



ISSN (E): 2277- 7695

ISSN (P): 2349-8242

NAAS Rating: 5.03

TPI 2020; 9(5): 270-273

© 2020 TPI

[www.thepharmajournal.com](http://www.thepharmajournal.com)

Received: 24-03-2020

Accepted: 26-04-2020

**Syed Kashif Mohiuddin**

Department of Biotechnology,  
Joginpally B.R Pharmacy  
College, Moinabad, Hyderabad  
Telangana, India

**Dr. Pittu Vishnu Priya**

Department of Biotechnology,  
Joginpally B.R Pharmacy  
College, Moinabad, Hyderabad  
Telangana, India

**AVSSS Gupta**

Department of Biotechnology,  
Joginpally B.R Pharmacy  
College, Moinabad, Hyderabad  
Telangana, India

## Treatment of Duchenne muscular dystrophy (DMD) by gene therapy using adeno-associated viral vectors

Syed Kashif Mohiuddin, Dr. Pittu Vishnu Priya and AVSSS Gupta

### Abstract

Duchenne muscular dystrophy (DMD) is a severe type of muscular dystrophy. The symptom worsens quickly. The muscle loss occurs first in the thighs and pelvis followed by the arms. This can result in trouble standing up. Most of the patients are unable to walk by the age of 12. Affected muscles may look larger due to increased fat content. The disorder is the X-linked recessive. Where about two thirds of cases are inherited from a person's mother, and one third of cases are due to a new mutation by drugs. It is caused by a mutation in the gene for the protein dystrophin. Dystrophin is important to maintain the muscle fibre's cell membrane. Genetic testing can often make the diagnosis at birth. Medications used include steroids to slow muscle degeneration, anticonvulsants to control seizures and some muscle activity, and immunosuppressant's to delay damage to dying muscle cells. DMD affects about one in 5,000 males at birth It is the most common type of muscular dystrophy. The average life expectancy is 26 years Gene therapy can be used as treatment in early stages, DNA delivery anti-sense oligonucleotides and plasmid DNA, gene therapies and stem cell technologies. all show promise for being able to impact different types of muscular dystrophies. Focusing on developing direct gene replacement strategies to treat recessively inherited form of muscular dystrophy, particularly Duchenne muscular dystrophy (DMD) using adeno-associated viral vectors to deliver synthetic dystrophin genes for the purpose of developing gene therapy for DMD.

**Keywords:** Duchenne muscular dystrophy (DMD), protein dystrophin, genetic testing, gene therapy, anti-sense oligonucleotides, plasmid DNA, adeno-associated viral vectors

### Introduction

Duchenne muscular dystrophy (DMD) is a severe progressive muscle degenerative disease caused by dystrophin mutations in childhood, occurring in about one of every 5000 male births. DMD is caused by the absence of dystrophin or due to mutation of *DMD* located on the X chromosome and thus primarily affects males. (DMD) is a rare neuromuscular disorder that causes progressive weakness and early death. Gene therapy is an area of new therapeutic development by using adeno associated viral vectors (AAV) vectors are the leading platform for gene delivery for the treatment of a variety of human diseases. Capsids such as AAV8 and AAV9 can target multiple muscle types throughout the body, enabling rAAV gene therapies to be developed for multiple muscle diseases, especially those afflicting muscles of the entire body, such as Duchenne muscular dystrophy (DMD). After the transfer of dystrophin protein muscle can serve as a bio-factory to produce secreted therapeutic dystrophin proteins for the treatment of muscle diseases

**Symptoms:** A progressive neuromuscular disorder, is muscle weakness associated with muscle wasting with the voluntary muscles being first affected, especially those of the hips, pelvis area, thighs, shoulders, and calves. Muscle weakness also occurs later, in the arms, neck, and other areas. Calves are often enlarged. Symptoms usually appear before age six and may appear in early infancy

### Other physiological changes which occur are

1. Curved spine due to weak back muscles
2. Swollen calves due to fat and scar tissue build up
3. Arms held back for balance
4. Belly stick due to weak abdomen

**Corresponding Author:**

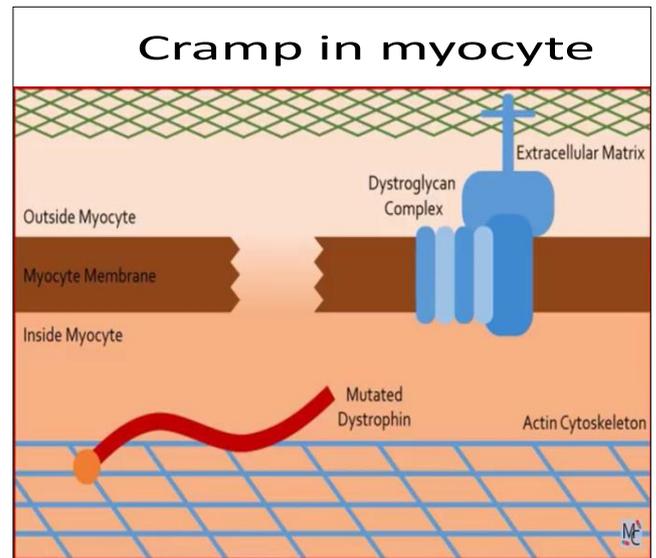
**Syed Kashif Mohiuddin**

Department of Biotechnology,  
Joginpally B.R Pharmacy  
College, Moinabad, Hyderabad  
Telangana, India

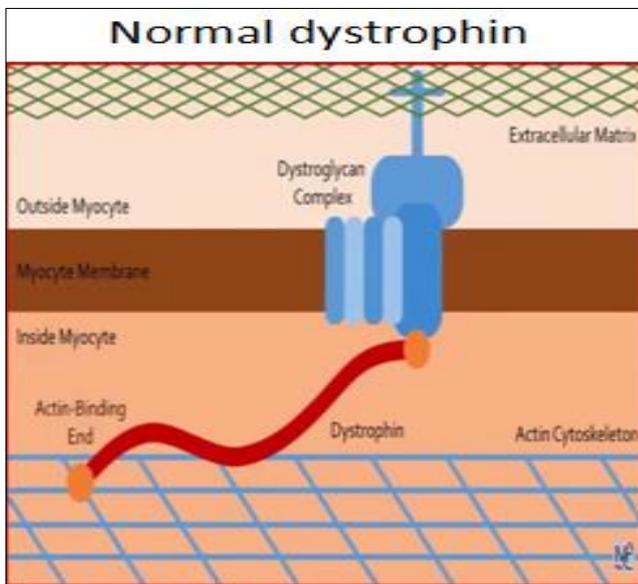
**Functions of dystrophin gene are**

In skeletal muscle, dystrophin protein is located just beneath the sarcolemma connecting the cytoskeleton to the extracellular matrix via a dystrophin associated glycoprotein complex (DGC) and thus stabilizes the sarcolemma of the muscle fibre and protects it from damage during the repeated cycles of contraction and relaxation. Mutations in the dystrophin gene, resulting in decreased or lack of dystrophin expression, such as in DMD patients, results in destabilization of the DGC complex and muscle fibre degeneration.

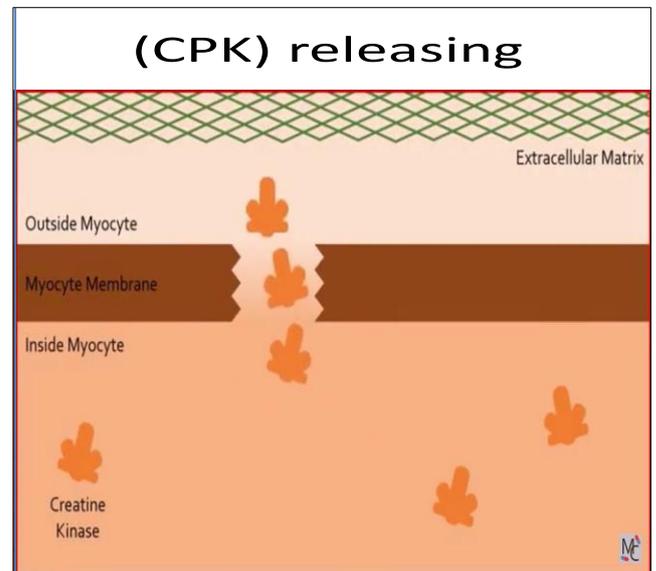
Dystrophin is a support between actin binding end and myocyte membrane (fig1) the patient with DMD has a dystrophin with shorter central rod (fig 2) so that it cannot provide support between actin binding end and myocyte membrane due to which cramps are formed in myocyte membrane (fig 3), as the cramps are formed. The creatine kinase used for muscle strength is released out into the blood circulation (fig 4) as the result the muscles get weak and scar, fat tissue is formed in the damaged myocyte resulting in swollen and weak muscles



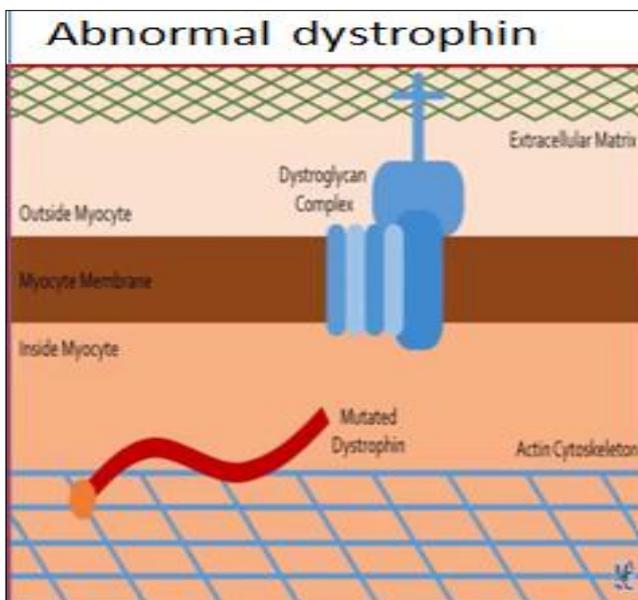
**Fig 3:** shows the cramp formed in myocyte due to abnormal function of mutated dystrophin.



**Fig 1:** shows normal dystrophin function



**Fig 4:** Shows the release of creatine kinase from the cramp in myocyte



**Fig 2:** shows the abnormal function of mutated dystrophin

**AAV over adenovirus**

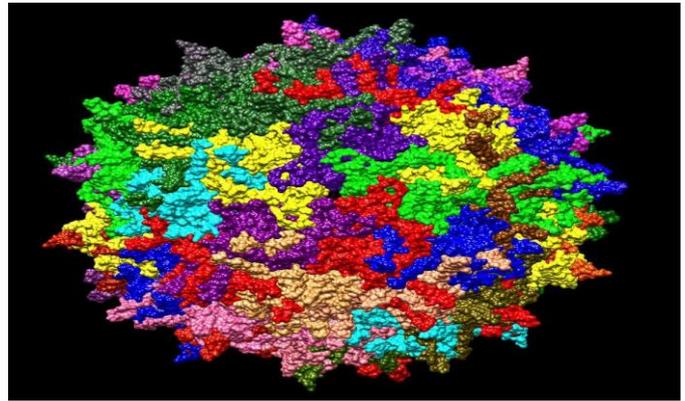
The adeno-associated viruses are small and relatively simple viruses that have greater potential as vectors for gene therapy. It is a single-stranded, non-enveloped DNA virus of 4.5. kb in size.

The viruses' apparent lack of pathogenicity.

It has the ability to stably integrate into the host cell genome at a specific site.

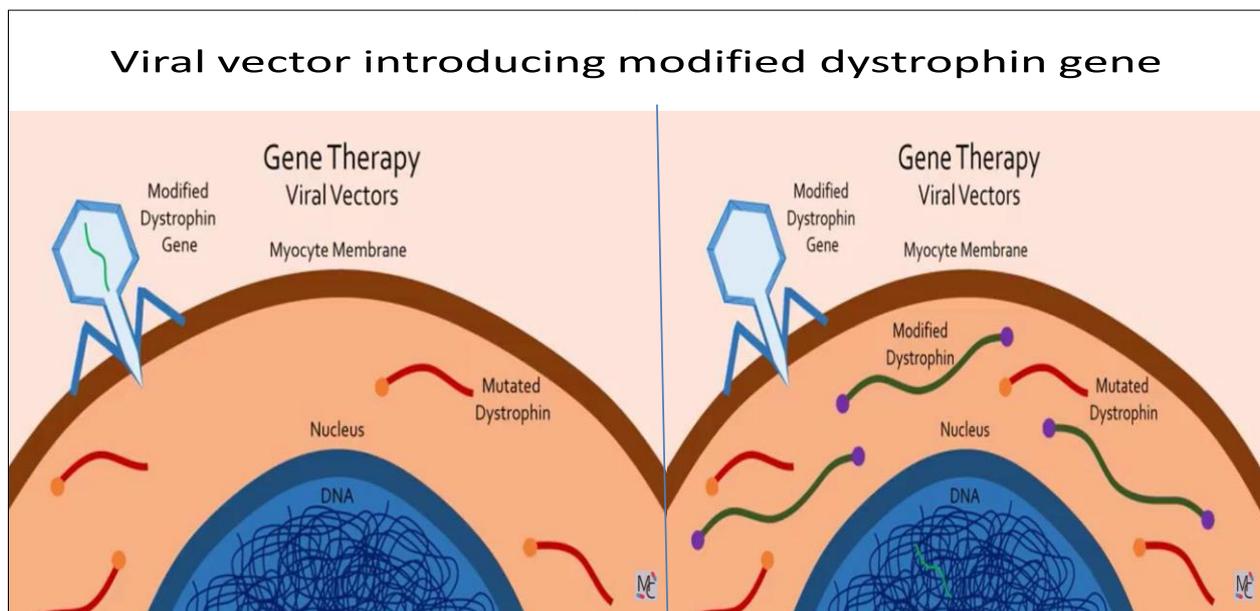
**(AAV) Adeno-Associated Viruses** AAV belongs to the genus Depend parvovirus within the family Parvoviridae. the life cycle is depends on the presence of a helper virus, such as AdV. AAV is found in multiple vertebrate species, including human and non-human primates. AAV does not cause any human diseases. The genome is flanked by two T-shaped inverted terminal repeats at the ends that largely serve as the viral origins of replication. The rep gene encodes four proteins required for viral replication; they are named after their molecular masses: Rep78, Rep68, Rep52 and Rep40. Capsids AAV8 and AAV9 targets the muscle types

throughout the body, rAAV gene therapy is developed for multiple muscle diseases, especially those afflicting muscles of the entire body, such as Duchenne muscular dystrophy (DMD). The treatment method is by using viral vectors some viruses are known to incorporate their own DNA into human cells, we have taken advantage of the skills making viruses carrying modified Dystrophin Gene instead of viral genes (fig 6) now the incorporated dystrophin gene which get incorporated into DNA and Myocytes and allowing them to produce modified Dystrophin protein (fig 7) in the individuals body and the modified dystrophin gene will function in the place of mutated dystrophin gene thus provide the support to the myocyte during the contraction and relaxation so there will be no further damage to the myocyte and in growing children the myocyte will be repaired hence there will be no release of CPK into the blood hence there is enough CPK for contractions and relaxation of muscle



**Fig 5:** Adeno-associated virus structure

**Example:** Adeno-Associated Virus-5, Avian Adeno Associated Virus, Adeno-Associated Virus-3



**Fig 6:** shows mutated dystrophin gene

**Fig 7:** shows the modified dystrophin gene introduced in body by AAV

**Conclusion:** Treatment of Duchenne muscular dystrophy (DMD) by gene therapy using adeno-associated viral vectors. The method was found to be highly reproducible. Our aim is to apply this technique to treat DMD patients from this promising therapy.

#### References

1. Kristy J Brown, Ramya Marathi, Alyson A Fiorillo, Eugene F Ciccimaro, Seema Sharma, David S Rowlands *et al.* Accurate Quantitation of Dystrophin Protein in Human Skeletal Muscle Using Mass Spectrometry.
2. Poysky J. Behavior patterns in Duchenne muscular dystrophy: report on the Parent Project Muscular Dystrophy behavior workshop 8-9 of December 2006, Philadelphia, USA. *Neuromuscul Disord.* 2007; 17:986-994. [PubMed] [Google Scholar]
3. Bushby K, Finkel R, Birnkrant DJ *et al.* Diagnosis and management of Duchenne muscular dystrophy, part 1: diagnosis, and pharmacological and psychosocial management. *Lancet Neurol.* 2010; 9:77-93. [PubMed] [Google Scholar]
4. O'Brien KF, Kunkel LM. Dystrophin and muscular dystrophy: past, present, and future. *Mol Genet Metab.* 2001; 74:75-88. [PubMed] [Google Scholar]
5. Backman E, Henriksson KG. Low-dose prednisolone treatment in Duchenne and Becker muscular dystrophy. *Neuromuscul. Disord.* 1995; 5:233-241. [PubMed] [Google Scholar]
6. Bartlett RJ, Stockinger S, Denis MM *et al.* *In vivo* targeted repair of a point mutation in the canine dystrophin gene by a chimeric RNA/DNA oligonucleotide. *Nat. Biotechnol.* 2000; 18:615-622. [PubMed] [Google Scholar]
7. Baumeister R, Ge L. The worm in us- *Caenorhabditis elegans* as a model of human disease. *Trends Biotechnol.* 2002; 20:147-148. Review. [PubMed] [Google Scholar]
8. Bushby K, Finkel R, Birnkrant DJ, Case LE, Clemens PR, Cripe L *et al.* Diagnosis and management of Duchenne muscular dystrophy, part 1: diagnosis, and pharmacological and psychosocial management. *Lancet*

- Neurol. 2010; 9(1):77-93.  
10.1016/S1474-4422(09)70271-6  
[PubMed] [CrossRef] [Google Scholar]
9. Flanigan KM. Duchenne and Becker muscular dystrophies. *Neurol Clin.* 2014; 32(3):671-88. viii. 10.1016/j.ncl.2014.05.002  
[PubMed] [CrossRef] [Google Scholar]
  10. Ryder S, Leadley RM, Armstrong N, Westwood M, de Kock S, Butt T *et al.* The burden, epidemiology, costs and treatment for Duchenne muscular dystrophy: an evidence review. *Orphanet J Rare Dis.* 2017; 12(1):79. 10.1186/s13023-017-0631-3 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
  11. Otto C, Steffensen BF, Højberg AL, Barkmann C, Rahbek J, Ravens-Sieberer U *et al.* Predictors of Health-Related Quality of Life in boys with Duchenne muscular dystrophy from six European countries. *J Neurol.* 2017; 264(4):709-23. 10.1007/s00415-017-8406-2  
[PubMed] [CrossRef] [Google Scholar]
  12. Uzark K, King E, Cripe L, Spicer R, Sage J, Kinnett K *et al.* Health-related quality of life in children and adolescents with Duchenne muscular dystrophy. *Pediatrics.* 2012; 130(6):e1559-66. 10.1542/peds.2012-0858 [PubMed] [CrossRef] [Google Scholar]
  13. Landfeldt E, Lindgren P, Bell CF, Guglieri M, Straub V, Lochmüller H *et al.* Quantifying the burden of caregiving in Duchenne muscular dystrophy. *J Neurol.* 2016; 263(5):906-15. 10.1007/s00415-016-8080-9 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
  14. Hatzmann J, Heymans HS, Ferrer-i-Carbonell A, van Praag BM, Grootenhuis MA. Hidden consequences of success in pediatrics: parental health-related quality of life-results from the Care Project. *Pediatrics.* 2008; 122(5):e1030-8. 10.1542/peds.2008-0582  
[PubMed] [CrossRef] [Google Scholar]
  15. Abi Daoud MS, Dooley JM, Gordon KE. Depression in parents of children with Duchenne muscular dystrophy. *Pediatr Neurol.* 2004; 31(1):16-9. 10.1016/j.pediatrneurol.2004.01.011  
[PubMed] [CrossRef] [Google Scholar]
  16. Kenneson A, Bobo JK. The effect of caregiving on women in families with Duchenne/Becker muscular dystrophy. *Health Soc Care Community.* 2010; 18(5):520-8. [PubMed] [Google Scholar]
  17. Magliano L, Patalano M, Sagliocchi A, Scutifero M, Zaccaro A, D'angelo MG *et al.* Burden, professional support, and social network in families of children and young adults with muscular dystrophies. *Muscle Nerve.* 2015; 52(1):13-21. [PMC free article] [PubMed] [Google Scholar]
  18. Adeno-associated virus vector as a platform for gene therapy delivery Dan Wang, Phillip W. L. Tai, and Guangping Gao.
  19. Atchison RW, Casto BC, Hammon WM. Adenovirus-associated defective virus particles. *Science* [PubMed] [Google Scholar] This report is among the first to identify by electron microscopy the presence of AAV as a defective virus in simian Adv preparations. 1965; 149:754-756.
  20. Hoggan MD, Blacklow NR, Rowe WP. Studies of small DNA viruses found in various adenovirus preparations: physical, biological, and immunological characteristics. *Proc. Natl Acad. Sci. USA.* 1966; 55:1467-1474. [PMC free article] [PubMed] [Google Scholar]
  21. Blacklow NR, Hoggan MD, Rowe WP. Isolation of adenovirus-associated viruses from man. *Proc. Natl Acad. Sci. USA.* 1967; 58:1410-1415. [PMC free article] [PubMed] [Google Scholar]
  22. Carter BJ. Adeno-associated virus and the development of adeno-associated virus vectors: a historical perspective. *Mol. Ther.* 2004; 10:981-989. [PubMed] [Google Scholar]
  23. Berns KI. My life with adeno-associated virus: a long time spent studying a short genome. *DNA Cell Biol.* 2013; 32:342-347. [PMC free article] [PubMed] [Google Scholar]
  24. ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). 2000 Feb 29 -. Identifier NCT01976091, Gene Transfer Clinical Trial for LGMD2D (Alpha-sarcoglycan Deficiency) Using scAAVrh74.tMCK.hSGCA; 2013 Nov 5 [cited 2017 Aug 30]; [about 5 screens].  
<https://clinicaltrials.gov/ct2/show/NCT01976091>.
  25. Al-Zaidy SA, Sahenk Z, Rodino-Klapac LR, Kaspar B, Mendell JR. Follistatin Gene Therapy Improves Ambulation in Becker Muscular Dystrophy. *J Neuromuscul Dis.* 2015; 2(3):185-92. 10.3233/JND-150083 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
  26. Vulin A, Barthélémy I, Goyenville A, Thibaud JL, Beley C, Griffith G *et al.* Muscle function recovery in golden retriever muscular dystrophy after AAV1-U7 exon skipping. *Mol Ther.* 2012; 20(11):2120-33. 10.1038/mt.2012.181 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
  27. Kornegay JN, Peterson JM, Bogan DJ, Kline W, Bogan JR, Dow JL *et al.* NBD delivery improves the disease phenotype of the golden retriever model of Duchenne muscular dystrophy. *Skelet Muscle.* 2014; 4:18 10.1186/2044-5040-4-18 [PMC free article] [PubMed] [CrossRef] [Google Scholar]
  28. Hollin IL, Peay HL, Bridges JF. Caregiver preferences for emerging duchenne muscular dystrophy treatments: a comparison of best-worst scaling and conjoint analysis. *Patient.* 2015; 8(1):19-27. 10.1007/s40271-014-0104-x [PubMed] [CrossRef] [Google Scholar]
  29. Hollin IL, Peay HL, Apkon SD, Bridges JFP. Patient-centered benefit-risk assessment in duchenne muscular dystrophy. *Muscle Nerve.* 2017; 55(5):626-34. [PubMed] [Google Scholar]
  30. Peay HL, Hollin I, Fischer R, Bridges JF. A community-engaged approach to quantifying caregiver preferences for the benefits and risks of emerging therapies for Duchenne muscular dystrophy. *Clin Ther.* 2014; 36(5):624-37. 10.1016/j.clinthera.2014.04.011 [PubMed] [Google Scholar]