



ISSN (E): 2277- 7695

ISSN (P): 2349-8242

NAAS Rating: 5.03

TPI 2020; 9(1): 318-324

© 2020 TPI

www.thepharmajournal.com

Received: 22-11-2019

Accepted: 26-12-2019

Vasantha SKI

Assistant Professor, Department of Veterinary Physiology, NTR college of Veterinary Science, Gannavaram, Andhra Pradesh, India

Nikhil Kumar Tej

Assistant Professor, Department of Veterinary Physiology, NTR College of Veterinary Science, Gannavaram, Andhra Pradesh, India

Saikiran BVS

MVSc, Department of Veterinary Physiology, NTR College of Veterinary Science, Gannavaram, Andhra Pradesh, India

Lavanya S

MVSc, Department of Veterinary Physiology, NTR College of Veterinary Science, Gannavaram, Andhra Pradesh, India

Sivaiah K

Scientist, Livestock Research Station, Lam farm, Guntur, Andhra Pradesh, India

Mutha Rao M

Professor and Officer in-Charge, Livestock Research Station, Lam farm, Guntur, Andhra Pradesh, India

Srinivasa Prasad CH

Professor and University Head, Department of Veterinary Physiology, NTR College of Veterinary Science, Gannavaram, Andhra Pradesh, India

Corresponding Author:

Vasantha SKI

Assistant Professor, Department of Veterinary Physiology, NTR college of Veterinary Science, Gannavaram, Andhra Pradesh, India

Hematological and biochemical changes in Ongole cows one week before and one week after parturition in relation to THI

Vasantha SKI, Nikhil Kumar Tej, Saikiran BVS, Lavanya S, Sivaiah K, Mutha Rao M and Srinivasa Prasad CH

Abstract

The objective of the present study was to assess the hematological and biochemical changes during periparturient period Ongole cattle. Eight Ongole pregnant cattle of four to six years of age during the summer season were used for the experiment. Temperature humidity index (THI) was calculated by recording the ambient temperature and relative humidity in the calving season. Whole blood was collected one week before and after calving. Haematological and biochemical parameters were estimated. The study revealed that transition period had no effect ($p>0.05$) on total erythrocytic count (TEC), haemoglobin (Hb), packed cell volume (PCV), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC). The total leucocyte count (TLC) was significantly ($p<0.05$) higher and lymphocyte percent was significantly ($p<0.05$) lower during postpartum compared to prepartum. There was no significant ($p>0.05$) difference between prepartum and postpartum with respect to biochemical parameters such as total protein, BUN and creatinine. Cholesterol, calcium and phosphorous levels were significantly ($p<0.05$) lower during postpartum compared to prepartum. Changes in hematological and biochemical parameters are indicative of parturition stress in Ongole cows and these parameters could be used as stress markers. The mean values obtained for hematological and biochemical parameters are within the reference range of cattle despite of high THI which is indicative of high thermo tolerance to the existing adverse ambience.

Keywords: Ongole cattle, hematological, biochemical, periparturient period, temperature humidity index

1. Introduction

Periparturient period is the most stressful condition for dairy animals as it induces several physiological changes for the onset of lactation and is also associated with oxidative stress (Konvicna *et al.*, 2015; Ambily *et al.*, 2019) [15, 4]. The animals are also subjected to various metabolic and endocrine changes during this phase in the life cycle of dairy animals (Tharwat *et al.*, 2013). Initiation of lactation compared to the late gestation induces more stress to the animals as the nutrient requirement by the mammary gland is several times more than that of a growing fetus (Tharwat *et al.*, 2012) [34]. This makes the animals highly susceptible to negative energy balance (Mohamed *et al.*, 2015) [23], fat mobilization, elevation of circulating nonesterified fatty acids and ketone bodies (Ingvarsten and Andersen, 2000; Seifi *et al.*, 2007) [16, 31]. Moreover, when transition period is accompanied by high environmental temperature and humidity, the health of the animals is more compromised making them prone for diseases. The alterations in hematological and biochemical parameters are used to understand the physiological adjustments during periparturient period. Moreover in tropical country like India, thermal stress is one of the major factors which further aggravate stress in cattle and buffaloes. Hence a comprehensive understanding of the effect of the THI on the periparturient period is of utmost importance to understand the physiology of animals and to adopt nutritional and managemental strategies to optimize production.

2. Materials and Methods

Eight female Ongole cattle of four to six years of age were randomly selected from the LAM Farm, Guntur, and Andhra Pradesh. The animals were kept in shed with appropriate facilities for feeding and watering. The animals were fed as per ICAR feeding standards (Ranjhan, 1998) [27] with concentrates and roughage stall fed.

2.1 Meteorological data

The experiment was conducted at different days in the month of May (summer). The meteorological data during May was recorded. Ambient temperature (AT) and relative humidity (RH) was recorded from outdoor atmosphere using Digital Thermo Hygrometer. Temperature humidity index (THI) was calculated using the equation, $THI = (0.8 \times T_{db}) + [(RH/100) \times T_{db} - 14.4] + 46.4$ (T_{db} = temperature of dry bulb, RH = relative humidity) (Mader *et al.*, 2006) [22] from daily recordings AT and RH.

2.2 Blood collection and analysis

Blood samples were collected on one week prepartum and one week postpartum for assessing the haematological parameters and biochemical parameters. Whole blood was collected between 10.00 h-11.00 h by venepuncture of the jugular vein using vacutainers containing sodium ethylene diamine tetra acetic acid salt (EDTA) as anticoagulant. Total erythrocyte count (TEC), total leucocyte count (TLC), Haemoglobin (Hb), Lymphocyte%, Monocyte%, Granulocyte%, packed cell volume (PCV), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) were estimated using Mindray Heamoanalyzer BC 2800 Auto hematology Analyzer purchased from Shenzhen Mindray Biomedical Electronics Co Ltd. Haematological indices such as MCV, MCH and MCHC were calculated from the erythrocytic series.

The serum was separated from another aliquot of blood samples by centrifugation at 3500 rpm for 10 minutes at room temperature and stored at -20 °C for estimation of biochemical parameters. total plasma proteins (biuret method, Doumas *et al.*, 1971) [9], cholesterol (phenol amino antipyrine method, Richmond, 1973) [28], total calcium (o-cresol Phenolphthalein complex one method, Baginski, 1973) [5], phosphorous (Ammonium molybdate method, Wang, 1983) [35], urea (GLDH-urease method, Young, 1990) and ceatinine (Jaffe's method, Bowers, 1980) [7] concentration was determined photometrically.

2.3 Statistical analysis

The data obtained on various parameters were statistically analysed using paired t test (Snedecor and Cochran, 1994). The whole data was analysed using computerized software programme SPSS Ver.20.0.

3. Results and Discussion

The mean \pm SE values of minimum and maximum temperature, relative humidity and THI during the study period are given in Table 1. The mean THI recorded in the present study was found to be 81.22 ± 1.69 . As per Helal *et al.* (2010) [15] a THI of 74 or less is considered as normal or comort zone, A THI of 75-78 is indicative that animal is in alert state, while a THI of 79-83 is danger state and above 84 is an emergency condition. In the current study, a high THI of above 72 in the present study was indicative that the animals were under significant heat stress during the study period.

3.1 Hematological parameters

The changes in hematological parameters from prepartum to postpartum were presented in Fig. 1 to 10. The results of our present study with respect to hematological parameters are as follows. The TEC concentration during prepartum and postpartum was found to be 6.2 ± 0.21 and 6.42 ± 0.24 $10^6/\mu\text{l}$

respectively; with no significant ($p > 0.05$) difference. Similarly, Abdulkareem (2013) [3] also did not notice significant difference in TEC, Hb, PCV, MCV, MCH, and MCHC between calving and postpartum period in Iraqi riverine buffaloes. The findings of Abdelrazek *et al.* (2018) [2] are also similar to our findings, with non-significant alteration in TEC, Hb, PCV, MCV, MCH, MCHC of Egyptian buffaloes in transition period. In contrary to present findings, Gavan *et al.* (2010) [12] reported TEC values of 6.94 and 5.02 $10^6/\mu\text{l}$ suggesting a decreased TEC at parturition in HF cows. Nevertheless TEC is within the reference range of 4.9-7.5 $10^6/\mu\text{l}$ for cattle (Wood and Rocha, 2010) [36].

The TLC concentration during prepartum and postpartum was found to be 8.98 ± 0.51 and 11.48 ± 0.79 $10^3/\mu\text{l}$ respectively; with significant ($p < 0.05$) increase in postpartum compared to prepartum. The results of Abdelrazek *et al.* (2018) [2] also showed an increase in TLC towards calving and few days postpartum. Contradictory to our findings Gavan *et al.* (2010) [12] reported values of 13.8 and 11.68 $10^3/\mu\text{l}$ suggesting a decreased TLC at parturition in HF cows. In the reports of Mohamed *et al.* (2015) [23] a significant decrease in TLC in the early lactating period when compared with the late pregnant Murrah buffalo heifers in Assiut city, Egypt was observed. At parturition, there was an increase in TLC due to high levels of cortisol secreted, which in turn stimulates the bone marrow (Kim *et al.* (2005) [19]. Nevertheless TLC is within the reference range of 5.1-13.3 $10^3/\mu\text{l}$ for cattle (Wood and Rocha, 2010) [36].

The Hb concentration during prepartum and postpartum was found to be 8.68 ± 0.25 and 8.85 ± 0.25 g/dl respectively with no significant ($p > 0.05$) difference. Contrarily, Gavan *et al.* (2010) [12] reported an Hb values of 11.10 and 7.88 g/dl during prepartum and postpartum with significant difference in HF cows. Nevertheless Hb is within the reference range of 8.4-12.0 g/dl for cattle (Wood and Rocha, 2010) [36].

The PCV observed during prepartum and postpartum was found to be 28.20 ± 1.17 and $29.03 \pm 1.11\%$ respectively with no significant ($p > 0.05$) difference. Gavan *et al.* (2010) [12] reported a PCV value of 30.5 and 21.31% during prepartum and postpartum respectively suggesting a decreased PCV in HF cows. The MCV observed during prepartum and postpartum was 45.23 ± 0.69 and 45.68 ± 0.49 fl respectively. Gavan *et al.* (2010) [12] reported MCV values of 43.8 and 41.66 fl during prepartum and postpartum in HF cows with no significant difference. In contrary to the present findings Mohamed *et al.* (2015) [23] reported a significant decrease in PCV, MCV in the early lactating period. Nevertheless PCV and MCV are within the reference range of 21-30% and 36-50 fl for cattle (Wood and Rocha, 2010) [36].

The MCH observed during prepartum and postpartum was found to be 13.73 ± 0.20 and 13.78 ± 0.19 pg respectively with no significant ($p > 0.05$) difference. Gavan *et al.*, (2010) [12] reported no significant difference between prepartum and postpartum with MCH values of 16.02 and 15.46 pg in HF cows. The MCHC observed during prepartum and postpartum was found to be 33.68 ± 2.01 and $31.80 \pm 1.80\%$ respectively. Gavan *et al.* (2010) [12] also reported no significant difference between prepartum and postpartum with MCHC values of 36.42 and 37.16% in HF cows. Contradicting our results, Mohamed *et al.* (2015) [23] reported a significant decrease in MCHC in the early lactating period. Nevertheless MCH and MCHC are within the reference range of 14-19 pg and 38-43 g/dl for cattle (Wood and Rocha, 2010) [36].

The monocyte%, Lymphocyte% and granulocyte% prepartum

and postpartum was found to be 9.38 ± 0.49 and 10.03 ± 0.26 , 42.5 ± 0.29 and 36.48 ± 4.25 and 50.88 ± 2.20 , 53.43 ± 4.35 respectively. There was no significant ($p > 0.05$) between pre and postpartum with respect to monocyte and granulocyte % whereas, lymphocyte% was significantly ($p < 0.05$) lower during postpartum compared to prepartum. Similar to present findings, Gavan *et al.* (2010) [12] reported no significant difference in monocyte% between pre and postpartum in HF cows. Abdelrazek *et al.* (2018) [2] observed significant decrease in Lymphocyte% and increase in Monocyte% at calving in Egyptian buffaloes in transition period. High lymphocyte% during postpartum could be due to increased cortisol levels at calving which causes Lymphocytopenia (Jacor *et al.*, 2001) [17]. Nevertheless monocyte and Lymphocyte counts are within the reference range of $1.8-8.1 \times 10^3/\mu\text{l}$ and $0.1-0.7 \times 10^3/\mu\text{l}$ for cattle (Wood and Rocha, 2010) [36].

3.2 Biochemical parameters

The changes in biochemical parameters from prepartum to postpartum were presented in Fig. 11 to 16. The changes in some of the parameters indicate the physiological changes across the parturition but do not necessarily indicate any disease (Tharwat *et al.*, 2015) [33]. The mean serum total protein concentration recorded in the present study was found to be 5.94 ± 0.13 and 5.63 ± 0.14 g/dl in prepartum and postpartum respectively with no significant difference. The present findings were in agreement with the reports of Piccione *et al.* (2012) [25] where no significant in protein concentration between pre and postpartum dairy cows with values of 5.52 ± 0.45 and 4.83 ± 0.88 g/dl. Whereas, Contradictory to the present findings, Tharwat *et al.* (2015) [33] found an increased trend in total protein in goats during transition period while Seifi *et al.* (2007) [31] also reported a significant decrease in protein concentration with values of 71.8 ± 1.1 and 61.6 ± 1.6 g/l in prepartum and postpartum dairy cows. Nevertheless the total protein value recorded from our study falls within the reference range of $6.2-8.2$ g/dl for cattle (Kahn, 2005) [21]. Serum plasma proteins indicate the maternal requirement of proteins for milk production and immunoglobulins (Mohri *et al.*, 2007; Piccione *et al.*, 2012) [24, 25].

The mean serum cholesterol concentration recorded in the present study was found to be 84.80 ± 1.59 and 79.15 ± 0.85 mg/dl. The serum cholesterol concentration was significantly ($p < 0.05$) lower during postpartum compared to prepartum. Similarly, Tharwat *et al.* (2015) [33] found significant decrease in cholesterol levels towards postpartum in goats. Contrary to the present findings, no significant difference in cholesterol concentration between pre and postpartum with mean values of 79.17 ± 12.30 and 63.50 ± 6.76 in HF cows. In the postpartum period there is an increase in circulating high density lipoproteins (HDL) as well decrease in low density lipoproteins (LDL), along with decrease in very low density lipoproteins; which contain the largest proportion of cholesterol amongst the lipoproteins (Bruss 1997) [8]. However the mean plasma cholesterol concentration recorded falls within the reference range of $62.1-192.5$ mg/dl for cattle (Kahn, 2005) [21]. This indicates that the values of our study are within the range. Although within the reference range, due to change in endocrine profile; lipid reserves are increased during the late pregnancy and utilized during lactation (Roche *et al.*, 2009; Piccione *et al.*, 2012) [29, 25].

The mean BUN concentration recorded in the present study

was found to be 22.33 ± 0.88 and 21.83 ± 0.60 mg/dl. The serum BUN concentration was significantly ($p < 0.05$) lower during postpartum compared to prepartum. Similar to our results, Tharwat *et al.* (2015) [33] found no significant change in levels of BUN between pre and postpartum in goats. Similarly, Seifi *et al.* (2007) [31] also reported no significant change of 2.63 ± 0.12 and 2.39 ± 0.11 mmol/l in prepartum and postpartum dairy cows respectively. In contrary to our findings, Piccione *et al.* (2012) [25] reported a significant increase ($P < 0.01$) during the postpartum in dairy cows with values of 16.00 ± 1.92 and 20.83 ± 4.30 mg/dl. The increase in urea levels during the lactation depends on the dietary protein intake (Roubies *et al.*, 2006; Piccione *et al.*, 2012) [30, 25] and feed intake (Bauchart, 1993; Seifi *et al.*, 2007) [6, 31]. However; the mean plasma BUN concentration recorded falls within the reference range of $7.8-24.6$ for cattle (Kahn, 2005) [21]. Quiroz-Rocha *et al.* (2009) [26] reported reference values of $2.1-8.0$ and $1.9-7.8$ mmol/lit in pre and post calving cows in Southern Ontario dairy farms. This indicates that the values of our study are within the range.

The mean serum creatinine concentration recorded in the present study was found to be 1.10 ± 0.11 and 1.20 ± 0.82 mg/dl. The creatinine concentration was significantly ($p < 0.05$) lower during postpartum compared to prepartum. Similar to our results, Tharwat *et al.* (2015) [33] found no significant change towards postpartum in goats. Piccione *et al.* (2012) [25] found that creatinine serum showed higher levels during the late pregnancy and early lactation of 1.21 ± 0.22 and 1.22 ± 0.24 mg/dl respectively. The mean plasma creatinine concentration recorded in the study animals agree with the reference value of $0.6-1.8$ for cattle (Kahn, 2005) [21]. Although within the reference range the values were on the higher side which could be due to the development of the foetal musculature and the load of organic waste of the newborn (Ferrell *et al.*, 1991; Piccione *et al.*, 2012) [11, 25].

The mean serum calcium concentration recorded in the present study was found to be 10.80 ± 0.81 and 8.07 ± 0.30 mg/dl. The serum calcium concentration was significantly ($p < 0.05$) lower during postpartum compared to prepartum. Similar to our results, Tharwat *et al.* (2015) [33] found significant decrease in calcium levels towards postpartum in goats. Piccione *et al.* (2012) [25] also reported significantly lower calcium concentration of 5.78 ± 1.00 during postpartum compared to prepartum with 7.75 ± 0.48 mg/dl. Quiroz-Rocha *et al.* (2009) [26] reported reference values of $2.18-2.65$ and $1.64-2.61$ mmol/lit in pre and post calving cows in Southern Ontario dairy farms. The drop in calcium concentration occurs at parturition or in the 1st days after calving (Goff, 2004) [13]. This is due to the onset of lactation, where calcium is drained in colostrum and milk (Goff, 2000) [14]. The mean serum phosphorous concentration recorded in the present study was found to be 4.72 ± 0.21 and 3.68 ± 0.17 mg/dl respectively. The serum phosphorus concentration was significantly ($p < 0.05$) lower during postpartum compared to prepartum. Similar to our results, Tharwat *et al.* (2015) [33] found significant decrease in levels towards postpartum in goats. Piccione *et al.* (2012) [25] also reported 5.67 ± 0.66 and 4.88 ± 0.86 mg/dl in prepartum and postpartum dairy cows. Quiroz-Rocha *et al.* (2009) [26] reported reference values of $1.48-2.65$ and $1.04-2.73$ mmol/lit in pre and post calving cows in Southern Ontario dairy farms. The decrease in phosphorus concentration after parturition could be due to initiation of lactation.

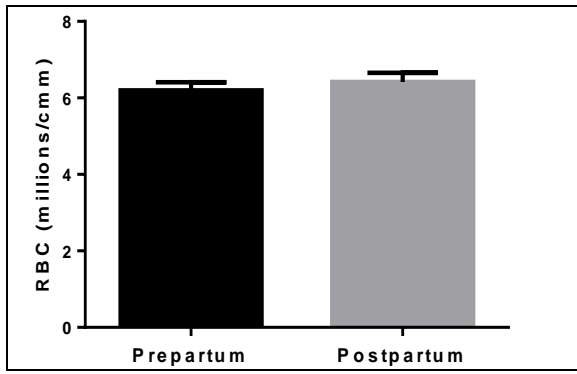


Fig 1

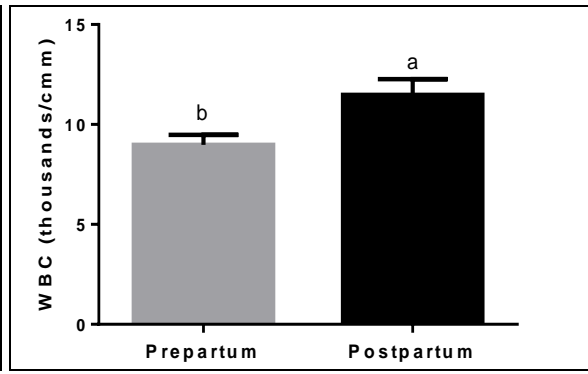


Fig 2

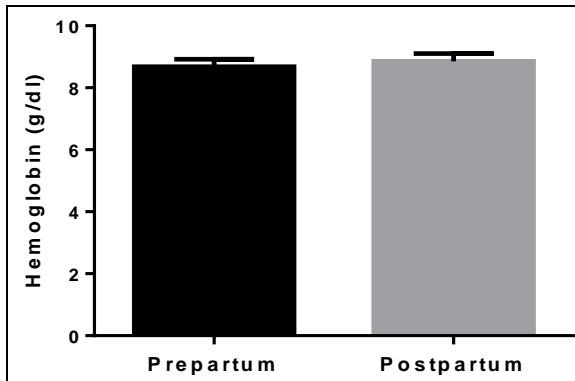


Fig 3

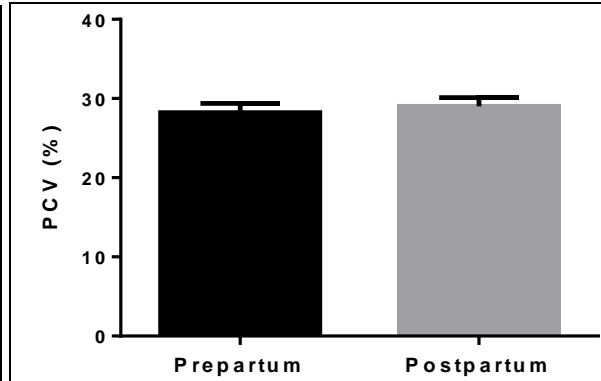


Fig 4

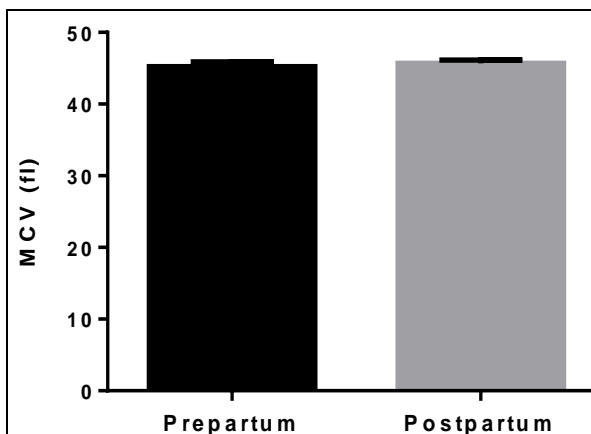


Fig 5

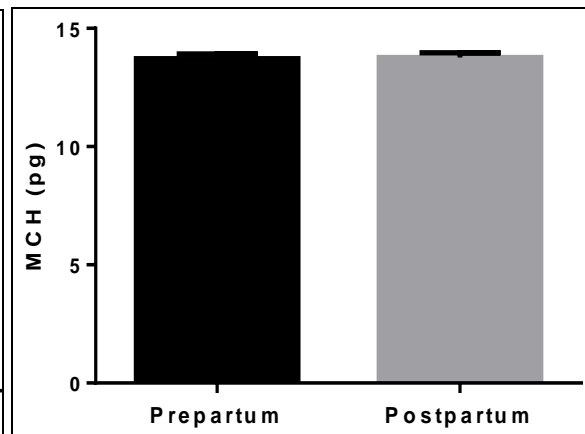


Fig 6

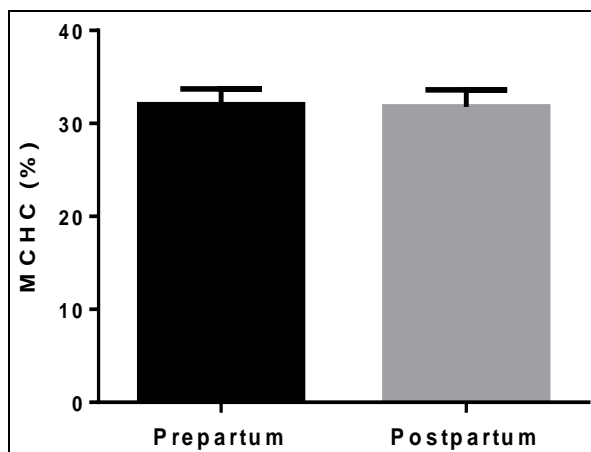


Fig 7

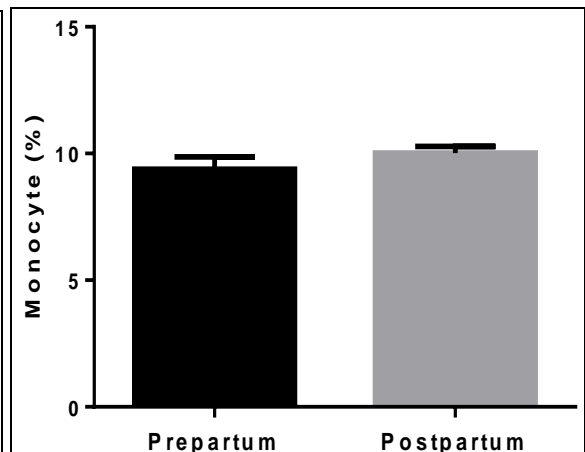


Fig 8

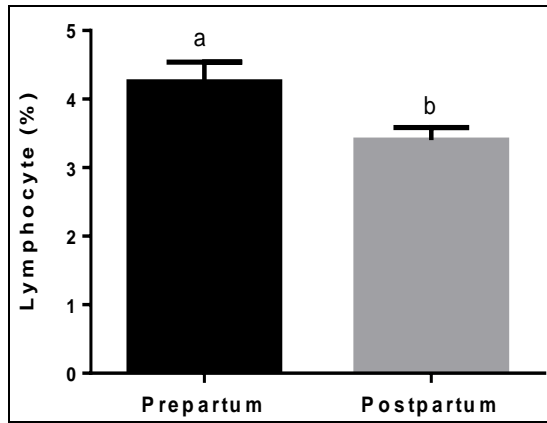


Fig 9

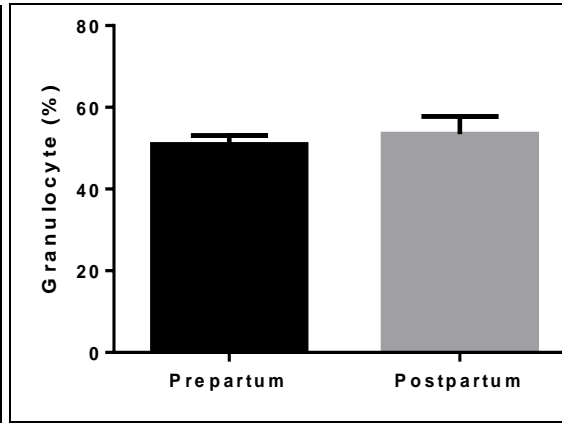


Fig 10

Fig 1 to 10: Shows the changes in hematological parameters from prepartum to postpartum during periparturient period in Ongole cows

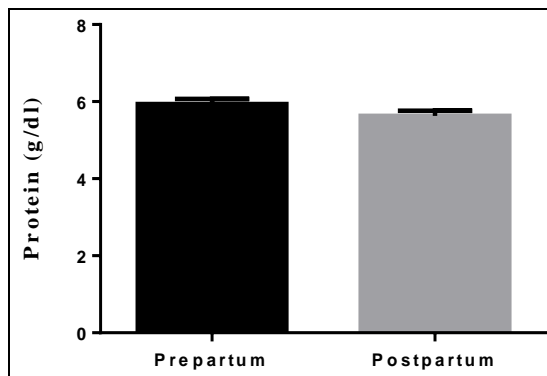


Fig 11

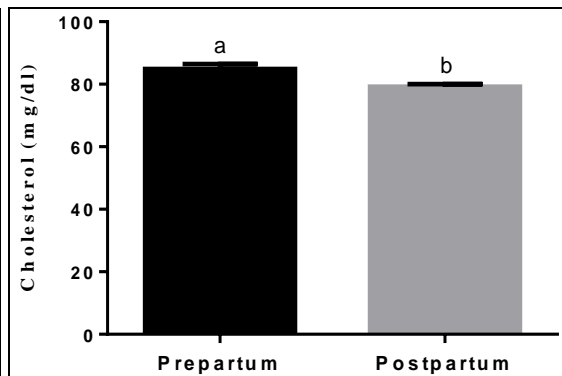


Fig 12

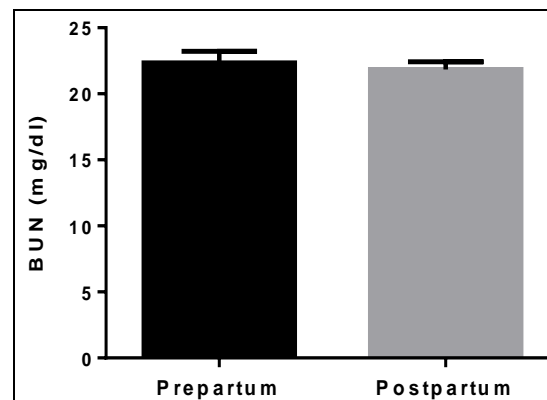


Fig 13

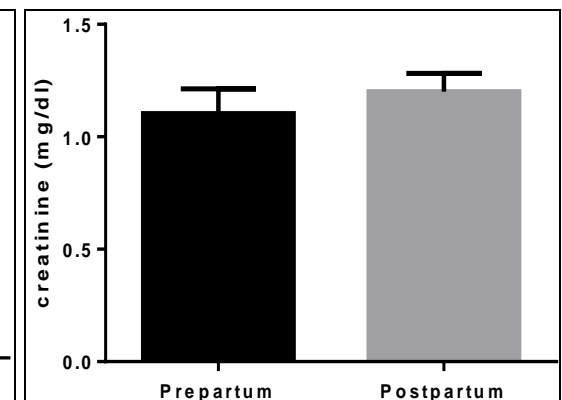


Fig 14

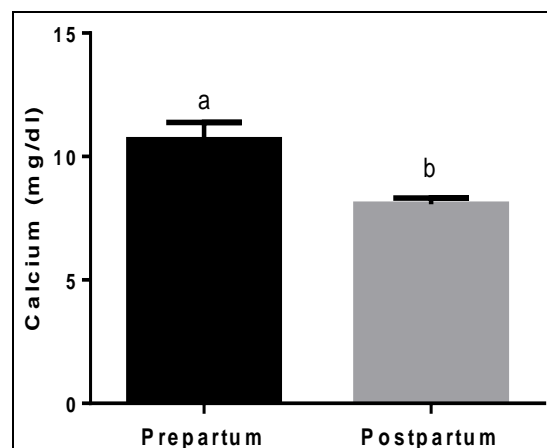


Fig 15

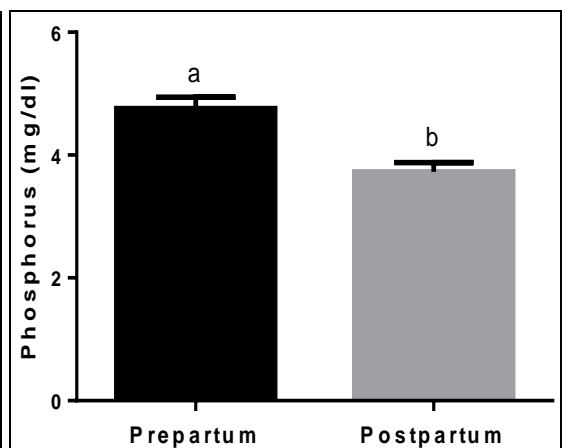


Fig 16

Fig 11 to 16: Shows the changes in biochemical parameters from prepartum to postpartum during periparturient period in Ongole cows

4. Conclusion

Periparturient period is the most stressful in the life of a dairy animal. Therefore estimation of hematological and biochemical parameters is of paramount importance to find out the physiological alterations and health status of the animals. A significant decrease in cholesterol, calcium, phosphorous, lymphocyte percent and a significant increase in TLC concentration during postpartum compared to prepartum is indicative of stressful condition during postpartum. These parameters could be used as markers to evaluate the quantum of stress during parturition in Ongole cows. However, the mean values obtained for various hematological and biochemical parameters were within the reference range despite of high THI which is further indicative of adaptability of Ongole cows to existing adverse ambience.

5. Acknowledgements

The research grant provided by NTR College of Veterinary Sciences, Gannavaram, is duly acknowledged.

6. Conflict of Interest

The authors declare that there is no any conflict of interest for this manuscript.

7. References

1. Abd Ellah MR, Hamed MI, Ibrahim DR, Rateb HZ. Serum biochemical and haematological reference intervals for water buffalo (*Bubalus bubalis*) heifers. *Journal of the South African Veterinary Association*. 2014; 85(1):01-07.
2. Abdelrazek H, Ismail TA, El-Azzazi FE, Elsayed DH. Hematological and Metabolic Alterations in Egyptian Buffaloes during Transition Period. *Egyptian Academic Journal of Biological Sciences. C, Physiology and Molecular Biology*. 2018; 10(1):69-78.
3. Abdulkareem TA. Some hematological and blood biochemical attributes of Iraqi riverine buffaloes (*Bubalus bubalis*) around calving and postpartum periods. *Al-Anbar Journal of Veterinary Sciences*. 2013; 6(1):143-150.
4. Ambily TR, Beena V, Karthiayini K, Ramnath V, Uma R Siddaramesh A. Effect of supra-nutritional supplementation of vitamin E and/or selenium on oxidant-antioxidant profile in transition dairy cows. *The Pharma Innovation Journal*. 2019; 8(8):97-106.
5. Baginski ES. *Clinica Chemica Acta*. 1973; 46(1):4-6.
6. Bauchart D. Lipid absorption and transport in ruminants. *Journal of Dairy Science*. 1993; 76(12):3864-3881.
7. Bowers LD. Kinetic serum creatinine assays I. The role of various factors in determining specificity. *Clinical Chemistry*. 1980; 26(5):551-554.
8. Bruss ML, Lipids and ketones. In *Clinical Biochemistry of Domestic Animals*. 5th edition. San Diego Academic Press, 1997, 83-115.
9. Dumas BT, Watson WA, Biggs HG. Albumin standards and the measurement of serum albumin with bromocresol green. *Clinica chimica acta*. 1971; 31(1):87-96.
10. Drackley JK. Biology of dairy cows during the transition period: the final frontier. *Journal of Dairy Science*. 1999; 82(11):2259-2273.
11. Ferrell CL. Maternal and fetal influences on uterine and conceptus development in the cow: II. Blood Flow and nutrient flux. *Journal of Animal Science*. 1991; 69(5):1954-1965.
12. Gavan C, Retea C, Motorga V. Changes in the hematological profile of Holstein primiparous in periparturient period and in early to mid-lactation. *Scientific Papers Animal Science and Biotechnologies*. 2010; 43(2):244-246.
13. Goff JP. Macromineral disorders of the transition cow. *Veterinary clinics of North America: Food Animal Practise*. 2004; 20(3):471-494.
14. Goff JP. Pathophysiology of calcium and phosphorus disorders. *Veterinary clinics of North America: Food Animal Practise*. 2000; 16(2):319-37.
15. Helal A, Hashem A, Abdel-Fattah M, El-Shaer H. Effect of heat stress on coat characteristics and physiological responses of Balady and Damascus goats in Sinai. *Egypt Am-Eurasian J Agric Environ Sci*. 2010; 7:60-69.
16. Ingvarsten KL, Andersen JB. Integration of metabolism and intake regulation: a review focusing on periparturient animals. *Journal of Dairy Science*. 2000; 83(7):1573-1597.
17. Jacor SK, Ramnath V, Philomina PT, Rahunandhanan KV, Kannan A. Assessment of physiological stress in periparturient cows and neonatal calves. *Indian journal of physiology and pharmacology*. 2001; 45(2):233-238.
18. Johnson HD, Ragsdale AC, Berry IL, Shanklin MD. Temperature-humidity effects including influence of acclimation in feed and water consumption of Holstein cattle. *University of Missouri Research Bulletin*, 1963, (846).
19. Kim IH, Na KJ, Yang MP. Immune responses during the peripartum period in dairy cows with postpartum endometritis. *Journal of Reproduction and Development*. 2005; 51(6):757-764.
20. Konvicna J, Vargova M, Paulikova I, Kovac G, Kostecka Z. Oxidative stress and antioxidant status in dairy cows during prepartal and postpartal periods. *Acta Veterinaria Brn*. 2015; 84(2):133-140.
21. Kahn CM, Editor. *The Merck veterinary manual*. Whitehouse Station, NJ: Merck, 2005.
22. Mader TL, Davis M, Brown-Brandl T. Environmental factors influencing heat stress in feedlot cattle. *Journal of Animal Science*. 2006; 84(3):712-719.
23. Mohamed GA, Abd-Elnaser EM, Elsayed HK. Preliminary study on lipid profile with relation to total antioxidant capacity and some hematological and biochemical changes of pre-post-partum buffalo heifers at assiut city. *Assiut Veterinary Medical Journal*. 2015; 61(144):159-165.
24. Mohri M, Sharifi K, Eidi S. Hematology and serum biochemistry of Holstein dairy calves: age related changes and comparison with blood composition in adults. *Research in Veterinary Science*. 2007; 83(1):30-39.
25. Piccione G, Messina V, Marafioti S, Casella S, Giannetto C, Fazio F. Changes of some haemato chemical parameters in dairy cows during late gestation, postpartum, lactation and dry periods. *Veterinarija Ir Zootechnika*. 2012; 58(80):59-64.
26. Quiroz-Rocha GF, LeBlanc SJ, Duffield TF, Wood D, Leslie KE, Jacobs RM. Reference limits for biochemical and hematological analytes of dairy cows one week before and one week after parturition. *The Canadian Veterinary Journal*. 2009; 50(4):383.
27. Ranjhan SK, editor. *Nutrient Requirements of Livestock and Poultry*. New Delhi: Indian Council of Agriculture

- Research, 1998.
28. Richmond W. Determination of cholesterol by CHOD-PAP method. *Clinical Chemistry*. 1973; 19:1350-1356.
 29. Roche JR, Frgens NC, Kay JK, Fisher MW, Stafford KJ, Berry DP. Invited review: Body condition score and its association with dairy cow productivity, health and welfare. *Journal of Dairy Science*. 2009; 92:5769-5801.
 30. Roubies N, Panouis N, Fytianou A, Katsoulos PD, Giadinis N, Karatzias H. Effects of age and reproductive stage on certain serum biochemical parameters of Chios sheep under greek rearing conditions. *Journal of Veterinary Medicine Series A*. 2006; 53(6):277-281.
 31. Seifi HA, Gorji-Dooz M, Mohri M, Dalir-Naghadeh B, Farzaneh N. Variations of energy-related biochemical metabolites during transition period in dairy cows. *Comparative Clinical Pathology*. 2007; 16(4):253-258.
 32. Snedecor GW, Cochran WG. *Statistical Methods*. Iowa State University Press, Ames, Iowa. 1967; 196:44-51.
 33. Tharwat M, Ali A, Al-Sobayil F. Hematological and biochemical profiles in goats during the transition period. *Comparative Clinical Pathology*. 2015; 24(1):1-7.
 34. Tharwat M, Takamizawa A, Hosaka YZ, Endoh D, Oikawa S. Hepatocyte apoptosis in dairy cattle during the transition period. *Canadian Journal of Veterinary Research*. 2012; (4):241-247.
 35. Wang J, Chen CC, Osaki S. *Clinical Chemistry*. 1983; 29:1255.
 36. Wood D, Quiroz-Rocha GF. *Schalm's Veterinary Hematology*, 2010, 829:4
 37. Young DS. *Effects of Drugs on Clinical Laboratory Tests*. Third Edition, 1990.