



ISSN (E): 2277- 7695
ISSN (P): 2349-8242
NAAS Rating: 5.23
TPI 2021; SP-10(12): 857-859
© 2021 TPI
www.thepharmajournal.com
Received: 16-10-2021
Accepted: 18-11-2021

QU Nazir

Division of Epidemiology and Preventive Medicine, SKUAST-K, Shuhama, Srinagar, Jammu & Kashmir, India

MI Yattoo

Division of Veterinary Clinical Complex, SKUAST-K, Jammu & Kashmir, India

M Shaheen

Division of Veterinary Clinical Medicine, SKUAST-K, Jammu & Kashmir, India

A Muhee

Division of Epidemiology and Preventive Medicine, SKUAST-K, Shuhama, Srinagar, Jammu & Kashmir, India

OR Parray

Division of Veterinary Clinical Medicine, SKUAST-K, Jammu & Kashmir, India

IU Haq

Division of Epidemiology and Preventive Medicine, SKUAST-K, Shuhama, Srinagar, Jammu & Kashmir, India

Corresponding Author

QU Nazir

Division of Epidemiology and Preventive Medicine, SKUAST-K, Shuhama, Srinagar, Jammu & Kashmir, India

Effect of different antimicrobials on oxidative indices in goats affected with caprine mycoplasmosis

QU Nazir, MI Yattoo, M Shaheen, A Muhee, OR Parray and IU Haq

Abstract

Present study was conducted to study/explore the effect of various therapeutic regimes on the oxidative indices in mycoplasma affected goats. Thirty (N=30) clinically mycoplasma affected goats were taken and allotted to three test groups viz., Group I, II and III and were treated with different antimicrobials with each group having 10 animals (n=10). Group I was treated with tylosin @ 20 mg/kg, Group II with oxytetracycline @10 mg/, and Group III was treated levofloxacin @ 1.5 mg/kg body weight IM. Supportive medicines given with antimicrobials included flunixin meglumine @ 1.1 mg/kg and chlorpheniramine @ 0.5 mg/kg intramuscularly daily for three days in all test groups. Results indicated that at the 0th hour of the experimental trial, there were significant ($P \leq 0.05$) alterations in oxidative indices/values of all the three test groups compared to healthy control. With course of the trial, significant ($P \leq 0.05$) changes were observed in total oxidant status (TOS) and total anti-oxidant status (TAS) in group I alone indicating Tylosin gives better results compared to other antimicrobial agents.

Keywords: antimicrobials, caprine mycoplasmosis, goats, oxidative indices

Introduction

Mycoplasmosis occurs generally as a contagious disease in small ruminants and the infections are caused by various species of Mycoplasma microorganisms (Yattoo *et al.*, 2018) ^[1]. They usually infect respiratory tract, reproductive tract, udder, eyes, joints and a few cause septicemia (Thiaucourt and Bolske, 1996; OIE, 2009) ^[2, 4]. Mycoplasmas are known to cause serious and often fatal illness in goats leading to heavy morbidity (60-100%) and mortality (10-100%) in affected animals, resulting in severe loss to farmers (OIE, 2017; Yattoo *et al.*, 2018) ^[5, 1]. The most commonly used as well as easily available antimicrobials till now are oxyteracycline, tylosin, enrofloxacin, tiamulin, danofloxacin (Sarkar *et al.*, 1992; Kumar *et al.*, 2012; Abraham *et al.*, 2015) ^[3, 7, 10]. Antimicrobial use in combination with anti-inflammatory, anti-pyretic, analgesic and anti-allergic drugs under field conditions is uncommon but is essential for preventing the pathogen induced severe inflammation, fever, thorax pain (pleurodynia) and allergic reactions.

Excessive production of free radicals resulting in oxidative stress or oxidant/antioxidant imbalance occurs in diseased or stressful conditions (Nazifi *et al.*, 2009) ^[11]. Irreversible damage to normal tissues might occur due to either excessive production or inadequate removal of free radicals (Lopaczyski and Zeisel, 2001) ^[12]. Altered levels of various oxidants and antioxidants have been reported in mycoplasma affected goats (Parray *et al.*, 2019) ^[6]. Parray *et al.*, 2019 ^[6] has reported decreased levels of TAS while increased levels of TOS in the mycoplasma affected goats.

Materials and Methods

Blood sampling

At the beginning of study, blood sample from each animal was collected by jugular puncture using sterile 18G needle. For estimation of oxidative indices (TAS, TOS and OSI) the blood was collected in clot activator (5ml) vials for serum extraction.

Procedure

The present study was conducted to explore oxidative changes in mycoplasma affected goats. Clinically affected goats (N=30) were divided in three groups viz. Group I, Group II and Group III with each group having minimum ten animals (n=10). An additional group with 10 healthy animals viz., Group IV was taken as healthy control or negative control. Group I, Group II and Group III were treated with tylosin @ 20 mg/kg, oxytetracycline @ 10 mg/kg and levofloxacin @ 1.5 mg/kg BW intramuscularly, respectively.

The treatment was repeated at the interval of 48 hours four times (0th hour, 48th hour, 96th hour, 144th hour). Supportive medicines given with antimicrobials included flunexin meglumine @ 1.1 mg/kg and chlorpheniramine @ 0.5 mg/kg intramuscularly daily for three days in all test groups. These groups were evaluated for oxidative indices at the beginning of the experimental trial viz., 0th hour and continued at the intervals of 48th hour, 96th hour, 144th hour. Improvement in oxidative indices were then used to aid in assessing the comparative efficacy of different therapeutic interventions. The therapeutic protocol is given in Table 1.

Total oxidant status (TOS) was evaluated as per the method of

Erel (2004) [8] and was estimated through a new automated calorimetric method given by Erel (2004) [8]. The reading was taken at the wavelength of 560nm while the total antioxidant status (TAS) was estimated through a new direct automated method of Erel (2004) [8]. The reading was taken at the wavelength of 600 nm. It was followed by evaluation of oxidative stress index (OSI) which is the ratio of the total oxidant status (TOS) to total antioxidant status (TAS) (Erel, 2005) [9]. Oxidative stress index (OSI) was calculated as per the formula; $OSI = TOS/TAS$ (Erel, 2005) [9]. Likewise, oxidative indices evaluation was also carried out in healthy control group.

Table 1: Therapeutic protocol

Group	Antibiotic (IM)	Supportive Treatment
Group I	Tylosin @ 20 mg/kg IM repeat 48 hourly	Flunexin meglumine @ 1.1 mg/kg and chlorpheniramine @ 0.5 mg/kg IM
Group II	Oxytetracycline @ 10 mg/kg repeat 48 hourly	Flunexin meglumine @ 1.1 mg/kg and chlorpheniramine @ 0.5 mg/kg IM
Group III	Levofloxacin @ 1.5 mg/kg repeat 48 hourly	Flunexin meglumine @ 1.1 mg/kg and chlorpheniramine @ 0.5 mg/kg IM
Group IV	NA	NA

Results and Discussion

Effect of different antibiotic based therapeutic regimens on oxidative indices

a) Total oxidative status (TOS) ($\mu\text{mol H}_2\text{O}_2$ equivalent/L)

At the beginning of the experimental trial, TOS levels were observed significantly ($P \leq 0.05$) higher in Group I, Group II as

well in Group III compared to healthy control/ negative control group. However, post-treatment, significant decrease ($P \leq 0.05$) was observed in Group I with values coming near close to healthy control at 144th hour of the therapeutic trial as shown in Table 2.

Table 2: Effect of different therapeutic regimes on TOS levels ($\mu\text{mol H}_2\text{O}_2$ equivalent/L)

Hours post treatment	Group I	Group II	Group III	Group IV
0	3.1480 \pm 0.149 ^a	3.1660 \pm 0.154 ^a	3.160 \pm 0.161 ^a	1.5640 \pm 0.068 ^{Aa}
48	2.6130 \pm 0.131 ^b	2.8450 \pm 0.144 ^a	2.890 \pm 0.146 ^a	1.6120 \pm 0.07 ^{Aa}
96	2.080 \pm 0.114 ^{Ac}	2.5250 \pm 0.14 ^{ABa}	2.6340 \pm 0.133 ^{BCa}	1.5750 \pm 0.073 ^{Da}
144	1.5590 \pm 0.101 ^{Ad}	2.1990 \pm 0.142 ^{Bb}	2.625 \pm 0.134 ^{BCa}	1.5730 \pm 0.069 ^{ADa}

^{ABCD} values with different superscript differ significantly ($P \leq 0.05$) in a row

^{abcd} values with different superscript differ significantly ($P \leq 0.05$) in a column

b) Effect on Total antioxidant status (TAS) ($\mu\text{mol Trolox}$ equivalent/L)

In all three test groups the total antioxidant status (TAS) was significantly lower ($P \leq 0.05$) as compared to healthy control group (Group IV) at the beginning of the study. With

commencement of treatment significant ($P \leq 0.05$) increase in Group I while non-significant ($P > 0.05$) increase in Group II and Group III was observed at all intervals as shown in Table 3.

Table 3: Effect of different therapeutic regimes on TAS levels ($\mu\text{mol Trolox}$ equivalent/L)

Hours post treatment	Group I	Group II	Group III	Group IV
0	0.6257 \pm 0.014 ^a	0.6228 \pm 0.009 ^a	0.6193 \pm 0.013 ^a	1.4650 \pm 0.083 ^{Aa}
48	0.8235 \pm 0.016 ^b	0.7205 \pm 0.010 ^a	0.7067 \pm 0.012 ^a	1.4610 \pm 0.078 ^{Aa}
96	1.0091 \pm 0.01 ^{Ac}	0.8240 \pm 0.011 ^{Ba}	0.7883 \pm 0.011 ^{BCa}	1.4670 \pm 0.077 ^{Da}
144	1.1999 \pm 0.016 ^{Ad}	0.9229 \pm 0.011 ^{Bb}	0.8740 \pm 0.014 ^{BCa}	1.4680 \pm 0.074 ^{Da}

^{ABCD} values with different superscript differ significantly ($P \leq 0.05$) in a row

^{abcd} values different superscript differ significantly ($P \leq 0.05$) in a column

c) Effect on Oxidative stress index (OSI)

Pre-treatment at 0 hr, Group I, Group II and Group III showed significantly higher ($P \leq 0.05$) status in OSI compared to Group IV (healthy control). Post treatment significant ($P \leq 0.05$) decrease of OSI was observed in Group I and Group

II of all the test groups. However, values for OSI in Group I were comparable to healthy control at 144th hour of the trial and percent decrease in OSI was more (74.2 %) in Group I as shown in Table 4.

Table 4: Effect of different therapeutic regimes on OSI

Hours post treatment	Group I	Group II	Group III	Group IV
0	5.018 \pm 0.163 ^a	5.073 \pm 0.207 ^a	5.136 \pm 0.303 ^a	1.107 \pm 0.087 ^{Aa}
48	3.163 \pm 0.119 ^{Ab}	3.941 \pm 0.172 ^{Bb}	4.110 \pm 0.238 ^{BCb}	1.139 \pm 0.087 ^{Da}
96	2.056 \pm 0.096 ^{Ac}	3.058 \pm 0.149 ^{Bc}	3.349 \pm 0.179 ^{BCbc}	1.109 \pm 0.09 ^{Da}
144	1.295 \pm 0.075 ^{Ad}	2.375 \pm 0.135 ^{Bd}	2.757 \pm 0.158 ^{BCcd}	1.104 \pm 0.08 ^{ADa}
% <(144 th hr)	74.2	53.2	46.3	0

^{ABCD} values with different superscript differ significantly ($P \leq 0.05$) in a row

^{abcd} values with different superscript differ significantly ($P \leq 0.05$) in a column

In different test groups, from the experimental trial, improvement in oxidative indices post-treatment in different test groups was observed which indicate better and fast improvement in disease condition by action of antimicrobials combined with anti-inflammatory and anti-allergic agents (Yatoo *et al.*, 2018)^[1]. Tylosin showed significantly ($P \leq 0.05$) better results compared to other treatment agents used in the experimental trial in terms of improvement in oxidative indices. Effectiveness of antimicrobial drug tylosin has also been reported by Yatoo *et al.* (2018)^[1].

References

1. Yatoo MI, Parray OR, Mir MS, Qureshi S, Kashoo ZA, Nadeem M *et al.* Mycoplasmosis in small ruminants in India: A review. *J Exp. Biol. Agric. Sci* 2018;6(2):264-281.
2. Thiaucourt F, Bolske G. Contagious caprine pleuropneumonia and other pulmonary mycoplasmoses of sheep and goats. *Revue Scientifique Et Technique* 1996;15(4):1397-1414.
3. Sarkar AK, Verma BB, Thakur DK. Treatment of natural cases of pneumonia associated with *Mycoplasma* infection. *Indian Vet. J* 1992;69:1041-1042.
4. OIE. Contagious caprine pleuropneumonia. In *Terrestrial Animal Health Code*. World Organisation for Animal Health, Paris, 2009, 687-690.
5. OIE. OIE - Terrestrial Animal Health Code Twenty-sixth edition, 2017. World Organisation For Animal Health 12, rue de Prony, 75017 Paris, FRANCE, 2017.
6. Parray OR, Yatoo MI, Muheet, Bhat RA, Malik HU, Bashir ST *et al.* Seroepidemiology and risk factor analysis of contagious caprine pleuropneumonia in Himalayan pashmina goats. *Small Rumin. Res* 2019;171:23-36.
7. Kumar A, Verma AK, Gangwar NK, Rahal A. Isolation, characterization and antibiogram of *Mycoplasma bovis* in sheep pneumonia. *Asian J Anim. Vet. Adv* 2012;7:149-157.
8. Erel O. A novel automated direct measurement method for measuring total oxidant status. *Clinical Biochemistry* 2004;38:1103-1111.
9. Erel O. A new automated calorimetric method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clinical Biochemistry* 2005;37:277-285.
10. Abraham SS, Asha TT, Julie B, Prathiush PR, Nandakumar S, Prasad PM. Pathological and molecular characterization of contagious caprine pleuropneumonia (CCPP) outbreak in Kerala. *Indian J Vet. Pathol* 2015;39(2):121-124.
11. Nazifi S, Saeb M, Ghafari N, Razeghian I, Razavi SM, Vosoughi F *et al.* Reference values of oxidative stress parameters in adult native Iranian goats. *Bulg. J. Vet. Med* 2009;12(2):119-124.
12. Lopaczyski W, Zeisel SH. Antioxidants, programmed cell death, and cancer. *Nutr. Res* 2001;21:295-307.