



ISSN (E): 2277- 7695
ISSN (P): 2349-8242
NAAS Rating: 5.03
TPI 2021; 10(3): 249-252
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www.thepharmajournal.com
Received: 25-01-2021
Accepted: 28-02-2021

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Influence of herbal supplementation on dressed yield and percent organ weights of broilers during coccidiosis

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Abstract

Present investigation was conducted to evaluate the influence of herbal supplementation on percent dressed yield and percent organ weights of broilers during coccidiosis. 168, one day old broiler chicks were divided into 7 groups, with 2 replicates of 12 chicks each. Broilers of group I and II were provided unsupplemented diet. Broilers of group III and IV were provided 0.0125% amprolium supplemented diet, whereas broilers of group V and VI were provided diet supplemented with 0.2% madar leaf powder and broilers of group VII were provided 0.4% madar leaf powder supplemented diet. On 15th day of experiment, broilers of group II, IV, VI and VII were infected with 50,000 mixed *Eimeria sp.* sporulated oocysts. Carcass traits as dressed yield % and organ weight % were higher in both infected and uninfected madar leaf powder supplemented groups followed by amprolium and unsupplemented groups, but difference was non-significant for percent dressed yield whereas significant for percent organ weights.

Keywords: Poultry coccidiosis, percent dressed yield, percent organ weight, herbal powder

Introduction

Coccidiosis is commonly called as Red Dysentery. It is caused by the intracellular protozoan parasite *Eimeria*, which undergoes its life cycle in the intestinal mucosa of the infected bird. Nine *Eimeria* species (*Eimeria tenella*, *Eimeria acervulina*, *Eimeria necatrix*, *Eimeria brunetti*, *Eimeria maxima*, *Eimeria mitis*, *Eimeria mivati*, *Eimeria hageni* and *Eimeria praecox*) infect chicken. *Eimeria* parasites colonise intestinal cells and may lead to a massive epithelial destruction. Consequently, the host may suffer from diarrhoea, malabsorption and poor weight gain (Novaes *et al.*, 2012) [16]. *Eimeria tenella*, cause of caecal coccidiosis is the most common and pathogenic species among all *Eimeria sp.* affecting chickens (Ayaz *et al.*, 2003) [3]. Difficulties in tackling avian coccidiosis with chemical coccidiostats and vaccines due to resistance development, residual effects as well as high cost stimulated the scientists to explore further newer methods of control and the natural products are being investigated to this effect (Kayser *et al.*, 2003) [10]. A number of natural feed additives have shown anticoccidial activity. Some plants i.e. *Azadirachta indica*, *Hobrrhena antidysentrica*, *Barberis aristata*, *Embelia ribes*, *Acorus calamus*, *Artemisia annua*, and *Artemisia absinthium* have been shown to possess anti-coccidial activity (Kostadinovic *et al.*, 2019) [11]. Zhang *et al.*, 2020 [26] also evaluated the anticoccidial property of *Camellia sinensis*.

Plant of our study, *Calotropis procera* (Madar) was investigated by many scientists for medicinal properties. Study of Mueen *et al.* (2003) [15] observed the antioxidant properties of madar. Similarly Arya and Kumar (2005) [2] reported its anti-inflammatory activity. Al-Qarawi *et al.* (2001) [1] also illustrated the anthelmintic activity of *Calotropis procera* latex against *Haemonchus contortus* infection in Najdi Sheep. Singh *et al.* (2009) [21] and Zaman *et al.* (2011) [25] studied the anticoccidial effect of *Calotropis sp.* supplementation. Based on various beneficial and therapeutic effects of *Calotropis procera*, current experiment was conducted to study the influence of 0.4% as well as 0.2% madar leaf powder and amprolium on percent dressed yield and percent organ weights of coccidiosis infected broiler chicks.

Materials and Methods

The experiment was conducted on 168, one day old broiler chicks for 30 days. On arrival, chicks were randomly allocated to seven groups. Each group had two replicates with 12 chicks each.

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Chicks of different replicates were kept in separate cages and maintained under similar managerial conditions. The groups were designated as group I, II, III, IV, V, VI and VII. Broilers of group I and II were provided standard control diet without any supplement. In broilers of group III and IV, standard feed was supplemented with 0.0125% amprolium and broilers of group V and VI were provided with basal diet supplemented with 0.2% madar leaf powder. Broilers of group VII were provided standard control diet supplemented with 0.4% madar leaf powder (Table 3.1). On 15th day of experiment broilers of group II, IV, VI and VII were infected with 1 ml suspension containing 50,000 sporulated oocysts of *Eimeria species*. *Eimeria* spp. mixed culture contained *E. tenella* (80%), *E. necatrix* (10%), *E. acervulina* (6%), *E. maxima* (2%) and *E. mitis* (2%), which were identified on the basis of guidelines of Levine (1985) [13]. To isolate the oocysts of *Eimeria sp.*, method described by Holdsworth *et al.* (2004) [8] was used with few modifications.

For percent dressed yield and percent organ weight studies, 3

$$\text{Percent dressed yield (\%)} = \frac{\text{Live weight} - \text{Weight loss as blood, skin, head, shank, wing tips and viscera} \times 100}{\text{Live weight}}$$

$$\text{Percent organ weight (\%)} = \frac{\text{Organ weight} \times 100}{\text{Live weight}}$$

Results and Discussion

Per cent dressed yield and percent organ weights as gizzard, liver, heart or giblet of different groups of broilers on 30th day of experiment or 15th day post infection are presented in Table 1.

On 15th days post infection or 30th day of experiment, among uninfected groups, numerically maximum percent dressed yield were recorded in group V (66.33), whereas minimum in group I (65.22). Group III showed moderate percent dressed yield (65.81), but significant difference was not found between treatments. Durrani *et al.* (2006) [6] reported higher dressing percentage and giblet weight in broilers fed with diet containing turmeric. Among uninfected groups weight gain was significantly maximum in madar leaf powder supplemented groups. This may be due to the growth promoting effect of its saponin content (Kumar *et al.*, 2013; Tiwari *et al.*, 2014; Zaman *et al.*, 2011) [12, 22, 25] and antioxidant effect of its phenols and flavonoides content (Joshi *et al.*, 2009) [9]. Minimum weight gain among uninfected groups was observed in broilers of amprolium supplemented group. Similar findings were observed by Bahadhoran *et al.* (2014) [4].

Among infected groups, maximum per cent dressed yield was showed by broilers of group VII (66.30) followed by group VI (66.27) and group IV (65.71), whereas minimum per cent dressed yield was recorded in group II (65.15), but significant effect of supplementations on dressed yield was not observed. Weight gain among infected groups was maximum in madar leaf powder supplemented group and concentration dependent effect was observed. Coccidiosis neutralizing effect of madar leaf powder may be attributed to its constituent as saponin, which act on the protozoan development by interacting with cholesterol present on the parasitic cell membrane and resulting into parasitic death (Wang *et al.*, 1998) [23]. Singh (2006) [19] noticed significantly higher dressed yield in wheat grass juice supplemented infected group than control infected group. Singh *et al.* (2007) [18] reported that supplementation of amla and turmeric powder in broiler diet improved dressing

broilers from each replicate were slaughtered at 15th days post infection or 30th day of experiment. Before slaughter, each broiler was weighed and weight gain was recorded. It was slaughtered by severing the jugular vein. After complete bleeding, weight was recorded. The weight was again recorded after manual defeathering using hot water at 50-55 °C. Head, shank and wing tips were removed by giving cuts at atlanto-occipital, hock and knee joints respectively and their weights were taken.

After this, a horizontal cut was applied posterior to keel bone. Breast was pushed forward to expose the viscera and the carcass was eviscerated. Weight of the carcass was again recorded and from this dressed yield per cent was calculated. Liver, heart and gizzard were detached from rest of the viscera. Gall bladder was removed from liver. Gizzard was opened and its contents were removed and epithelial linings were detached. Individual weights of all these organs were recorded. Dressing yield were calculated by following formula:

percentage in broilers. Durrani *et al.* (2008) [7] noticed significantly higher dressing percentage in aqueous aloe-vera gel supplemented infected group than infected unsupplemented group. Waskar *et al.* (2009) [24] also revealed that supplementation of herbal toxin binder product in basal diet is efficacious in improving overall meat quality attributes such as carcass yield and dressing percentage (%). Minimum weight gain among infected groups was observed in unsupplemented group, which is due to pathogenic effect of *Eimeria spp.* When the coccidial infection was introduced into the flock, the growth depressing effect of amprolium was neutralized by the effective control of the infection. Bozkurt *et al.*, 2013 [5] also have similar findings.

In both infected and uninfected groups, organ weights as gizzard, heart, liver or giblet percent weights were recorded significantly higher in madar leaf powder supplemented groups. Among infected groups, concentration dependent effect of madar leaf powder was not observed on percent giblet weight. Singh *et al.* (2008) [20] also showed significant effect ($P < 0.05$) of herbal complex and sodabicarb on eviscerated weight, edible weight and giblet weight of treated groups. Milosevic *et al.* (2013) [14] also observed the effect of antioxidant rich plant garlic on carcass traits and found that internal organs as liver and heart were also under the influence of garlic as liver weight was higher in garlic supplemented groups compared to the control group.

Minimum per cent organ weights of gizzard, liver, heart and giblet among infected groups were recorded in infected untreated group i.e. group II.

In our study, gizzard, liver, heart percent weight as well as giblet percent weight was found to be higher in infected groups than respective uninfected groups, this may be due to the hypertrophy of these organs due to effect of coccidiosis, similar results were recorded by Panda and Comb (1964) [17] who noticed hypertrophy of liver, pancreas, adrenal gland, spleen and bursa in chicks which recovered from experimentally induced coccidiosis. Singh *et al.* (2008) [20] also recorded higher % weight of liver, gizzard, pancreas,

spleen and bursa in infected group than uninfected group indicating hyperactivity of these organs.

Conclusion

It was concluded from the study that percent dressed yield among different groups were non-significant as it was ratio of dressed weight and live weight and decrement or increment in both weights occur simultaneously. Madar leaf powder

supplementation affect weight gain in infected and uninfected groups positively, but its significant effect was not observed on dressed yield. Percent giblet weight was significantly higher in uninfected as well as infected madar leaf powder supplemented groups and concentration dependent effect of madar leaf powder on percent giblet weight was not observed during coccidiosis.

Table 1: Carcass yield, organ weighs (% live weight) in broilers of different groups on 15 days post infection or 30th day of experiment

Groups		Weight gain (g)	Dressed yield without giblet (%)	Gizzard (%)	Liver (%)	Heart (%)	Giblet (%)
I	UC	1314.31 ^b	65.22 ^{de}	2.14 ^f	229 ^g	0.603 ^f	5.03 ^f
II	IC	937.76 ^f	65.15 ^e	220 ^e	2.35 ^f	0.612 ^e	5.16 ^e
III	UA	1258.94 ^c	65.81 ^{abcd}	225 ^d	2.40 ^e	0.618 ^d	5.27 ^d
IV	IA	1233.94 ^d	65.71 ^{abcde}	2.30 ^e	2.46 ^d	0.624 ^c	5.39 ^e
V	UM	1416.35 ^a	66.33 ^a	2.45 ^b	2.52 ^c	0.628 ^b	5.60 ^b
VI	IM-1	1172.37 ^e	66.27 ^{abc}	2.50 ^a	2.55 ^b	0.630 ^a	5.68 ^a
VII	IM-2	1228.11 ^d	66.30 ^{ab}	2.49 ^a	2.57 ^a	0.629 ^{ab}	5.69 ^a
CD at 5%		15.154	0.620	0.017	0.017	0.019	0.026

*Significant a, b, c, d means bearing different superscripts in a column differ significantly ($P \leq 0.05$)

Acknowledgement

The authors are thankful to the Dean, College of Veterinary and Animal Sciences, G.B. Pant University of Agriculture and Technology, Pantnagar, Uttarakhand, India for fund and providing necessary facilities during the course of the study.

References

- Al-Qarawi AA, Mahmoud OM, Sobaih MA, Haroun EM, Adam SEI. A Preliminary Study on the Anthelmintic Activity of *Calotropis procera* Latex against *Haemonchus contortus* Infection in Najdi Sheep. *Vet Res Commun* 2001;25(1):61-70.
- Arya S, Kumar VL. Anti-inflammatory Efficacy of Extracts of Latex of *Calotropis procera* Against Different Mediators of Inflammation. *Mediators Inflamm* 2005;4:228-232.
- Ayaz M, Akhtar M, Hayat CS, Hafeez MA, Haq A. Prevalence of coccidiosis in broiler chickens in Faisalabad, Pakistan. *Pakistan Vet J* 2003;23:51-52.
- Bahadoran S, Hassanpour H, Kheirabadi KP, Shekarchian S. Effect of clopidol and amprolium/ethopabate on performance and intestinal morphology of chickens with experimental coccidiosis. *KVFD* 2014;20(4):571-576.
- Bozkurt M, Giannenas I, Kucukyilmaz K, Christaki E, Florou-Paneri P. An update on approaches to controlling coccidia in poultry using botanical extracts. *British Poultry Sci* 2013;54(6):713-727.
- Durrani FR, Ismail M, Sultan A, Suhail SM, Chand N, Durrani Z. Effect of different levels of feed added turmeric (*Curcuma longa*) on the performance of broiler chicks. *J Agri Bio Sci* 2006;1:9-11.
- Durrani FR, Ullah S, Chand N, Durrani Z, Akhtar S. Using aqueous extract of aloe gel as anticoccidial and immunostimulant agent in broiler production. *Sarhad J Agric* 2008;24(4):665-669.
- Holdsworth PA, Conway DP, McKenzie ME, Dayton AD, Chapman HD, Mathis GF, Skinner JT, Mundt HC, Williams RB. World Association for the Advancement of Veterinary Parasitology (WAAVP) guidelines for evaluating the efficacy of anticoccidial drugs in chickens and turkeys. *Vet Parasitol* 2004;121:189-212.
- Joshi R, Sharma A, Lal B. Analysis of antioxidant activity in extracts of *Calotropis procera* (Ait.) R. Br. *J Appl Biosci* 2009;17:899-903.
- Kayser O, Kiderlen AF, Croft SL. National products as antiparasitic drugs. *Parasitol. Res* 2003;90:55-62.
- Kostadinovic L, Popovic S, Pelic DL, Cabarkapa I, Duragic O, Levi J. Medicinal plants as natural alternative to coccidial synthetic drugs in broiler chicken production. *Journal of Agronomy, Technology and Engineering Management* 2019;2(5):325-334.
- Kumar PS, Suresh E, Kalavathy S. Review on a potential herb *Calotropis gigantea* (L.) R. Br., *Sch Acad J Pharm* 2013;2(2):135-143.
- Levine ND. *Veterinary protozoology*. Iowa State University Press, Ethopia 1985, P414.
- Milosevic N, Stojcic MD, Stanacev V, Peric L, Veljic M. The performance and carcass traits of broilers feed with garlic (*Allium sativum*) additive. *European Poultry Sci* 2013;77:254-259.
- Mueen AKK, Rana AC, Dixit VK. Free radical scavenging activity of *Calotropis* species. *Indian Drugs* 2003;40:654-655.
- Novaes J, Thibirio L, Rangel LD, Ferro M, Abe RY, Manha APS *et al*. A comparative transcriptome analysis reveals expression profiles conserved across three *Eimeria* spp. of domestic fowl and associated with multiple developmental stages. *International J Parasitol* 2012;42:39-48.
- Panda B, Combs GF. Effect of coccidiosis on different glands of the growing chicks. *Avian Dis* 1964;8:7-12.
- Singh N, Singh JP, Singh V. Effect of dietary supplementation of herbal formulation on dressing percentage and mortality in broiler chicks. *Indian J Field Vet* 2007;2:22-24.
- Singh VS. Dietary modulation for the management of poultry coccidiosis. Thesis, Ph.D., G.B. Pant University of Agriculture and Technology, Pantnagar 2006.
- Singh VS, Palod J, Shukla SK, Shukla PK. Influence of a herbal anticoccidial on growth, parasitological, haematological parameters and carcass traits of the broilers experimentally infected with mixed field isolates of *Eimeria* species. *Indian J Anim Sci* 2008;78(10):1057-62.
- Singh VS, Yadav CL, Rajput MS. Efficacy of

- combination of aqueous solution of seeds of *Embelia ribes*, *Azadiracta indica* and *Calotropis procera* against poultry coccidiosis. Proceedings of the International Summit on 'Advancing Veterinary Medical Care Challenges and Strategies' & 27th Indian Society for Veterinary Medicine (ISVM) Convention Satellite Seminars on Veterinary Internal Medicine 2009;19-21.
22. Tiwari A, Singh S, Singh S. Chemical analysis of leaf extracts of *Calotropis procera*, International J of Scient. and Res. Publications 2014;4(1):1-2.
 23. Wang Y, Mcallister TA, Newbold CJ, Rode LM, Cheeke PR, Cheng KJ. Effects of *Yucca schidigera* extract on fermentation and degradation of steroidal saponins in the rumen simulation technique (RUSITEC). Animal Feed Sci and Technol 1998;74:143-153.
 24. Waskar VS, Devangare AA, Gosavi PP, Ravikanth K, Maini S, Rekhe DS. Meat quality attributes of broilers supplemented with herbal toxin binder product. Vet World 2009;2(7):274-277.
 25. Zaman MA, Iqbal Z, Abbas RZ, Khan MN. Anticoccidial activity of herbal complex in broiler chickens challenged with *Eimeria tenella*. Parasitol 2011;139(2):237-243.
 26. Zhang K, Li X, Na C, Abbas A, Abbas RZ, Zaman MA. Anticoccidial effects of *Camellia sinensis* (green tea) extract and its effects on blood and serum chemistry of broiler chickens. Pakistan Veterinary Journal 2020;40(1):77-80.