlings by alleviation of oxidative damage and expression of stress proteins. *J Plant Physiol.* 2007;164:283-294.
  37. Shah, *et al.* A Effect of sowing dates on yield and yield components of different wheat varieties. *J Agron.* 2006;5(1):106-110.
  38. Dhaka *et al.* Phenological development, yield and yield attributes of different wheat genotypes as influenced by sowing time and irrigation levels, *Pakistan Journal of Agricultural;* c2006.
  39. Shahzad MA, Wasi-ud-Din Sahi, Khan ST, Ehsanullah MM, Ahmad M. Effect of sowing dates and seed treatment on grain yield and quality of wheat, *Pakistan Journal of Agricultural Science.* 2007;44(4):581-583.
  40. Singh, *et al.* Management practices to mitigate the impact of high temperature on wheat: a review *IIOAB J.* 2011;2(7):11-22.
  41. Patil, *et al.* Nitrogen uptake and grain protein in emmer wheat genotype as influenced by sowing dates and nitrogen levels. *Karnataka J Agric. Sci.* 2002;15:349-352.
  42. Singh, *et al.* Growth, yield and phenological response of wheat cultivars to delayed sowing. – *Indian Journal of Plant Physiology.* 2002;8(3):277-286.
  43. Fattah, *et al.* Physiological response of Niger (*Guizotia abyssinica*) to mixtalol and potassium naphthenate application. *Bangladesh Journal of Botany.* 1998;27(2):97-102.
  44. Emam Y, Cartwright P. Effects of drying soil and CCC on root: shoot growth and water use in barley plants. *Monograph-British Society for Plant Growth Regulation.* 1998;21:389-392.
  45. Darussalam M, Cole MA, Patrick JW. Auxin control of photo-assimilate transport to and within developing grain of wheat. *Australian Journal of Plant Physiology.* 1998;25:69-78.
  46. Lejeune P, Prinsen E, Van Onckelen H, Bernier G. Hormonal control of ear abortion in a stress-sensitive maize (*Zea mays*) inbred. *Australian Journal of Plant Physiology.* 1998;25:481-488.
  47. Espindula MC, Rocha VS, Grossi JAS, Souza MA, Souza LT, Favarato LF. Use of growth retardants in wheat. *Planta Daninha.* 2009;27:379-387.
  48. Kim YH, Khan AL, Hamayun M, Kim JT, Lee JH, Hwang IC, *et al.* Effects of prohexadione calcium on growth and gibberellins contents of *Chrysanthemum morifolium* R. cv Monalisa White. *Scientia Horticulture.* 2010;123:423-427.
  49. Hussain Z, Leitch MH. The effect of sulphur and growth regulators on growth characteristics and grain yield of spring sown wheat. *Journal of Plant Nutrition.* 2007;30:67-77.
  50. Meena JP. Photosynthesis, growth and yield of wheat (*Triticum aestivum* L.) varieties at different dates of sowing. Thesis M.Sc. Ag. (Agronomy), G.B. Pant University of Agriculture and technology, Pant nagar (U.S. nagar); c2009. p.172.
  51. Patil S, Subbanna. Effect of triacntanol on growth, yield and ascorbic acid content of lettuce (*Lactuca sativa* L.). *Crop Research.* 1988;1(2):141-145.
  52. Regmi AP, Ladha JK, Pathak H, Pasuquin E, Dawe D, Hobbs PR. Analyses of yield and soil fertility trends in a 20-year rice-wheat experiment in Nepal. *Soil Science Society of America Journal* 2002;66:857-867.
  53. Jia W, Wang Y, Zhang S, Zhang J. Salt-stress-induced ABA accumulation is more sensitively triggered in roots than in shoots. *J Exp. Bot.* 2003;53:2201-2206.
  54. Hayat S, Ahmad A, Mobin M, Hussain A, Fariduddin Q. Photosynthetic rate, growth, and yield of mustard plants sprayed with 28-homobrassinolide. *Photosynthetica.* 2001;38:469-71.
  55. Yan W, Zhang XX, Yuan A. Effects of two plant growth regulators on the growth and recovery of alfalfa seedlings exposed to Aluminum stress. *J Shanghai Jiaotong Univ (Agric Sci).* 2011;29(2):75-82.
  56. Morinaka Y, Sakamoto, Yoshiaki I, Masakazu A, Hidemi K, Motoyuki A, Makoto M. Morphological alteration caused by Brassinosteroid insensitivity increases the biomass and grain production of rice. *Plant Physiol.* 2006;141:924-931.
  57. Arshad M, Frankenberger WT. Plant growth-regulating substances in the rhizosphere: Microbial production and functions. *Advances in Agronomy.* 1998;62:46-152.
  58. Hobbs PR, Gupta RK. Resource conserving technologies for wheat in rice-wheat systems. In Ladha JK, James EH, Duxbury JD, Gupta RK, Buresh RJ. (eds.) *Improving the productivity and sustainability of rice-wheat systems: Issues and impact.* ASA Special Publication, ASA, Madison, Wisconsin, USA; c2002.
  59. Sharma-Natu P, Sumesh KV, Lohot VD, Ghildiyal MC. High temperature effect on grain growth in wheat cultivars: An evaluation of responses. *Indian Journal of Plant Physiology.* 2006;11:239-245.
  60. Ugarte C, Calderini DF, Slafer GA. Grain weight and grain number responsiveness to pre-anthesis temperature in wheat, barley and triticale. *Field Crops Research.* 2007;100:240-248.
  61. Hensel LL, Grbic V, Baumgarten DA, Blecker AB. Developmental and age-related processes that influence

- the longevity and senescence of photosynthetic tissues in Arabidopsis. *Plant Cell*. 1993;5:553-564.
62. Pandey A, Chaudhry S, Sharma A, Choudhary VS, Malviya MK, Chamoli S, Rinu K. Recovery of *Bacillus* and *Pseudomonas* spp from the 'Fired Plots' under shifting cultivation in Northeast India. *Curr. Microbiol*. 2011;62:273-280.
63. Hussain A, Maqsood M, Ahmad A, Aftab S, Ahmad Z. Effect of irrigation during various development stages on yield, component of yield and harvest index of different wheat cultivars. *Pakistan J Agric. Sci*. 1998;34:104-107.
64. Tripathi SC, Sayre KD, Kaul JN, Narang RS. Growth and morphology of spring wheat (*Triticum aestivum* L.) culms and their association with lodging: effects of genotypes, N levels and ethephon. *Field Crops Research*. 2003;84:271-290.
65. Pan M, Zhou X, Chen M. Cellulose Nano whiskers isolation and properties from acid hydrolysis combined with high pressure homogenization. *Bio Resources*. 2013 Jan 9;8(1):933-43.