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Effect of herbal treatment on haemato-biochemical recovery in a *Colibacillosis* calves

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Abstract

The objective of the present study was to assess the *in-vitro* antimicrobial efficacy of *Punica granatum* rind and *Psidium guajava* leaf extract on enteropathogenic *Escherichia coli* and to evaluate its effectiveness in the treatment of *colibacillosis* in calf. The experiment was conducted at Department of Veterinary Medicine, College of Veterinary Science and Animal Husbandry, Birsa Agricultural University, Kanke Ranchi. A total no. of 60 *colibacillosis* positive calves (*Escherichia coli* positive) and six (06) clinically normal healthy calves (Healthy control) were taken to investigate the haemato-biochemical profiles. *E. coli* was isolated from the *colibacillosis* affected calves and the animals found positive were used for the study of haemato-biochemical profile. Faecal sample were cultured on MacConkey and EMB agar to diagnose *E. coli*. Analysis of blood and serum samples of the *colibacillosis* affected calves revealed significant increase in Haemoglobin, Total erythrocyte count, Total leukocyte count, Blood urea nitrogen, Total serum protein, Serum creatinine, and Serum potassium, while significant decrease in Blood glucose and Serum sodium. However, no significant changes were observed in Alanine transaminase.

Keywords: calf, *colibacillosis*, *E. coli*, haemato-biochemistry

Introduction

Colibacillosis is a leading cause of calf mortality and morbidity during first few week of life (Radostits *et al.* 2000) ^[1]. *Colibacillosis* is one of the challenging tasks to veterinary practitioners. The various known causes of calf scour are grouped under infectious and noninfectious (Izzo *et al.* 2011) ^[2]. Various antimicrobial agents have been the drugs of choice for the clinician but it has been observed that prolong use of oral antibiotics is detrimental to the intestinal mucosa resulting into mal-absorption diarrhoea and even development of drug resistant (Mero *et al.* 1985) ^[3]. Guava leaves and Pomegranate peels have been widely used as potent anti-diarrheal agent by rural peoples (Lutterrodt, 1989; and Alnieida *et al.* 1995) ^[4, 5]. Thus the present work has been designed to assess the treatment effect of *Psidium guajava* leaves and *Punica granatum* rind extract in restoration of haemato-biochemical changes in *colibacillosis* calves.

Materials and Methods

A total of 60 *colibacillosis* affected calves presented or reported at Veterinary Clinical Complex (RVC), Instructional bovine farm of Ranchi Veterinary College, and from local organized or unorganized Dairy farm situated in an around Ranchi Veterinary College with the history of passage of soft fluid faeces, abnormal colour of faeces (white to yellowish green and mucoid) along with rapid loss of body weight, animal becoming lean and dehydrated was selected for the present study. The faecal sample were collected directly from the rectum of *colibacillosis* affected calves with the help of sterilized swab, the swab materials were then kept in a pre-enriched peptone water broth for its culture and isolation of bacteria. Pre-enriched sample were cultured on MacConkey Agar and on eosine methylene blue agar as per standard method Soulsby (1982) ^[6]. Gram's staining was done as per method described by Stokes *et al.* (1993) ^[7]. Further, biochemical test like IMViC test was done for its confirmation of isolates biochemical testing of *E. coli* was performed by using commercially available KB001 HiMedia *E. coli* Identification Kit (HiMedia Laboratories Pvt. Limited).

Experimental design

On the basis of culture and sensitivity test the selected calves were randomly distributed into 3 equal groups i.e. group-A, group-B and group-C consist of 20 animals in each group.

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Group-A was treated with Ofloxacin tab. orally, group-B was treated with ethanolic extract of *Punica granatum* rind orally and group-C was treated with ethanolic extract of *Psidium guajava* leaf orally. The entire treatments are continued for upto 7 days along with supportive therapy consisting of 5% DNS, B-complex inj. and liver tonic inj.

Collection of Blood Samples

Blood samples were collected on day 0 (pre-treatment) and on 2nd, 4th, 6th and 8th days of post-treatment for estimation of hemato-biochemical parameters. A total of 8 ml blood sample was collected aseptically through sterilized disposable syringe, directly from jugular vein. 2 ml of them were transferred to a test tube containing EDTA for its hematological examination (Hemoglobin, Total leukocyte count, Total erythrocyte count) by using Veterinary hematological Analyzer (Vet Scan HM5). Rest 6 ml of blood sample was transferred to a clean, dry, wide mouthed glass test tube having no anticoagulant, further, this test tube was allowed to stand in slanting position (for 30 minutes) at room temperature for serum separation. The collected serum was kept in refrigerator for its quantitative estimation of biochemical parameters (Blood glucose, Total serum protein, Blood urea nitrogen, Serum creatinine, Alanine transaminase) by using blood Auto analyzer (Model: EM 360) furthermore, Serum Sodium, Potassium were estimated using Easylyte plus (K/Na/Cl analyzer).

Statistical analysis: Statistical difference between respective means for various parameters was evaluated using tow way Anova as per the method described by Snedecor and Cochran (2004) [18].

Result and Discussion

Hematological observation

An increase in hemoglobin values (g/dl) of *colibacillosis* affected calves was observed among all the treatment group-A (13.250±2.962 g/dl), group-B (13.365±2.989 g/dl) and group-C (13.490±3.016 g/dl) on 0 day (pre-treatment) in comparison to healthy control calves (10.733±0.532 g/dl). Following different treatment regimen an increased hemoglobin followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration of normal hemoglobin within the same treatment group throughout the study periods. A normal hemoglobin level in the entire treatment group was achieved on 8th days of observation as 10.905±2.438 g/dl, 11.115±2.485 g/dl and 10.935±2.445 g/dl in the treatment group-A, group-B and group-C respectively. The haemocontraction that have occurred in *colibacillosis* affected calves might be due to excessive loss of fluid through faeces which finally leads to hemoconcentration as well as decrease in per unit of blood volume. Hence due to decrease in blood volume and dehydration there is increase in hemoglobin values (Jain 1986; Santos *et al.*, 2002; Ghanem *et al.*, 2012; Kumar and Mandial, 2002) [9, 10, 11, 12]. These observation recorded are in agreement with Kumar and Mandial (2002) [12], Tikoo and soodan (2009) [13], Asati *et al.*, (2010) [14], Kumar *et al.*, (2010) [15], Tarunpreet *et al.*, (2018) [16] and Nayak *et al.*, (2019) [17].

An increase in TEC ($10^6/\mu\text{l}$) values in *colibacillosis* affected calves was observed among all the treatment group-A ($9.139 \pm 0.290 \times 10^6/\mu\text{l}$), group-B ($9.265 \pm 0.340 \times 10^6/\mu\text{l}$) and group-C ($8.861 \pm 0.401 \times 10^6/\mu\text{l}$) on 0 day (pre-treatment) in comparison to healthy control calves $7.033 \pm 0.195 \times 10^6/\mu\text{l}$.

Following different treatment regimen an increased TEC followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration of normal TEC within the same treatment group. An early achievement in normal TEC value was noted on 6th days of observation in group-C ($7.668 \pm 0.342 \times 10^6/\mu\text{l}$) whereas in group-A ($7.294 \pm 0.226 \times 10^6/\mu\text{l}$) and group-B ($7.831 \pm 0.309 \times 10^6/\mu\text{l}$) it was achieved on 8th days of observation. The main reason of increased in TEC values in *colibacillosis* affected calves might be the consequence of fluids loss which ultimately derived to hemoconcentration (Reddy, 1975; Boyd *et al.*, 1974) [18, 19]. Our trends of results are in agreement with the findings of Tikoo and Soodan (2009) [13], Mir *et al.*, (2010) [20], Kumar *et al.*, (2010) [15], Hassan *et al.*, (2013) [21], Tarunpreet *et al.*, (2018) [16], Nayak *et al.*, (2019) [17] and Manu *et al.*, (2019) [22].

An increase in TLC ($10^3/\mu\text{l}$) of *colibacillosis* affected calves was observed in group-A ($14.795 \pm 1.132 \times 10^3/\mu\text{l}$), group-B ($14.278 \pm 0.861 \times 10^3/\mu\text{l}$) and group-C ($14.469 \pm 0.73 \times 10^3/\mu\text{l}$) on 0 day (pre-treatment) in comparison to healthy calves. Following different treatment regimen an increased TLC value followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration of normal TLC value ($10.550 \pm 0.670 \times 10^3/\mu\text{l}$) within the same treatment group throughout the study periods. A normal TLC value in the entire treatment group was achieved on 8th days of observation as $11.309 \pm 0.713 \times 10^3/\mu\text{l}$, $10.382 \pm 0.760 \times 10^3/\mu\text{l}$ and $10.733 \pm 0.561 \times 10^3/\mu\text{l}$ in the treatment group-A, group-B and group-C respectively. An increase in TLC values of *colibacillosis* affected calves might be occurred due to reaction of body defense mechanism against *E. coli* infection as well as dehydration that causes an increase in the viscosity of blood (Tiko and Soodan 2009; Fernandes *et al.*, 2009) [23]. Our results of variation in TLC are in concurrence with the findings of other scientists Asati *et al.*, (2010) [14], Mir *et al.*, (2010) [13, 20], Kumar *et al.*, (2010) [15], Hassan *et al.*, (2013) [21], Mehesara (2018) [24], Tsarunpreet *et al.*, (2018) [16], Nayak *et al.*, (2019) [17] and Manu *et al.*, (2019) [22].

Biochemical observation

A significant decrease in the blood glucose level (mg/dl) of *colibacillosis* affected calves among all the treatment group-A ($38.625 \pm 0.791 \text{ mg/dl}$), group-B ($38.525 \pm 0.429 \text{ mg/dl}$) and group-C ($39.720 \pm 0.887 \text{ mg/dl}$) on 0 day (pre-treatment) in comparison to healthy control calves $56.466 \pm 1.493 \text{ mg/dl}$. Following different treatment regimen the level of blood glucose followed a significant ($p \leq 0.01\%$) increasing trend towards restoration of normal level within the same treatment group. The normal level of blood glucose was not achieved till end of treatment in all the groups-A, group-B and group-C. However, a maximum increased was observed in group-C $52.560 \pm 1.063 \text{ mg/dl}$ than group-B ($51.890 \pm 0.859 \text{ mg/dl}$) and group-A ($48.440 \pm 0.428 \text{ mg/dl}$) on 8th days of observation. The probable reason of decrease in blood glucose level (Hypoglycemia) in *colibacillosis* calves might be associated with anorexia, decreased intestinal absorption of glucose and reduce rate of conversion of lactic acid to glucose (Morris *et al.*, 1985) [25]. Our results of variation in blood glucose leves are in agreement with findings of Asati *et al.*, (2010) [14], Pal and Pachuri (2011) [26], Mehesare (2018) [24] and Tarunpreet *et al.*, (2018) [16].

An increase in mean values of blood urea nitrogen (mg/dl) of *colibacillosis* affected calves among the entire treatment group-A ($28.910 \pm 1.061 \text{ mg/dl}$), group-B ($30.547 \pm 0.819 \text{ mg/dl}$) and group-C ($30.836 \pm 0.912 \text{ mg/dl}$) on 0 day i.e. before

start of treatment in comparison to healthy control animal (20.636±0.243 mg/dl). Following different treatment regimen an increase in blood urea nitrogen followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration of normal level within the same treatment group throughout the study periods. A nearer in normal blood urea nitrogen level was achieved on 8th days of observation. However, a maximum decreased was observed in group-C (21.927±0.813mg/dl) than group-B (22.298±0.811m/dl) and group-A (24.338±1.020mg/dl). The reason behind elevation in BUN level of *colibacillosis* affected calves might be due to catabolism of protein, fat and carbohydrates to generate water (Radiostitis *et al.*, 2007) [27]. These results are in agreement with the findings of Chagkja (2002) [28], Seifi *et al.*, (2006) [29], Jesse *et al.*, (2016) [30] and Mehesare (2018) [24].

An increase in mean values of total serum protein (g/dl) of *colibacillosis* affected calves among all the treatment group-A (8.804±0.117 g/dl), group-B (8.446±0.415 g/dl) and group-C (8.013±0.259 g/dl) on 0 day (pre-treatment) in comparison to healthy control calves (7.223±0.197 g/dl). Following different treatment regimen an increased total serum protein followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration in normal total serum protein within the same treatment group. An early achievement of normal total serum protein level was noted in group-C (7.266±0.202 g/dl) than group-B (7.269±0.386 g/dl) on 6th days of observation, whereas it was achieved on 8th days of observation in group-A (7.453±0.309 g/dl). An increase in the mean values of total serum protein of *colibacillosis* affected calves might be due to dehydration which causes reduction in fluid volume leading to haemo-concentration as a result hyper-proteinaemia occurs (Kaneko 1989) [31]. Our findings of result are in similar trends with the findings of Asati *et al.*, (2010) [14], Mir *et al.*, (2010) [20], Singh *et al.*, (2014) [32], Tarunpreet *et al.*, (2018) [16] and Manu *et al.*, (2019) [22].

An increase in mean values of serum creatinine level (mg/dl) was observed among all the treatment group-A (2.382±0.090 mg/dl), group-B (2.30±0.087 mg/dl) and group-C (2.280±0.075 mg/dl) on 0 day (pre-treatment) in comparison to healthy control calves 1.323±0.149 mg/dl. Following different treatment regimen an increased serum creatinine level followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration of normal serum creatinine level within the same treatment throughout the study periods. A normal serum creatinine level as in group-B (1.354±0.061mg/dl) and group-C (1.363±0.071mg/dl) was noted on 8th days of observation whereas in group-A a nearer normal level (1.634±0.111mg/dl) was achieved till end of the treatment. An increase in serum creatinine values of *colibacillosis* affected calves could be due to insufficient perfusion pressure through kidneys which have occurred due to reduced blood volume as a result of severe dehydration (Radiostitis *et al.*, 2007) [27]. Similar types of findings during present research work in *colibacillosis* affected calves are in corroboration with Changkja (2002) [28], Seifi *et al.*, (2006) [29], Jesse *et al.*, (2016) [30] and Mehesare (2018) [24].

An increase in mean values of ALT level in *colibacillosis* affected calves was observed among all the treatment group-A (18.590±0.44 IU/L), group-B (18.030±0.430 IU/L) and group-C (18.560±0.799 IU/L) on 0 day (pre-treatment) in comparison to the values of healthy control calves 17.296±0.825 IU/L. Following treatment on subsequent days a non-significant difference was noticed within and between different treatment groups, however in different treatment

regimen, the level of ALT showed decreasing trend in both the herbal treatment group-B and group-C and near normal level such as 17.360±0.455 IU/L and 17.580±0.819 IU/L was archived in both these group-B and group-C respectively. This decrease in ALT level was more in herbal treatment group then the antibiotics treatment group-A which remains to 18.320±0.498 IU/L at the end of treatment i.e. on 8th days of observation. In the present study a non-significant change in ALT activities has indicated the absence of marked hepatic damage (Lewis *et al.*, 1975) [33]. Our findings of ALT are in corroboration with the findings of Lewis *et al.*, (1975) [33], Singh *et al.*, (2014) [32] and Shekhar *et al.*, (2017) [34].

A decrease in mean values of serum sodium concentration (mEq/L) of *colibacillosis* affected calves was observed among all the treatment group-A (118.414±0.865 mEq/L), group-B (118.769±0.585 mEq/L) and group-C (119.927±0.740 mEq/L) on 0 day (pre-treatment) in comparison to healthy control calves 127.913±2.800 mEq/L. Following different treatment regimen a decreased serum sodium level followed a significant ($p \leq 0.01\%$) increasing trends towards restoration of normal serum sodium level within the same treatment group. An early achievement in normal serum sodium level was noted on 4th days of observation in group-C (127.071±0.670 mEq/L) whereas in group-A (127.632±0.623 mEq/L) and group-B (128.323±0.536 mEq/L) it was achieved on 8th days of observation. The reason of decrease in the mean values of serum sodium concentration of *colibacillosis* affected calves might due to loss of sodium bicarbonate (normally present in intestinal contents) through faeces which ultimately leads to fall in sodium ion concentration (Benjamin1985) [35]. Our findings of results are in agreement with the findings of Asati *et al.*, (2010) [14], Mir *et al.*, (2010) [20], Mehesare (2018) [24], Tarunpreet *et al.*, (2018) [16] and Manu *et al.*, (2019) [22].

An increase in mean values of serum potassium concentration (mEq/L) of *colibacillosis* affected calves was observed among all the treatment group-A (6.725±0.243 mEq/L), group-B (6.845±0.180 mEq/L) and group-C (6.795±0.222 mEq/L) on 0 day (pre-treatment) in comparison to healthy control calves 4.953±0.881. Following different treatment regimen an increased serum potassium level followed a significant ($p \leq 0.01\%$) decreasing trends towards restoration of normal serum potassium level within the same treatment group throughout the study periods. A normal serum potassium level among the entire treatment group was achieved on 8th days of observation as 5.110±0.226 mEq/L, 5.075±0.123 mEq/L and 4.710±0.195 mEq/L in the treatment group-A, group-B and group-C respectively. The main reason of increase in the mean values of serum potassium concentration of *colibacillosis* affected calves might be due to metabolic acidosis, which might have stimulates the transfer of potassium ion from intracellular space to extracellular space because excess hydrogen ions are buffered intra-cellularly, therefore potassium ion along with sodium ion leave to maintain neutrality (Brobst 1986) [36]. The results of present findings are in close agreement with the findings of Asati *et al.*, (2010) [14], Mir *et al.*, (2010) [20], Mehesare (2018) [24], Tarunpreet *et al.*, (2018) [16] and Manu *et al.*, (2019) [22].

Conclusions

After, critical analyzing the various features of *colibacillosis*, it was concluded that treatment with *Psidium guajava* leaf extract@ 40mg/kg b.wt. for upto 7th days can prove to be more beneficial than *Punica granatum* rind extract and Ofloxacin in achieving 100% recovery in hemato-biochemical

changes that have occurred due to *colibacillosis* in calves.

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References

- Radostits OM, Gay CC, Blood DC, Hinchcliff KW. Veterinary Medicine: A Textbook of the Diseases of Cattle, Sheep, Pigs, Goats and Horses. WB Saunders, New York, USA 2000.
- Izzo MM, Kirkland PD, Mohler VL, Perkins NR, Gunna AA, House JK. Prevalence of major enteric pathogens in Australian dairy calves with diarrhoea. Australian Veterinary Journal 2011;89:167-73.
- Mero KN, Rollin RE, Phillips RW. Malabsorption due to selected oral antibiotics, Veterinary Clinics North American Food Animal Practice 1985;1:581-588.
- Lutterodt GD. Inhibition of gastrointestinal release of acetylcholine by quercetin as a possible mode of action of *Psidium guajava* leaf extracts in the treatment of acute diarrhoeal diseases. Journal of Ethnopharmacology 1989;25:235-247.
- Alneida CC, Karnikowski MG, Flieto R, Baldisserotto B. Analysis of antidiarrhoic effect of plants used in popular medicine. Revista De Saude Publica 1995;29:428-433.
- Soulsby E.J.L. Helminths, Arthropods and Protozoan of Domesticated Animals. 7th Ed. (ELBS) Bailliere Tindall, London 1982.
- Stokes EJ, Ridgway GL, Wren MWD. Clinical Microbiology, 7th Edn., Jhon Wiley & Sons, London 1993, 101.
- Snedecor GW, Cochran WG. Statistical Methods. 8th Edn. Oxford and IBH Publishing Co. Pvt. Ltd., Kolkata 2004.
- Jain NC. In: Schalm's Veterinary Haematology. 4th Edn. Lea and Febiger, Philadelphia 1986.
- Santos RL, Tsohis RM, Baumler AJ, Adams G. Haematologic and serum biological changes in *Salmonella typhimurium* infected calves. American Journal of Veterinary Research 2002;63(8):1145-1150.
- Ghanem MM, Fkhrany SF, Abd EI-Roaf YM, EI-Attar HM. Clinical and haematological evaluation of diarrheic neonatal buffalo calves (*Bubalus Bubalis*) with reference to antioxidant changes. Benha veterinary medical journal 2012;23(2):275-288.
- Kumar R, Mandial RK. Clinico-biochemical and therapeutic studies on clinical *colibacillosis* in crossbred calves. Indian Vet. J 2002;79:672-676.
- Tikoo A, Soodan JS. *Colibacillosis* in neonatal calves and its management. Intas Polivet 2009;10(2):212-213.
- Asati CK, Roy S, Roy M. Evaluation of oral rehydration solution and amoxicillin-salbactam treatment of colibacillosis in calves. Indian Vet. J 2010;87(5):454-456.
- Kumar BK, Shekher P, Kumar N. A clinical study on neonatal calf diarrhoea. Intas Polivet 2010;11(2):233-235.
- Tarunpreet SK, Sharma Singh AP, Deepika Goklaney. Prevalence of colibacillosis disease and clinico-haemato biochemical changes in lambs in Southern part of Rajasthan. Veterinary Practitioner 2018;20(1):95-99.
- Nayak TC, Singh Savita AP, Kachhawa JP Gupta SR, Yadav R. A study on alteration in clinico-haematological parameters in colibacillosis affected diarrhoeic cattle calves. International Journal of Chemical Studies 2019;7(5):1559-1562.
- Reddy V Raghuma. Clinical and therapeutic studies on calf scours. M.V. Sc. Thesis, G.B. Pant Univ. of Agril. and Tech. Pantnagar 1975, 73.
- Boyd JW, Baker JR, Leyland A. Neonatal diarrhea in calves. Vet. Rec. 1974;95:310-313.
- Mir N, Shukla PC, Baghel RPS, Dixit P, Saroori AR. Efficacy of baelpulp in calf diarrhoea with rehydration therapy. Veterinary Practitioner 2010;11(1):63-65.
- Hassan N, Sheikh GN. Prevalence of colibacillosis disease in lambs in Kashmir valley. Indian J Anim. Sci 2013;83(8):784-785.
- Manu Jaiswal PC, Shukla Alok Mishra, Preeti Bisht. Clinical score and haemato-biochemical alterations in acute diarrhoea in calves. The Pharma Innovation Journal 2019;8(2):529-532.
- Fernandes CE, Roy K, Shukla PC, Rao MLV. Changes in the haematological profile in calf diarrhoea in response to therapy. Intas Polivet 2009;10(2):214-215.
- Mehesare SS. Phytopharmacological and therapeutic study of polyherbal antidiarrhoeal formulation in goats. Ph.D. Thesis, submitted to Maharashtra Animal and Fishery Science University, Nagpur 2018.
- Morris JA, Weils GAH, Scott AC, Sojka WJ. A comparison of methods for demonstrating colonization in the small intestine of piglets by enterotoxigenic *E. coli*. Br. Vet. J 1985;141:484-489.
- Pal B, Pachauri SP. Comparative efficacy of oral rehydration solutions for restoration of plasma biochemical changes in neonatal diarrhoeic calves. 11th Indian Veterinary Congress and XVIII Annual Conference of Indian Association for Advancement of Veterinary Research and National Symposium on Veterinary science and education on move: critical gaps and needs held at Jaipur, India 2011.
- Radostits OM, Gay CC, Hinchcliff KW, Constable PD. Veterinary Medicine, A textbook of the diseases of cattle, sheep, goats, pigs and horse. 10th ed. Elsevier Saunders, London, UK 2007.
- Changkija B. Clinico-biochemical and therapeutic studies of calf diarrhoea. M.V. Sc. thesis, IVRI, Deemed University, Izatnagar 2002.
- Seifi HA, Mahri M, Shoorei E, Farzaneh N. Using haematology and serum biochemical findings as prognostic indicators in calf diarrhoea. Comparative Clinical Pathology 2006;15:143-147.
- Jeese FFASM, Abba Y, Chung ELT, Adamu L, Bitrus AA, Hambali DU. Clinico-Pathological Findings of Septicaemic *Colibacillosis* in a Calf. J. Dairy Vet. Anim. Res 2016, 4(3).
- Kaneko JJ. Clinico-Biochemistry of Domestic Animals. 4th Ed. Department of Clinical pathology school of Vet. Med. University of California, Davis, California 1989.
- Singh M, Gupta KV, Mandal BD, Bansal KS, Sharma KD, Shakya M *et al.* A study on alteration in Haematological parameter in *colibacillosis* affected calves. International Journal of Advance Research 2014;2(7):746-750.
- Lewis LP, Phillip RW, Elliot CD. Change in plasma glucose and lactate concentration and enzyme activities

- in the neonatal calf with diarrhea. *Am. J. Vet. Res.* 1975;36:413-16.
34. Shekhar S, Ranjan R, Singh CV, Kumar P. Prevalence, Clinicohaemato-Biochemical alterations in colibacillosis in neonatal Calves. *International Journal of Current Microbiology and Applied Sciences* 2017;6:3192-3198.
 35. Benjamin MM. *Outline of Veterinary Clinical Pathology*. 1edn. Kalyani Publications, New Delhi 1985, 109.
 36. Brobst D. Review of pathophysiology of alterations in potassium homeostasis. *J. Am. Vet. Med. Ass.* 1986;188:1019-1025.