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Antihyperlipidemic activity of isoflavones

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Abstract

Cyclocarya paliurus has been used commonly to treat diabetes in China. However, the effective components and the effect of plant origin remain unclear. In this study, *C. paliurus* leaves with different chemical compositions were selected from five geographical locations, and their effects on streptozotocin (STZ)-induced diabetic mice were evaluated with both ethanol and aqueous extracts. Glucose levels, lipid levels, and biomarkers of liver and kidney function were measured. Results showed that *C. paliurus* extracts with better antihyperglycemic effects were characterized by higher contents of total flavonoids, especially quercetin-3-O-glucuronide and kaempferol-3-O-glucuronide. Furthermore, significantly negative correlations were found between triterpenoids contents and lipid levels. The anti-hyperlipidemic effect of flavone-rich *B. chinensis* leaf extract (HTP) in ICR mice fed a high-fat diet. Treatment with HTP significantly decreased peri-epididymal fat weight ($p < 0.01$ and $p < 0.05$ for 200 and 100 mg/kg, respectively), lowered serum and hepatic lipid, and decreased glucose area under curve (AUC) in oral glucose tolerance test ($p < 0.01$ for 200 mg/kg). Western blot and ELISA analysis showed that administration of HTP (200 mg/kg) significantly increased AMPK ($p < 0.05$) phosphorylation and PPAR α expression in liver ($p < 0.05$).

Keywords: *Cyclocarya paliurus*, diabetes, flavonoids, triterpenoids, geographical locations, extraction solvents, *Belamcanda chinensis*, Flavone, Hyperlipidemia, Adenosine 5'-monophosphate-activated protein kinase, Peroxisome proliferator activated receptor-alpha, Glucose tolerance

Introduction

Cyclocarya paliurus (Batal) Iljinskaja belongs to the Juglandaceae family and is widely distributed in mountainous regions of sub-tropical China [8]. Leaves of this plant are traditionally used in China as an ingredient in nutraceutical tea or drug formulations for the treatment of hypertension, diabetes mellitus, and hyperliposis [9-12]. Previous studies have reported that *C. paliurus* leaves contain abundant bioactive components including polysaccharides, triterpenoids, flavonoids, and phenolic compounds [13-15]. Thus, chemical identification and the anti-diabetic effect of *C. paliurus* extracts have attracted the attention of many scholars, however the constituents responsible for anti-diabetic effect still remains controversial. For example, previous studies have revealed that the water extract of *C. paliurus* could reduce postprandial triglyceride level in hyperlipidemic mice, and predicted polysaccharides to be the active components [16, 17]. Polysaccharides of *C. paliurus* are also found to exhibit a lipid-lowering effect on high-fat-diet-induced rats [18, 19].

However, it is demonstrated that polysaccharides did not appear to be the active anti-diabetic constituent, based on the comparison of anti-diabetic effects of *C. paliurus* ethanol extract (without polysaccharides) and aqueous extract in streptozotocin (STZ)-induced diabetic rats [20]. In addition, several studies have shown that triterpenic acid-enriched *C. paliurus* fraction or extracts possess better antihyperlipidemic activities in mice fed with high-fat-diet [21-23]. Thus, further investigation of the anti-diabetic activities of *C. paliurus* extracts with detailed component information is required. To fill the knowledge gap mentioned above, in this study, *C. paliurus* leaves with different chemical compositions were selected from five geographical locations and their effects on streptozotocin (STZ)-induced diabetic mice were evaluated with both ethanol and aqueous extracts. Principal components analysis (PCA) and canonical correspondence analysis (CCA) were used to analyze the chemical variability and its relationship with anti-diabetic activities. The potential antihyperglycemic capacity of *C. paliurus* flavonoids and the antihyperlipidemic effect of *C. paliurus* triterpenoids were revealed. Taken together, this study provides essential information supporting the use of *C. paliurus* extract as a natural medicine for diabetic patients.

Belamcanda chinensis belongs to the family of Iridaceae and their rhizomes have been widely used as traditional medicine in China.

Many isoflavones such as tectoridin, iridin, irigenin and tectorigenin have been identified from this plant [14]. The antifebrile, antioxidant, antiinflammatory and hepatoprotective activities of *B. chinensis* have been well-documented [15-17]. Previously, we reported the hypoglycemic and anti-hyperglycemic effects of flavone-rich *B. chinensis* leaf extract (HTP) in normal and STZ induced diabetic rats [18, 19]. Apart from these, no study to-date has investigated the antihyperlipidemia activity of HTP. In this study, we assessed the anti-hyperlipidemia effects of HTP on high-fat diet induced obese mice. The stimulating effects of HTP on AMPK and PPAR α were also studied.

Material and Methods

Plant Materials

Five geographical locations from five provinces in China for *C. paliurus* sampling were identified based on the major distribution of the species. Voucher specimens were deposited in Silviculture Lab of Nanjing Forestry University (Voucher code: 2011GX, 2011SC, 2011HB, 2011ZJ, 2011HN). The detailed geographic, climatic information, and soil index of the sample locations are listed in Table 1. Leaf samples were collected from each location in September 2014. At each location, 6–30 trees (generally dominant or co-dominant tree in the stand) were selected based on tree age (over 20 years-old), stem form, and growth vigor. Number of trees for collecting leaves for each location was determined according to quantity and stand area of *C. paliurus* which distributed on the area (about 10% of the total). About 400 g fresh fully developed leaves were collected from the middle crown for each tree and then sealed up with silica gel for transportation. Leaves collected in each location were mixed together in the lab and then dried at 70 °C for 48 h to constant weight, and ground into fine powder before extraction. All samples were stored at room temperature prior to analysis.

Chemical reagents and References

Streptozotocin (STZ) was purchased from Sigma Chemical Co. (St. Louis, MO, USA). Metformin Hydrochloride Tablets (MHT) was bought from Sino-American Shanghai Squibb Pharma (Shanghai, China), and Xiaoke Pill (XKP) from Guangzhou Zhongyi Pharmaceutical Enterprise (Guangdong, China). Commercial test kits used for measuring TG, TC, LDL-c, HDL-c, BUN, CREA, TBIL, AST, and ALT were purchased from Jiancheng Institute of Biotechnology (Nanjing, China). The reference standards of 3-O-caffeoylquinic acid, 4-O-caffeoylquinic acid, isoquercitrin, 4,5-di-O-caffeoylquinic acid, kaempferol-3-O-glucuronide, quercetin-3-O-glucuronide, arjunolic acid, quercetin-3-O-rhamnoside, quercetin-3-O-galactoside, and kaempferol-3-O-glucoside (purity >98%) were purchased from BioBioPha Co., Ltd. (Kunming, China), and arjunolic acid, oleanolic acid (purity >98%) were purchased from Shanghai Yuanye Biotechnology Co., Ltd. (Shanghai, China), whereas cyclocaric acid B, kaempferol-3-O-rhamnoside, and pterocaryoside A and pterocaryoside B (purity >98%) were isolated and purified from the laboratory of China Pharmaceutical University (Nanjing, China) [24].

Plant materials

Leaves of *Belamcanda chinensis* were collected from Hainan province, South China, and authenticated by Dr Hubiao Chen (Health Science Center, Peking University, China). The

preparation of HTP was performed as previously reported [18, 19]. A voucher specimen has been deposited in School of Life Science, Beijing Institute of Technology (NO. HTP20120911).

Animals and experiment design

Male ICR mice (20 \pm 2 g, Peking University Laboratory Animal Center, Beijing, China) were housed at 22 \pm 2 °C and 55 \pm 5% relative humidity; 12 h light-dark cycle and allowed free access to water and feed. The study was carried out in accordance with International Guidelines for Care and Use of Laboratory Animals [20] and approved by Animal Ethical Committee of Beijing Institute of Technology (reg. no. 201209007/BITAEC). Seventy-two ICR mice were kept in a week. Animals were then randomly divided into 6 groups with 12 mice in each group: BC, fed a standard diet and received 0.5% sodium carboxy methyl cellulose (CMC-Na) solution only; NC, fed a humidity controlled room on a 12-h light-dark cycle with food and water available ad libitum for one high-fat diet (HFD) and received 0.5% CMC-Na solution only; PC, fed a HFD and treated with gemfibrozil (200 mg/kg); and HTP groups, fed a HFD and treated with HTP (100, 200 and 400 mg/kg, respectively). HFD consists of 10% lard, 10% sugar, 10% egg York, 1% cholesterol, 0.2% sodium cholate, and 68.8% standard diet. HTP and gemfibrozil were administered by oral gavage. Body weight was monitored weekly. During the experiment, blood was collected from tail vein for the measurement of blood glucose. At the end of the 7th week, blood sample was collected from orbital venous plexus and the serum was prepared for biochemical analysis. Liver and peri-epididymal fat were removed, weighted and stored at -70 °C refrigerator (Thermo Fisher Scientific, Waltham, USA.). Serum and hepatic levels of triglyceride (TG) and total cholesterol (TCH) were determined by corresponding kits (Nanjing Jiancheng Bioengineering Institute, Nanjing, China) according to the manufacturer's instruction. The hepatic PPAR α level was determined by a mouse PPAR- α ELISA kit (HanKe Biotech co., Ltd, Beijing, China) according to the manufacture's instruction.

Oral glucose tolerance test (OGTT)

Animals were orally administered corresponding reagents after overnight fast. PC group was treated with acarbose (50 mg/kg). After another 2 h, glucose (2.5 g/kg) was given to each animal orally. Blood were collected from tail vein of each mouse at 0, 30, 60, and 120 min after glucose administration, and glucose levels were determined by a blood glucose meter (Roche Diagnostics, Basel, Switzerland).

Western blot

Western blot analysis was performed on liver tissue extract as previously reported [21]. Antibodies against phospho-AMPK α (Thr172) and AMPK α were from Cell Signaling Technology Inc. (Beverly, USA), and antibody against GAPDH was from Abcam, Inc. (Cambridge, USA).

Statistical analysis

Data are presented as the means \pm S.D. Oneway ANOVA was used to determine significant differences among groups, after which the modified Student's t-test with the Bon ferroni correction was used for comparison between individual groups. $P < 0.05$ was considered statistically significant.

Results and Discussion

Table 1: Index of liver and kidney function in different groups of experimental mice at the end of the diabetic trial (mean \pm SD). Different letters indicate significant differences ($p < 0.05$ by Duncan's test) between treatments ($n = 6$).

Treatment	ALT (U/L)	AST (U/L)	TBIL (μ mol/L)	CREA (μ mol/L)	BUN (mmol/L)
DC	52.8 \pm 2.17 a	145.6 \pm 3.08 a	7.50 \pm 0.89 a	25.8 \pm 1.17 a	17.3 \pm 0.99 a
NC	28.0 \pm 1.84 e	99.8 \pm 6.87 fg	1.00 \pm 0.18 g	14.7 \pm 1.36 e	9.80 \pm 0.18 d
EL1	36.1 \pm 2.63 cd	113.3 \pm 8.10 def	1.20 \pm 0.15 fg	20.8 \pm 0.75 bcd	13.4 \pm 0.86 b
EL2	35.9 \pm 1.80 cd	94.8 \pm 8.59 g	1.02 \pm 0.02 g	19.2 \pm 1.94 cd	12.6 \pm 1.08 bc
EL3	35.1 \pm 3.36 d	94.5 \pm 4.45 g	1.19 \pm 0.18 fg	18.2 \pm 2.13 d	13.1 \pm 1.26 bc
EL4	36.9 \pm 3.35 bcd	95.8 \pm 9.05 g	1.46 \pm 0.16 fg	21.0 \pm 0.89 bcd	11.0 \pm 1.17 cd
EL5	36.2 \pm 3.38 cd	114.0 \pm 7.66 de	1.38 \pm 0.18 fg	21.0 \pm 1.26 bcd	12.7 \pm 0.26 bc
AL1	40.3 \pm 4.51 bcd	130.2 \pm 4.56 b	1.47 \pm 0.18 fg	21.7 \pm 1.86 bc	14.7 \pm 0.81 b
AL2	43.4 \pm 6.36 b	128.9 \pm 5.30 bc	2.03 \pm 0.10 ef	22.3 \pm 1.21 b	14.2 \pm 0.69 b
AL3	41.6 \pm 1.36 bcd	123.6 \pm 6.69 bcd	2.57 \pm 0.28 e	22.8 \pm 0.98 b	14.3 \pm 0.92 b
AL4	43.1 \pm 5.16 b	125.8 \pm 4.16 bcd	4.87 \pm 0.17 bc	21.8 \pm 1.17 bc	14.2 \pm 1.17 b
AL5	40.0 \pm 3.19 bcd	115.5 \pm 6.08 cde	3.63 \pm 0.21 d	21.2 \pm 0.98 bc	14.2 \pm 1.32 b
MHT	42.4 \pm 2.06 bc	121.1 \pm 6.06 bcde	5.60 \pm 0.55 b	21.0 \pm 1.67 bcd	14.3 \pm 0.76 b
XKP	43.5 \pm 2.39 b	108.2 \pm 8.57 efg	4.41 \pm 0.22 cd	22.0 \pm 1.79 bc	13.9 \pm 0.82 b

The development of diabetes often leads to the leakage of the circulatory system, and high levels of AST, ALT, TBIL, CREA, and BUN were often observed in STZ-induced diabetic mice [36, 37], which is in agreement with our results (Table 2). Results indicated that *C. paliurus* extracts improved the liver and kidney function by decreasing the serum AST, ALT, and TBIL, CREA, and BUN levels in the diabetic mice. Moreover, significantly positive correlations were found between the values of AST, ALT, CREA, BUN, and lipid index (TC, TG, and LDL-c), which is in agreement with other reports in hyperlipidemic mice [38, 39]. Overall, these results

suggested that *C. paliurus* extracts could offer protection to liver and kidney by effectively lowering the lipid level. These findings, together with evidence collected from previous reports suggest that the composition of *C. paliurus* compounds might help for the design of therapeutic alternatives for the treatment of diabetes mellitus. However, geographic origins and the extraction solvents can also affect the effectiveness of the treatment as these factors influence the chemical compositions and their antihyperglycemic and antihyperlipidemic activities.

Table 2: Effect of HTP on the body weight, peri-epididymal fat weight and fat index

Group (n=12)	Body weight (g)	Peri-epididymal fat weight (g)	Fat index (mg/g BW)
BC	44.8 \pm 4.3	0.40 \pm 0.16**	8.14 \pm 2.62**
NC	46.1 \pm 3.0	0.83 \pm 0.30##	18.07 \pm 6.74##
PC	43.1 \pm 2.3*	0.55 \pm 0.18*	12.70 \pm 3.84
HTP100	44.8 \pm 3.9	0.59 \pm 0.42	9.57 \pm 3.15**
HTP 200	44.8 \pm 3.6	0.39 \pm 0.25**	8.62 \pm 5.33**
HTP 400	45.4 \pm 2.8	0.57 \pm 0.29*	11.13 \pm 5.06*

BC = fed a standard diet; NC = fed a high fat-diet (HFD); PC = fed a HFD and treated with gemfibrozil (200 mg/kg); HTPs = fed a HFD and treated with indicated dose of HTP. Values are mean \pm S.D.; ## $p < 0.01$ NC vs. BC; * $p < 0.05$, ** $p < 0.01$ vs. NC.

Animals in all groups showed a steady increase in body weight throughout the experimental period. At the end of the study, the average body weight of NC group was significantly higher than BC group ($p < 0.05$). Treating with HTP for 7 weeks did not influence the weight gain but significantly decreased the peri-epididymal fat weight and fat index (Table 2).

Conclusion

In conclusion, based on the analysis of chemical variability and its relationship with anti-diabetic activities, our results revealed the potential antihyperglycemic capacity of *C. paliurus* flavonoids and the antihyperlipidemic effect of *C. paliurus* triterpenoids, while these potential capacities were closely related to their contents. Further studies on the antihyperglycemic effect of *C. paliurus* kaempferol or quercetin glycosides and the antihyperlipidemic mechanism of *C. paliurus* triterpenoids will be carried out in future research.

The results presented in this study suggest that the flavone-rich *B. chinensis* leaf extract (HTP) has good potentials for

lipid management. Up regulation of AMPK and PPAR α are two possible mechanisms for its antihyperlipidemic activity.

References

- Jia W, Gao W, Tang L. Antidiabetic herbal drugs officially approved in China. *Phytother. Res.* 2003;17:1127-1134.
- Ahmad Aufa Z, Hassan FA, Ismail A, Mohd Yusof BN, Hamid M. Chemical compositions and antioxidative and antidiabetic properties of underutilized vegetable palm hearts from *Plectocomiopsis geminiflora* and *Eugeissona insignis*. *J Agric. Food Chem.* 2014;62:2077-2084.
- Jung M, Park M, Lee HC, Kang YH, Kang ES, Kim, S.K. Antidiabetic agents from medicinal plants. *Curr. Med. Chem* 2006;13:1203-1218.
- Dehghan-Kooshkghazi M, Mathers JC. Starch digestion, large-bowel fermentation and intestinal mucosal cell proliferation in rats treated with the α -glucosidase inhibitor acarbose. *Br. J Nutr* 2004;91:357-365.
- Umeno A, Horie M, Murotomi K, Nakajima Y, Yoshida Y. Antioxidative and antidiabetic effects of natural

- polyphenols and isoflavones. *Molecules* 2016;21:708.
6. Phan MAT, Wang J, Tang J, Lee YZ, Ng K. Evaluation of α -glucosidase inhibition potential of some flavonoids from *Epimedium brevicornum*. *Food Sci. Technol* 2013;53:492–498.
 7. Numonov S, Edirs S, Bobakulov K, Qureshi MN, Bozorov K, Sharopov F *et al.* Evaluation of the Antidiabetic Activity and Chemical Composition of Geranium collinum Root Extracts-Computational and Experimental Investigations. *Molecules* 2017;22:983.
 8. Fang S, Wang J, Wei Z, Zhu Z. Methods to break seed dormancy in *Cyclocarya paliurus* (Batal) Iljinskaja. *Sci. Hort* 2006;110:305-309.
 9. Liu Y, Fang S, Yang W, Shang X, Fu X. Light quality affects flavonoid production and related gene expression in *Cyclocarya paliurus*. *J Photoch. Photobio. B* 2018;179:66–73.
 10. Xie JH, Dong CJ, Nie SP, Li F, Wang ZJ, Shen MY, *et al.* Extraction, chemical composition and antioxidant activity of flavonoids from *Cyclocarya paliurus* (Batal.) Iljinskaja leaves. *Food Chem* 2015;186:97-105.
 11. Kurihara H, Asami S, Shibata H, Fukami H, Tanaka T. Hypolipemic effect of *Cyclocarya paliurus* (Batal) Iljinskaja in lipid-loaded mice. *Biol. Pharm Bull* 2003;26:383-385.
 12. Xie JH, Shen MY, Xie MY, Nie SP, Chen Y, Li C, *et al.* Ultrasonic-assisted extraction, antimicrobial and antioxidant activities of *Cyclocarya paliurus* (Batal.) Iljinskaja polysaccharides. *Carbohydr Polym* 2012;89:177-184.
 13. Liu Y, Qian C, Ding S, Shang X, Yang W, Fang S. Effect of light regime and provenance on leaf characteristics, growth and flavonoid accumulation in *Cyclocarya paliurus* (Batal) Iljinskaja coppices. *Bot. Stud* 2016;57:28.
 14. Fang S, Yang W, Chu X, Shang X, She C, Fu X. Provenance and temporal variations in selected flavonoids in leaves of *Cyclocarya paliurus*. *Food Chem* 2011;124:1382-1386.
 15. Wright M, Byrd J, Gao Y, Stubblefield J, Park H, Dunlap N. Isolation and structural clarification of triterpenes from *Cyclocarya paliurus*: Cyclocaric acid A and B. *Planta Med* 2014;80:PD19.
 16. Kurihara H, Fukami H, Kusumoto A, Toyoda Y, Shibata H, Matsui Y, *et al.* Hypoglycemic action of *Cyclocarya paliurus* (Batal.) Iljinskaja in normal and diabetic mice. *Biosci. Biotechnol. Biochem* 2013;67:877–880.
 17. Li S, Li J, Guan XL, Li J, Deng SP, Li LQ *et al.* Hypoglycemic effects and constituents of the barks of *Cyclocarya paliurus* and their inhibiting activities to glucosidase and glycogen phosphorylase. *Fitoterapia* 2011;82:1081-1085.
 18. Yang ZW, Ouyang KH, Zhao J, Chen H, Xiong L, Wang WJ. Structural characterization and hypolipidemic effect of *Cyclocarya paliurus* polysaccharide in rat. *Int. J Biol. Macromol* 2016;91:1073-1080.
 19. Hu WB, Zhao J, Chen H, Xiong L, Wang WJ. Polysaccharides from *Cyclocarya paliurus*: Chemical composition and lipid-lowering effect on rats challenged with high-fat diet. *J Funct. Foods* 2017;36:262–273.
 20. Wang Q, Jiang C, Fang S, Wang J, Ji Y, Shang X, *et al.* Antihyperglycemic, antihyperlipidemic and antioxidant effects of ethanol and aqueous extracts of *Cyclocarya paliurus* leaves in type 2 diabetic rats. *J Ethnopharmacol* 2013;150:1119-1127.
 21. Jiang C, Wang Q, Wei Y, Yao N, Wu Z, Ma Y, *et al.* Cholesterol-lowering effects and potential mechanisms of different polar extracts from *Cyclocarya paliurus* leave in hyperlipidemic mice. *J Ethnopharmacol* 2015;176:17-26.
 22. Ma Y, Jiang C, Yao N, Li Y, Wang Q, Fang S, *et al.* Antihyperlipidemic effect of *Cyclocarya paliurus* (Batal.) Iljinskaja extract and inhibition of apolipoprotein B48 overproduction in hyperlipidemic mice. *J Ethnopharmacol* 2015;166:286–296.
 23. Wu ZF, Meng FC, Cao LJ, Jiang CH, Zhao MG, Shang XL, *et al.* Triterpenoids from *Cyclocarya paliurus* and their inhibitory effect on the secretion of apolipoprotein B48 in Caco-2 cells. *Phytochemistry* 2017;142:76–84.
 24. Cao Y, Fang S, Yin Z, Fu X, Shang X, Yang W, *et al.* Chemical Fingerprint and Multicomponent Quantitative Analysis for the Quality Evaluation of *Cyclocarya paliurus* Leaves by HPLC-Q-TOF-MS. *Molecules* 2017;22:1927.
 25. Yang L, Wen KS, Ruan X, Zhao YX, Wei F, Wang Q. Response of Plant Secondary Metabolites to Environmental Factors. *Molecules* 2018;23:762.
 26. Zhang L, Hogan S, Li J, Sun S, Canning C, Zheng SJ, Zhou K. Grape skin extract inhibits mammalian intestinal α -glucosidase activity and suppresses postprandial glycemic response in streptozocin-treated mice. *Food Chem* 2011;126:466–471.
 27. Shehata AM, Quintanilla-Fend L, Bettio S, Singh CB, Ammon HPT. Prevention of multiple low-dose streptozotocin (MLD-STZ) diabetes in mice by an extract from gum resin of *Boswellia serrata* (BE). *Phytomedicine* 2011;18:1037–1044.
 28. Zhang M, Du N, Wang L, Wang X, Xiao Y, Zhang K, *et al.* Conjugated fatty acid-rich oil from *Gynostemma pentaphyllum* seed can ameliorate lipid and glucose metabolism in type 2 diabetes mellitus mice. *Food Funct* 2017;8:3696–3706.
 29. Mensah-Brown EPK, Al Rabesi Z, Shahin A, Al Shamsi M, Arsenijevic N, Hsu DK, *et al.* Targeted disruption of the galectin-3 gene results in decreased susceptibility to multiple low dose streptozotocin-induced diabetes in mice. *Clin. Immunol* 2009;130:83–88.
 30. Fang XK, Gao J, Zhu DN. Kaempferol and quercetin isolated from *Euonymus alatus* improve glucose uptake of 3T3-L1 cells without adipogenesis activity. *Life Sci* 2008;82:615–622.
 31. Mukhopadhyay P, Prajapati AK. Quercetin in anti-diabetic research and strategies for improved quercetin bioavailability using polymer-based carriers-a review. *RSC Advances* 2015;5:97547-97562.
 32. Yao Y, Chen F, Wang M, Wang J, Ren G. Antidiabetic activity of Mung bean extracts in diabetic KK-Ay mice. *J Agric. Food Chem* 2008;56:8869–8873.
 33. Somova LO, Nadar A, Rammanan P, Shode FO. Cardiovascular, antihyperlipidemic and antioxidant effects of oleanolic and ursolic acids in experimental hypertension. *Phytomedicine* 2003;10:115–121.
 34. Ejelonu OC, Elekofehinti OO, Adanlawo IG. *Tithonia diversifolia* saponin-blood lipid interaction and its influence on immune system of normal wistar rats. *Biomed. Phar* 2017;87:589–595.
 35. James DB, Elebo N, Sanusi AM, Odoemene L. Some biochemical effect of intraperitoneal administration of *Phyllanthus amarus* aqueous extracts on normoglycemic

- albino rats. *Asian J Med. Sci* 2010;2:7–10.
36. Kew MC. Serum aminotransferase concentration as evidence of hepatocellular damage. *Lancet* 2000;355:591-592.
 37. Ostfeld R, Spinelli M, Mookherjee D, Holtzman D, Shoyeb A, Schaefer M, *et al.* The association of blood urea nitrogen levels and coronary artery disease. *Einstein J Biol. Med* 2016;25:3–7.
 38. Cao J, Wang S, Yao C, Xu Z, Xu X. Hypolipidemic effect of porphyran extracted from *Pyropia yezoensis* in ICR mice with high fatty diet. *J Appl. Phycol* 2016;28:1315-1322.
 39. Shi F, Li J, Yang L, Hou G, Ye M. Hypolipidemic effect and protection ability of liver-kidney functions of melanin from *Lachnum YM226* in high-fat diet fed mice. *Food Funct* 2018;9:880–889.
 40. Wu C, Wang X, Wang H, Shen B, He X, Gu W, *et al.* Extraction optimization, isolation, preliminary structural characterization and antioxidant activities of the cell wall polysaccharides in the petioles and pedicels of Chinese herbal medicine Qian (*Euryale ferox* Salisb.). *Int. J Biol. Macromol* 2014;64:458–467.
 41. Srinivasan K, Viswanad B, Asrat L, Kaul CL, Ramarao P. Combination of high-fat diet-fed and low-dose streptozotocin-treated rat: a model for type 2 diabetes and pharmacological screening. *Pharmacol. Res* 2005;52:313–320.
 42. Raza J, Babb J, Movahed A. Optimal management of hyperlipidemia in primary prevention of cardiovascular disease. *Int. J Cardiol* 2004;97:355366.
 43. Patel A, Barzi F, Jamrozik K, Lam TH, Ueshima H, Whitlock G, *et al.* Serum triglycerides as a risk factor for cardiovascular diseases in the Asia Pacific region. *Circulation* 2004;110:2678-2686.
 44. Boullart A, Graaf J, Stalenhoe A. Serum triglycerides and risk of cardiovascular disease. *Biochim Biophys Acta* 2012;1821:867-875.
 45. Sinzinger H, Wolfram R, Peskar BA. Muscular side effects of statins. *J Cardiovasc Pharmacol* 2002;40:163-171.
 46. Rosenbaum D, Dallongeville J, Sabouret P, Bruckert E. Discontinuation of statin therapy due to muscular side effects: A survey in real life. *Nutr Metab Cardiovas* 2013;23:871-875.
 47. Benkhalti F, Prost J, Paz E, Jimenez F, Modafar C, Boustani E. Effects of feeding virgin olive oil or their polyphenols on lipid of rat liver. *Nutr Res* 2002;22:1067-1075.
 48. Kim B, Ku C, Pham T, Park Y, Martin D, Xie L, *et al.* *Aronia melanocarpa* (chokeberry) polyphenol-rich extract improves antioxidant function and reduces total plasma cholesterol in apolipoprotein E knockout mice. *Nutr Res* 2013;33:406-413.
 49. Qian Q, Liu X, He W, An Y, Chen Q, Wu J, *et al.* TG accumulation inhibitory effects of Jinqi formula by AMPK signaling pathway. *J Ethnopharmacol* 2012;143:41-48.
 50. Guo P, Lian ZQ, Sheng LH, Wu CM, Gao J, Li J, *et al.* The adenosine derivative 2',3',5'-tri-O-acetyl-N6-(3-hydroxyaniline) adenosine activates AMPK and regulates lipid metabolism in vitro and in vivo. *Life Sci* 2012;90:1-7.
 51. Hye Yang M, Vasquez Y, Ali Z, Khan IA, Khan SI. Constituents from *Terminalia* species increase PPARalpha and PPARgamma levels and stimulate glucose uptake without enhancing adipocyte differentiation. *J Ethnopharmacol* 2013;149:490498.
 52. Chao CY, Yin MC, Huang CJ. Wild bitter gourd extract up-regulates mRNA expression of PPARalpha, PPARgamma and their target genes in C57BL/6J mice. *J Ethnopharmacol* 2011;135:156-161.
 53. Mahamuni S, Khose R, Mena F, Badole S. Therapeutic approaches to drug targets in hyperlipidemia. *Bio Med* 2012;2:137-146.
 54. Yoon M. The role of PPARα in lipid metabolism and obesity: Focusing on the effects of estrogen on PPARα actions. *Pharmacol Res* 2009;60:151-159.
 55. Zhang YY, Wang Q, Qi LW, Qin XY, Qin MJ. Characterization and determination of the major constituents in *Belamcandae Rhizoma* by HPLC/DAD-ESI-MS(n). *J Pharm Biomed Anal* 2011;56:304-314.
 56. Wozniak D, Janda B, Kapusta I, Oleszek W, Matkowski A. Antimutagenic and anti-oxidant activities of isoflavonoids from *Belamcanda chinensis* (L.) DC. *Nutr Res* 2010;696:148-153.
 57. Kim Y, Yamada M, Lim S, Lee S, Ryu N, Shin K, Ohuchi K. Inhibition by tectorigenin and tectoridin of prostaglandin E2 production and cyclooxygenase-2 induction in rat peritoneal macrophages. *Biochim Biophys Acta* 1999;1438:399-407.
 58. Jung SH, Lee YS, Lim SS, Lee S, Shin KH, Kim YS. Antioxidant activities of isoflavones from the rhizomes of *Belamcanda chinensis* on carbon tetrachloride-induced hepatic injury in rats. *Arch Pharm Res* 2004;27:184-188.
 59. Wu C, Li Y, Chen Y, Lao X, Sheng L, Dai R, *et al.* Hypoglycemic effect of *Belamcanda chinensis* leaf extract in normal and STZ-induced diabetic rats and its potential active fraction. *Phytomedicine* 2011;18:292-297.
 60. Chen Y, Wu CM, Dai RJ, Li L, Yu YH, Li Y, *et al.* Combination of HPLC chromatogram and hypoglycemic effect identifies isoflavones as the principal active fraction of *Belamcanda chinensis* leaf extract in diabetes treatment. *J Chromatogr B Analyt Technol Biomed Life Sci* 2011;879:371-378.
 61. World Health Organization (WHO). Principles of Laboratory Animal Care. *Chronicle* 1985;39:51-56
 62. Zhang X, Wu C, Wu H, Sheng L, Su Y, Zhang X, *et al.* Antihyperlipidemic effects and potential mechanisms of action of the caffeoylquinic acid-rich *Pandanus tectorius* fruit extract in hamsters fed a high fat-diet. *PLoS One* 2013;8:e61922.
 63. Wu C, Shen J, He P, Chen Y, Li L, Zhang L, *et al.* The alpha-glucosidase inhibiting isoflavones isolated from *Belamcanda chinensis* leaf extract. *Rec Nat Prod* 2012;6:110120.
 64. Sharma B, Balomajumder C, Roy P. Hypoglycemic and hypolipidemic effects of flavonoid rich extract from *Eugenia jambolana* seeds on streptozotocin induced diabetic rats. *Food Chem Toxicol* 2008;46:23762383.
 65. Narender T, Khaliq T, Puri A, Chander R. Antidyslipidemic activity of furano-flavonoids isolated from *Indigofera tinctoria*. *Bioorg Med Chem Lett* 2006;16:3411-3414.
 66. Lee KT, Sohn IC, Kim DH, Choi JW, Kwon SH, Park HJ. Hypoglycemic and hypolipidemic effects of tectorigenin and kaikasaponin III in the streptozotocin-induced diabetic rat and their antioxidant activity in vitro. *Arch Pharm Res* 2000;23:461-466.

67. Xiong Y, Yang Y, Yang J, Chai H, Li Y, Jia Z, *et al.* Tectoridin, an isoflavone glycoside from the flower of *Pueraria lobata*, prevents acute ethanol-induced liver steatosis in mice. *Toxicology* 2010;276:64-72.
68. Guo Q, Wang PR, Milot DP, Ippolito MC, Hernandez M, Burton CA, *et al.* Regulation of lipid metabolism and gene expression by fenofibrate in hamsters. *Biochim Biophys Acta* 2001;1533:220232.