



ISSN (E): 2277- 7695
ISSN (P): 2349-8242
NAAS Rating: 5.03
TPI 2019; 8(9): 68-74
© 2019 TPI
www.thepharmajournal.com
Received: 14-07-2019
Accepted: 16-08-2019

Abhishek Parmar
Department of Animal
Nutrition, College of Veterinary
Science & Animal Husbandry,
Navsari Agricultural University,
Navsari, Gujarat, India

Vipul Patel
Department of Animal
Nutrition, College of Veterinary
Science & Animal Husbandry,
Navsari Agricultural University,
Navsari, Gujarat, India

Jignesh Patel
Department of Veterinary
Pathology, College of Veterinary
Science & Animal Husbandry,
Navsari Agricultural University,
Navsari, Gujarat, India

Sajani Usadadia
Department of Veterinary
Pathology, College of Veterinary
Science & Animal Husbandry,
Navsari Agricultural University,
Navsari, Gujarat, India

Sawan Rathwa
Department of Veterinary
Physiology and Biochemistry,
College of Veterinary Science &
Animal Husbandry, Navsari
Agricultural University, Navsari,
Gujarat, India

Dhruvil Prajapati
Department of Animal
Nutrition, College of Veterinary
Science & Animal Husbandry,
Navsari Agricultural University,
Navsari, Gujarat, India

Akshesh Rathva
Department of LPM, College of
Veterinary Science & Animal
Husbandry, Navsari Agricultural
University, Navsari Gujarat,
India

Correspondence
Abhishek Parmar
Department of Animal
Nutrition, College of Veterinary
Science & Animal Husbandry,
Navsari Agricultural University,
Navsari, Gujarat, India

Quercetin, a health promising phytoadditive for poultry production: Trends & advances

Abhishek Parmar, Vipul Patel, Jignesh Patel, Sajani Usadadia, Sawan Rathwa, Dhruvil Prajapati and Akshesh Rathva

Abstract

Plants sources are densely rich with the polyphenolic compound. Among that flavonoid is the major group found and due to their immense availability have rising a great interest for its multidimensional properties. Quercetin, the most plentiful flavonoid has combined properties of growth promoter, antioxidant, anti-inflammatory, anti-lypolytic, potent hepatoprotective, immune boosting and gut health enhancing potential. Their strong antioxidant activity was due to presence of the central C-ring along with high number of hydroxyl groups and conjugated π orbitals in its chemical structure. A substantial amount of literature on different animal studies with positive effects on growth, antioxidant status, hepatoprotection, immunomodulation, gut morphology, gut microbiota with ultimate improvement in gut health and birds health status. Current trends have been focused to find a natural phytoadditive with having a multidimensional action and potentially health promoting effects in animals and birds by nutritionist. Therefore, the aim of this review is to explore the biological actions of quercetin, aware the medicinal importance and highlighting that it can be a safe and excellent phytoadditive for the fast-growing poultry industry as an alternative of banned synthetic growth promoting agents for the welfare of birds.

Keywords: Antioxidant, flavonoid, hepatoprotectant, poultry, quercetin

Introduction

Supplementation of antibiotic growth promoter has banned by the European union (EU) in poultry industry (El-Hack *et al.*, 2016) [14]. The possible toxic effect of synthetic feed additive has enhanced the interest of poultry nutritionist to seek alternative originated from plant sources. Currently inclination to improve growth, feed utilization, meat and egg quality, immune system, gut environment and health of avian species including Turkeys, broilers and layers (Salah *et al.*, 2015; Orayaga *et al.*, 2016; Aguihe *et al.*, 2017) [59, 48, 5] through natural feed additives. Antioxidants are the agents, essential for poultry production as they decrease lipid peroxidation, improve the organoleptic characteristics, nutritional value of eggs, meat and can extend shelf life (Fellenberg and Speisky, 2006) [18]. More emphasis has been given to herbal source of additive having combine property of antioxidant, lipolysis, and gut modulating and immunomodulatory activity.

Flavonoids have aroused enormous interest in the preceding decades, due to its multidimensional health promoting effects on human, animals and omnipresence in plant kingdom. They are known to as “functional ingredients” and “health promoting biomolecules” (Nijveldt *et al.*, 2001; Kamboh *et al.*, 2015) [46, 29]. Quercetin is a major concern when studies about flavonoids, Quercetin (3,3",4",5,7-pentahydroxyflavone) is polyphenolic flavonoid compounds found almost ubiquitous in plants and plant foods sources having strong antioxidant property as free radical terminators (Sikder *et al.*, 2014) [63]. Presence of central C-ring along with high number of hydroxyl groups and conjugated π orbitals makes the compound strong reducing agents (Rice - Evans *et al.*, 1996) [57]. It is found in various foods such as vegetables, tea, fruits, wine apples and onions, tomatoes and is known to exert positive effect on poultry production and health (Saeed *et al.*, 2017) [58]. Being a powerful free radical scavenger, it ameliorates the organ damage, inhibit effect of oxidative stress (Yi *et al.*, 2011 and Sikder *et al.*, 2014) [73, 63] possess immunomodulatory and anti-inflammatory properties (Serafini *et al.*, 2010) [61]. Thus, the aim of this review is to highlighting the beneficiary potential and effects of quercetin as a phytoadditive in poultry health and production.

Chemical Structure

The chemical name of quercetin is 2-(3, 4-dihydroxyphenyl)-3, 5, 7-trihydroxy-4 Hchromen-4-one (Quercetin, C₁₅H₁₀O₇) and its derivatives structure are shown in Figure 1. The different alternative names for quercetin are sophoretin, meletin, quercetine, xanthaurine, quercetol, quercitin, quertine and flavinmeletin (Saeed *et al.*, 2017) [58]. Quercetin as a natural antioxidant, polyphenol and is highly lipophilic and slightly hydrophilic properties. It is yellow colored in powder form practically insoluble in water, but soluble in aqueous alkaline solutions. The melting point of quercetin is 316 °C or 601°F and its molar mass is 302.23 g/mol (Ades, 2009) [3]. Presence of central C-ring along with high number of hydroxyl groups and conjugated π orbitals makes the compound strong reducing agents (Rice-Evans *et al.*, 1996) [57]. Quercetin and its glycosidic metabolites possess cell signaling pathways and having high reducing power, effectively neutralized the ROS reduction thereby oxidative DNA damage (Wilms *et al.*, 2005) [72]. Therefore, all consequent ROS mediated events like hepatic inflammation and subsequent liver damage were efficiently inhibited by quercetin.

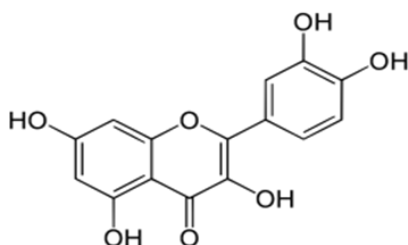


Fig 1: Molecular structure of Quercetin

Sources of Quercetin

Quercetin is a common flavonoid polyphenol compound enriched in most of vegetables and fruits in the form of glycosides (Alrawaiq and Abdullah, 2014) [7]. Synthetic production of flavonoids has not been practiced yet hence plants are the only major source of quercetin (Abdelmoaty *et al.*, 2010) [1]. It is widely distributed in plant sources such as onions, moringa, tomatoes, garlic, ginger, broccoli, barriers, tea, citrus fruits, brassica, cocoa, capers, apples, grapes, olive oil, flowers, nuts and leaves etc. (Saeed *et al.*, 2017) [58] as describe in Table 1.

Absorption and Metabolism of Quercetin

Quercetin is a glycosides, after oral ingestion efficiently hydrolyzed in the small intestine by beta-glucosidases to the

aglycone form and much of which is then absorbed (Scholz *et al.*, 2007) [60]. Once it absorbed, further processed in epithelial cells of the stomach and intestines by phase II enzymes (Alrawaiq and Abdullah, 2014) [7]. Quercetin and its derivatives are very stable against gastric acid in the gastrointestinal tract but evidences suggest its absorption site in the upper small intestine (Saeed *et al.*, 2017) [58]. Its absorption is influenced by differences in its glycosylation, the food sources from which it is ingested and dietary components such as fiber and fat (Guo *et al.*, 2013) [24]. After absorption, it will be got metabolized in different organs including the small intestine, colon, liver and kidney (Li *et al.*, 2016) [35]. The biotransformation enzymes generate different metabolites of quercetin includes the methylated, phenolic acid, sulfo-substituted and glucuronidated forms in the small intestine and liver (Day *et al.*, 2000) [12]. Studies suggest the highest accumulation of these metabolites lung, liver and kidneys in different animal models (Li *et al.*, 2016) [35]. These metabolites acted as an antioxidant through the resistance low-density lipoprotein (LDL) oxidation (Manach *et al.*, 1997) [39]. Quercetin metabolites concentrations were in the liver were lower than those in plasma, and the methylated (90%-95%) form were highest in hepatic metabolites (Manach *et al.*, 1999) [41]. Accumulation of quercetin is rising over continuous ingestion of quercetin rich sources which was highly correlated with dietary contents. Final, excretion of it, is being done by kidney (Li *et al.*, 2016) [35].

Biological Effects of Quercetin

Quercetin as a Growth Enhancer

Supplementation of quercetin in poultry ration has a better growth promoting and feed utilizing effect in birds as reported in previous studies. Liu *et al.* (2014) [37] had found improvement in feed conversion ratio with level of 0.367 to 0.369 g quercetin/kg of feed in laying hens, also found quadratic correlation with the level of quercetin (P=0.05) in laying rate and it was maximized by the supplementation level of 0.2 g/kg of diet. In same trend, quercetin @ 100, 200 and 300 mg/kg feed in combination with α -tocopherol feed imparted significant effect on gain in weight and feed conversion efficiency in broilers (Sohaib *et al.*, 2015) [64]. Similarly, Ramamneh, (2017) [54] noted improved chicken performance with high (P<0.001) weight gain, better feed conversion ratio with onion extract inclusion in broilers. That is attributed to a sulphur-containing organic compound known as Allicin, which characterized by antimicrobial activity that could be responsible for the growth promoting effect of onion (Ramamneh, 2017) [54]. Consistence better growth

Table 1: Various food sources and Quercetin contents (mg/100g).

| Food sources | | | |
|----------------------------|-------------------|-------------------------|-------------------|
| Vegetables | Quercetin content | Fruits | Quercetin content |
| Ancho pepper | 27.61 | Unsweetened apple sauce | 2.01 |
| Raw broccoli | 3.22 | Raw apple with skin | 4.43 |
| Cooked broccoli | 1.07 | Raw apricot | 2.54 |
| Red raw onion | 19.94 | Raw bilberry | 3.05 |
| Boiled onions | 19.37 | Black grapes | 2.56 |
| Loose-leaf lettuce | 1.94 | Frozen blueberry | 3.94 |
| Raw kale | 7.72 | Raw blueberry | 3.12 |
| Canned kale | 4.51 | Raw cherry | 1.24 |
| Chilli | 16.8 | Raw cowberry | 21.01 |
| Iceberg lettuce | 2.48 | Canned sweet cherry | 3.21 |
| Hot wax raw yellow pepper | 50.62 | Cherry raw tomato | 2.78 |
| Hot green raw chili pepper | 16.81 | Raw cranberry | 14.01 |

| | | | |
|-----------------|------|------------------------|-------|
| Raw green beans | 2.74 | Raw cranberry juice | 16.42 |
| Green tea | 2.00 | Frozen chokeberry | 8.91 |
| Capers | 234 | Raw lemon without peel | 2.3 |

(Source: Muhammad *et al.*, 2017) ^[45]

Performance and improvement in feed conversion efficiency was observed by Hassan *et al.* (2018) ^[25] with of rutin (i.e., rutoside, vitamin P, quercetin-3-rutinoside or sophorin) as a flavonolglycoside supplementation in broilers. These improved growth performance, likely due to favorable effect of flavonoids on gut morphology and the functional architecture of the small intestine (Awad *et al.*, 2011 ^[9]; Viveros *et al.*, 2011 ^[71]). Moreover, flavones up-regulate the combination of the growth hormone and the hepatic growth hormone receptor, which increases insulin-like growth factor 1 concentrations, thereby promoting animal growth (Ouyang *et al.*, 2016) ^[49]. In general, dietary flavonoid supplementation has been reported to improve the growth performance of broiler chickens (Starcevic, 2015) ^[65].

Quercetin as a Potent Antioxidant/Anti-Stressor

Health enhancing properties of polyphenol were directly related to their antioxidant (i) an ortho-dihydroxy or catechol group in the B-ring, (ii) a 2, 3, double bond, and (iii) hydroxyl substitution at position 3 and 5 in their chemical structure (Ozgen *et al.*, 2016) ^[50]. Experimental studies suggested enhanced endogenous antioxidant enzyme activities by quercetin and possible damages to the vital organs in diabetic rat (Elik *et al.*, 2007) ^[15]. Alike to that Zou *et al.* (2015) ^[75] reported free radical scavenging resultant oxidative stress preventing and anti-inflammatory effect of quercetin against the perfluorooctanoic acid induced liver damage. The free radical generating ROS (reactive oxygen species) is the major etiological factors responsible for many metabolic disorders (David *et al.*, 2017) ^[11]. Flavonoid supplementation also have protective mechanism in terms of decreased liver MDA (malondialdehyde) due to down regulation of the expression of TNF- α (tumor necrosis factor) and NF- κ B (necrosis factor) thereby the increased antioxidant capacity and mitigate the inflammation (Zengpeng *et al.*, 2018; Abu Hafsa and Ibrahim, 2018) ^[74, 2]. Similarly, either quercetin or β -sisterol reflect improved antioxidant status and protective effect in dietary induced dyslipidemic and hepatotoxic mice noticed by Marcolin *et al.* (2013) ^[42], Sikder *et al.* (2014) ^[63] and Iskender and Yenice, (2016) ^[28]. They suggested that quercetin (0.5 g/kg diet) being superiorly decreased MDA concentration and increased (glutathione peroxidase) GSH-Px, GR (glutathione reductase), GST (glutathione S-transferase), and SOD (super oxidizedismutase) activities and GSH (glutathione reduced) level in laying hens. Further, Oliveira *et al.* (2014) ^[47] reported that quercetin (50 mg/kg BW) reduce the LPO (lipid peroxidation) and SOD concentration and increased the GpX level in polychlorinated phenyl induced liver injury in Wister rat. Hassan *et al.* (2018) ^[25] observed that a high concentration of rutin (0.5 and 1 g/kg) supplementation in broilers, significantly reduced malondialdehyde (MDA) concentrations (P=0.001), with increased superoxide dismutase (SOD), catalase and glutathione peroxidase (GpX) activity and ameliorate the oxidative stress in broiler chickens. As in poultry birds are so sensitive to various stresses that ultimately causing various health alteration, being a strong antioxidant potential of quercetin seems to be effective ameliorating agents as phytoadditive in the poultry sector.

Quercetin as a Potent Hepatoprotective Agent

Numerous experiments have been reported the hepatoprotective activity of quercetin in liver damages (Table 2). Quercetin administration significantly decreased liver triglycerides, liver and serum total cholesterol, serum low-density lipoprotein cholesterol, and serum high density lipoprotein cholesterol (HDL-C) along with reduced (P<0.05) the elevated activity of serum alanine aminotransferase, aspartate aminotransferase and γ -glutamyltranspeptidase (GGT) in hepatic lipemic-oxidative injury, in addition also ameliorate the steatohepatitis in rats fed with a high cholesterol diet (HCD) reported by Mariee *et al.* (2012) ^[43]. Consistently, Porras *et al.* (2017) ^[53] and Gedikli *et al.* (2017) ^[20] were noted reduction in higher plasma triglyceride and ALT levels and protective effect of quercetin with high fat diet induced liver damages with alleviate the histomorphological changes and inflammatory activities in liver in rats or mice models, respectively. Corroborated hepatoprotective action was suggested by El-Faras *et al.* (2017) ^[13] in rats with paracetamol induced toxicity in rats. Overall the evidences suggesting anti-lypolytic and a potent hepatoprotective potential of quercetin due to the its chemical structure as strong free radical scavenger with reducing lipid peroxidation potential.

Quercetin as an Immunomodulators

As a resultant of oxidative stress, various metabolic processes generated wide range of the reactive species (RS) either reactive oxygen species (ROS) or reactive nitrogen species (RNS) in the body (Rehman *et al.*, 2018) ^[55]. The ROS play vital roles in the signaling pathways, cytokine transcription, immunomodulation, ion transport, and apoptosis (Gloire *et al.*, 2006 ^[21] Rehman *et al.*, 2018) ^[56]. Over production of ROS may causing damage to DNA, protein, and lipid structures, leading to the disruption of the cell functions (Markesbery *et al.*, 2013) ^[44]. Non-specific response a stress, one of the most pesky issues in the modern poultry industry causing compromised immune status, health, growth and productive performance of poultry birds (Surai *et al.*, 2016) ^[67]. Dietary polyphenol with having potent antioxidant potential not only stimulate immune response of birds additionally that also leads a modulation of detoxification enzymes, scavenging of oxidative agents and regulation of gene expression in cells (Kamboh *et al.*, 2015) ^[29]. Flavonoids regulate mucosal and cellular immunity and modulate the endocrine and circulatory markers of health; dietary supplementation with flavonoids can be, therefore, used for improving immunity and health of broiler chickens (Kamboh *et al.*, 2018) ^[30]. Numerous literatures suggested a immunostimulant activity of quercetin. In these concern, Huang *et al.* (2010) ^[26] reported quercetin effectively inhibited LPS (Lypopolysaccharides)-induced dendritic cells (DC) activation by reducing the production of proinflammatory cytokines/chemokines and the expression levels of MHC (Major histocompetability complex) class II and costimulatory molecules, additionally also abrogated the ability of LPS-stimulated DCs to induce Ag (Antigen)-specific T cell activation, both *in vitro* and *in vivo* study. Manach *et al.* (2004) ^[40] reported immune regulatory activity through enhancing Ig Y antibody titer, lymphoid

Table 2: Biological Activities of Quercetin/Flavonol.

| Activities | Flavonol/Quercetin Compound | Reference |
|---------------------|------------------------------|--|
| Hepatoprotective | Quercetin | Amalia <i>et al.</i> , 2007 ^[8] ; Vieira <i>et al.</i> , 2011 ^[70] ; Shizuma, 2014 ^[62] |
| Antilipid/Lypolytic | Quercetin | Ahn <i>et al.</i> , 2008 ^[6] ; Lee <i>et al.</i> , 2011 ^[33] |
| | Onion powder | Chandra <i>et al.</i> , 2013 ^[10] ; Emamat <i>et al.</i> , 2016 ^[16] |
| | Grape seed powder (Flavonol) | Adisakwattana <i>et al.</i> , 2010 ^[4] ; Fawzia <i>et al.</i> , 2014 ^[17] ; Abu Hafsa and Ibrahim, 2018 ^[2] |
| | Genistein | Park <i>et al.</i> , 2013 ^[52] ; Zhenpeng <i>et al.</i> , 2018 ^[75] |
| Anti inflammatory | Rutin | Liu <i>et al.</i> , 2017 ^[38] ; Hassan <i>et al.</i> , 2018 ^[25] |
| | Quercetin | García-Mediavilla <i>et al.</i> , 2007 ^[19] ; Stewart <i>et al.</i> , 2008 ^[66] ; Panchal <i>et al.</i> , 2012 ^[51] |

Organs weights and natural killer cells activity and resultant better performance of animals. Alike to that Konard *et al.* (2011) ^[32] noted a positive impact on immune parameters includes total blood leucocytes count and proinflammatory cytokines (IL-6, TNF α , GM-CSF, IFN γ , IL-1 β , IL-2, IL-8 and IL-10) in humans fed with quercetin. Similarly, Liu *et al.* (2012) ^[36] confirmed the immunostimulant function of quercetin with better effect on IL-8, IL-1 and IL-1 level in rats. Further, enhancing the IgY level showing a promoting effect of quercetin on mucosal immunity in broilers (Hager-Theodorides *et al.*, 2014) ^[25].

Quercetin as a Gut Modulator

Dietary bioactive constituents directly or indirectly purported to act on continuously interacting elements that define gut ecology such as gut microbiota composition and metabolic activity, gut integrity and inflammatory status (Suzuki and Hara, 2011; Lee *et al.*, 2017) ^[69, 34]. The status of the gut and its microscopic structure are good indicators of the stress and nutrient response of the intestinal tract (Viveros *et al.*, 2011) ^[71]. The increased villus surface area (height or width) is an indication of increasing the absorption capacity of mucosa for the available nutrients which regulates the nutritional status and ultimate health of birds (Kamboh *et al.*, 2015) ^[29]. As the crypt is known to be a villus factory, a large crypt depth suggesting a rapid tissue turnover and a high demand for generation of new tissues. The intestinal epithelial cells originate at the base of crypts as immature proliferative cells, differentiate and migrate along the villus surface upward to the villus tip and are finally extruded into the intestinal lumen (Hu and Guo, 2007) ^[27].

Few literatures are available for gut modulatory activity of quercetin. Although, Zou *et al.* (2016) ^[76] reported a positive effect on intestinal morphology with increased jejunal villus height in pigs with stressful condition. In the ceecal digesta, birds fed grape pomace concentrate and grape seed extract diets had higher populations of *Escherichia coli*, *Lactobacillus*, *Enterococcus*, and *Clostridium* than birds in any other treatment group. Viveros *et al.* (2011) ^[71] noted a higher microbial count such as *Escherichia coli*, *Lactobacillus*, *Enterococcus*, and *Clostridium* and modulatory effect on gut morphology in broilers fed with grapes polyphenols. In same line, Kamboh and Zhu, (2014) ^[31] suggest a flavonoid such as genistein and hesperidin was promoted intestinal morphology and absorptive functions of growing broilers. This gut modulating activity is not exactly known for quercetin but it might be due to the polyphenolic structure and strong antioxidant potential of quercetin. Thus, these seems to prove that quercetin can be a very good flavonol with improving gut morphology and microbiota, thereby gut health and performance of poultry birds.

Conclusions

The current evidences suggesting that, among the various phytochemical flavonoids, quercetin could be very effective ROS scavengers due to its chemical structural characters in poultry birds. This quercetin possesses multifarious biological actions including growth enhancer, antioxidant, anti-inflammatory, anti-lypotropic, potent hepatoprotective, immunomodulating and gut modulating activities. These positive actions and the present scientific findings supports that quercetin can be an excellent natural phytoadditive and would be a promising in modern poultry industry to alleviates the sparking issues of this sector, the stress by improvement in health, production and welfare of chickens. However, further studies is needed to decide incorporation level in poultry ration and found an ideal practical techniques for extraction additionally to find economically feasible sources of quercetin to reach up by the farmers as well as distributed it at a commercial level in poultry production.

References

1. Abdelmoaty MA, Ibrahim MA, Ahmed NS, Abdelaziz MA. Confirmatory studies on the antioxidant and antidiabetic effect of quercetin in rats. *Indian Journal of Clinical Biochemistry*. 2010; 25(2):188-192.
2. Abu Hafsa SH, Ibrahim SA. Effect of dietary polyphenol-rich grape seed on growth performance, antioxidant capacity and ileal microflora in broiler chicks. *Journal of Animal Physiology and Animal Nutrition*. 2018; 102:268-275.
3. Ades TB. Quercetin American Cancer Society Complete Guide to Complementary and Alternative Cancer Therapies (2nd edition). American Cancer Society, 2009.
4. Adisakwattana S, Moonrat J, Srichairat S, Chanasit C, Tirapongporn H, Chanathong B, Sapwarobol S. Lipid-Lowering mechanisms of grape seed extract (*Vitis vinifera* L) and its antihyperlipidemic activity. *Journal of Medicinal Plants Research*. 2010; 4:2113-2120.
5. Aguihe PC, Kehinde AS, Ospina-Rojas IC, Murakami AE. Comparative effect of different detoxified rubber seed meal on haematological and serum biochemical indices of broilers. *Journal of Animal Health and Production*. 2017; 5(2):50-57.
6. Ahn J, Lee H, Kim S, Park J, Ha T. The anti-obesity effect of quercetin is mediated by the AMPK and MAPK signaling pathways. *Biochemical and Biophysical Research Communications*. 2008; 373(4):545-549.
7. Alrawaiq NS, Abdullah A. A Review of Flavonoid Quercetin: Metabolism, Bioactivity and Antioxidant Properties. *International Journal of Pharm Tech Research*. 2014; 6(3):933-941.
8. Amalia PM, Possa MN, Augusto MC, Francisca LS. Quercetin prevents oxidative stress in cirrhotic rats.

- Digestive Diseases and Sciences. 2007; 52:2616-2621.
9. Awad WA, Ghareeb K, Bohm J. Evaluation of the chicory inulin efficacy on ameliorating the intestinal morphology and modulating the intestinal electrophysiological properties in broiler chickens. *Journal of Animal Physiology and Animal Nutrition*. 2011; 95(1):65-72. [http:// dx.doi.org/10.1111/j.1439-0396.2010.00999.x](http://dx.doi.org/10.1111/j.1439-0396.2010.00999.x)
 10. Chandra A, Sabharwal R, Chander R, Mahdi F, Mahdi AA. Effect of some Indian herbs on dyslipidemia in streptozotocin induced diabetic Rats. *International journal of Medical and Dental Sciences*. 2013; 2(1):24-33.
 11. David AVA, Arulmoli R, Parasuraman S. Overviews of biological importance of quercetin: A bioactive flavonoid. *Pharmacognosy Review*. 2016; 10(20):84-89.
 12. Day AJ, Bao Y, Morgan MR, Williamson G. Conjugation position of quercetin glucuronides and effect on biological activity. *Free Radical Biology & Medicine*. 2000; 29:1234-1243.
 13. El Faras AA, Elsawaf AL. Hepatoprotective activity of quercetin against paracetamol induced liver toxicity in rats. *Tanta Medical Journal*. 2017; 45:92-98.
 14. El-hack MEA, Alagawany M, Farag MR, Tiwari R, Karthik K, Dhama K, Zorriehzahra J *et al*. Beneficial impacts of thymol essential oil on health and production of animals, fish and poultry: a review. *Journal of Essential Oil Research*. 2016; 28(5):365-382. <https://doi.org/10.1080/10412905>.
 15. Elik M, Serdaroglu G, Ozkan R. The investigation of antioxidant activities of myricetin and quercetin with different methods, C.U. Fen-Edebiyat Fakultesi Fen Bilimleri Dergisi. 2007; 28:2.
 16. Emamat H, Foroughi F, Zinab HE, Taghizadeh M, Rismanchi M, Hekmatdoost A *et al*. The effects of onion consumption on treatment of metabolic, histologic, and inflammatory features of nonalcoholic fatty liver disease. *Journal of Diabetes & Metabolic Disorders*. 2016; 15:25.
 17. Fawzia AH, Khalid M, Basyony MM. Influence of grape seeds powder as a natural antioxidant on growth performance, antioxidant status and carcass characteristics of rabbits under hot conditions. *Proceedings of the 7th International Conference on Rabbit Production in Hot Climate, 8–12 September, 2014, Marsa Alam, Egypt*. 2014; 395-412.
 18. Fellenberg MA, Speisky H. Antioxidants: their effects on broiler oxidative stress and its meat oxidative stability. *World's Poultry Science Journal*. 2006; 62:53-70.
 19. García-Mediavilla V, Crespo I, Collado PS, Esteller A, Sanchez-Campos S, Tunon MJ *et al*. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappa B pathway in Chang Liver cells. *European Journal of Pharmacology*. 2007; 557(2-3):221-229.
 20. Gedikli S, Ozkanlar S, Gur C, Sengul E, Gelen V. Preventive effects of quercetin on liver damages in high-fat diet-induced obesity. *Journal of Histology & Histopathology*. 2017; 4:7.
 21. Gloire G, Legrand-Poels S, Piette J. NF- κ B activation by reactive oxygen species: fifteen years later. *Biochemical Pharmacology*. 2006; 72(11):1493-1505.
 22. Gong J, Chen SS. Polyphenolic antioxidants inhibit peptide presentation by antigen-presenting cells. *International Immunopharmacology*. 2003; 3:1841-1852.
 23. Guo Y, Mah E, Davis CG, Jalili T, Ferruzzi MG, Chun OK *et al*. Dietary fat increases quercetin bioavailability in overweight adults. *Molecular Nutrition & Food Research*. 2013; 57:896-905.
 24. Hager-Theodorides, Ariadne, Goliomytis, Michael, Delis S, Deligeorgis *et al*. Effects of dietary supplementation with quercetin on broiler immunological characteristics. *Animal Feed Science and Technology*. 2014; 198. 10.1016/j.anifeedsci.2014.09.021.
 25. Hassan FAM, Roushdy EM, Kishawy ATY, Zagloul AW, Tukur HA, Saadeldin IM *et al*. Growth performance, antioxidant capacity, lipid-related transcript expression and the economics of broiler chickens fed different levels of rutin. *Animals*. 2018; 9:7-11.
 26. Haung RY, Yu YL, Cheng WC, Yang CNO, Fu E, Chu CL *et al*. Immunosuppressive effect of quercetin on dendritic cell activation and function. *The Journal of Immunology*. 2010; 184:6815-6821.
 27. Hu Z, Guo Y. Effects of dietary sodium butyrate supplementation on the intestinal morphological structure, absorptive function and gut flora in chickens. *Animal Feed Science Technology*. 2007; 132(3-4):240-249.
 28. Iskender H, Yenice G, Dokumacioglu E, Kaynar O, Hayirli A, Kaya A *et al*. The effects of dietary flavonoid supplementation on the antioxidant status of laying hens. *Brazilian Journal of Poultry Science*. 2016; 18:663-668.
 29. Kamboh AA, Arain MA, Mughal MJ, Zaman A, Arain ZM, Soomro AH *et al*. Flavonoids: Health promoting phytochemicals for animal production- a review. *Journal of Animal Health and Production*. 2015; 3(1):6-13.
 30. Kamboh AA, Khan MA, Kaka U, Awad EA, Memon AM, Saeed M *et al*. Effect of dietary supplementation of phytochemicals on immunity and haematology of growing broiler chickens. *Italian Journal of Animal Science*. 2018; 17:4. 10381043.
 31. Kamboh AA, Zhu WY. Individual and combined effects of genistein and hesperidin on immunity and intestinal morphometry in lipopolysaccharide-challenged broiler chickens. *Poultry Science*. 2014; 93(9):1-9. [http://dx.doi.org/10.3382/ ps.2014-03971](http://dx.doi.org/10.3382/ps.2014-03971)
 32. Konrad Manuela, Nieman David, Henson A, Dru, Kennerly M, Krista Jin, Fuxia Holasek Sandra *et al*. The Acute Effect of Ingesting a Quercetin-Based Supplement on Exercise-Induced Inflammation and Immune Changes in Runners. *International journal of sport nutrition and exercise metabolism*. 2011; 21:338-46. 10.1249/01.MSS.0000402819.42267.6d.
 33. Lee KH, Park E, Lee HJ, Kim MO, Cha YJ, Kim JM *et al*. Effects of daily quercetin-rich supplementation on cardio metabolic risks in male smokers. *Nutrition Research and Practice*. 2011; 5(1):28-33.
 34. Lee SA, Apajalahti J, Vienola K, Gonzalez-Ortiz G, Fontes CMGA, Bedford MR *et al*. Age and dietary xylanase supplementation affects ileal sugar residues and short chain fatty acid concentration in the ileum and caecum of broiler chickens. *Animal Feed Science Technology*. 2017; 234:29-42.
 35. Li Y, Yao J, Han C, Yang J, Chaudhry MT, Wang S *et al*. Quercetin, inflammation and immunity. *Nutrients*, 2016; 8(167). Doi: 10.3390/nu8030167.
 36. Liu H, Zhang L, Lu SP. Evaluation of antioxidant and

- immunity activities of quercetin in isoproterenol-treated rats. *Molecules*. 2012; 17:4281-4291. doi: 10.3390/molecules17044281.
37. Liu HN, Liu Y, Hu LL, Suo YL, Zhang L, Jin F *et al*. Effects of dietary supplementation of quercetin on performance, egg quality, ceecal microflora populations, and antioxidant status in laying hens. *Poultry Science*. 2014; 93(2):347-353.
 38. Liu Q, Pan R, Ding L, Zhang F, Hu L, Ding B *et al*. Rutin exhibits hepatoprotective effects in a mouse model of non-alcoholic fatty liver disease by reducing hepatic lipid levels and mitigating lipid-induced oxidative injuries. *International Immunopharmacology*. 2017; 49:132-141.
 39. Manach C, Morand C, Demigne C, Texier O, Regerat F, Remesy C *et al*. Bioavailability of rutin and quercetin in rats. *FEBS letters*, 1997; 409(1):12-16.
 40. Manach C, Scalbert A, Morand C, Remesy C, Jimenez L. Polyphenols: Food sources and bioavailability. *American Journal of Clinical Nutrition*. 2004; 79:727-747.
 41. Manach C, Texier O, Morand C, Crespy V, Regerat F, Demigne C *et al*. Comparison of the bioavailability of quercetin and catechin in rats. *Free Radical Biology & Medicine*. 1999; 27:1259-1266.
 42. Marcolin E, Forgiarini LF, Rodrigues G, Tieppo J, Borghetti GS, Bassani VL *et al*. Quercetin decreases liver damage in mice with non-alcoholic steatohepatitis. *Basic & Clinical Pharmacology & Toxicology*. 2013; 112:385-391.
 43. Mariee DA, Abd-Allah MG, El-Beshbishy AH. Protective effect of dietary flavonoid quercetin against lipemic-oxidative hepatic injury in hypercholesterolemic rats. *Pharmaceutical Biology*. 2013; 50(8):1019-1025.
 44. Markesbery WR, Lovell MA. Damage to lipids, proteins, DNA, and RNA in mild cognitive impairment. *Archives of Neurology*. 2007; 64(7):954-956.
 45. Muhammad F, Ahsan M, Abdul W. Quercetin- A Mini Review. *Mod Concep Dev Agrono*, 2017, 1(2). DOI: 10.31031/MCDA.2018.01.000507
 46. Nijveldt RJ, van Nood E, van Hoorn DE, Boelens PG, van Norren K, Van LPA *et al*. Flavonoids: A review of probable mechanisms of action and potential applications. *American Journal of Clinical Nutrition*. 2001; 74(4):418-25.
 47. Oliveira CRD, Ceolin J, Oliveira RRD, Schemitt EG, Colares JR, Bauermann LDF *et al*. Effects of quercetin on polychlorinated biphenyls-induced liver injury in rats. *Nutricion Hospitalaria*. 2014; 29(5):1141-1148.
 48. Orayaga KT, Oluremi OIA, Adenkola AY. Effect of water soaking of sweet orange (*Citrus sinensis*) fruit peels on hematology, carcass yield and internal organs of finisher broiler chickens. *Journal of Animal Health and Production*. 2016; 4(3):65-71.
 49. Ouyang K, Xu M, Jiang Y, Wang W. Effects of alfalfa flavonoids on broiler performance, meat quality, and gene expression. *Canadian Journal of Animal Science*. 2016; 96:332-341.
 50. Ozgen S, Kilinc OK, Selamoglu Z. Antioxidant Activity of Quercetin: A Mechanistic Review. *Turkish Journal of Agriculture - Food Science and Technology*. 2016; 4(12):1134-1138.
 51. Panchal SK, Poudyal H, Brown L. Quercetin ameliorates cardiovascular, hepatic, and metabolic changes in diet-induced metabolic syndrome in rats. *Journal of Nutrition*. 2012; 142:1026-1032.
 52. Park YJ, Ko JW, Jeon S, Hye Kwon Y. Genistein alleviates neurodegeneration in ApoE^{-/-} mice fed a high-fat diet. *FASEB Journal*. 2013; 27(1):861-883.
 53. Porras D, Nistal E, Florez SM, Vaquero SP, Olcoz LJ, Jover R. Protective effect of quercetin on high-fat diet-induced non-alcoholic fatty liver disease in mice is mediated by modulating intestinal microbiota imbalance and related gut-liver axis activation. *Free Radical Biology and Medicine*. 2017; 102:188-202.
 54. Ramamneh DA. Effect of using Liquid onion on Broiler Physiology, Production and Behavior. *Bulletin of Environment, Pharmacology and Life Sciences*. 2017; 6(8):87-92.
 55. Rehman ZU, Che L, Ren S. Supplementation of vitamin E protects chickens from Newcastle disease virus mediated exacerbation of intestinal oxidative stress and tissue damage. *Cellular Physiology and Biochemistry*. 2018; 47(4):1655-1666.
 56. Rehman ZU, Meng C, Sun Y, Safdar A, Pasha RH, Munir M *et al*. Oxidative Stress in Poultry: Lessons from the Viral Infections. *Hindawi Oxidative Medicine and Cellular Longevity*, 2018; 14. <https://doi.org/10.1155/2018/5123147>
 57. Rice-Evans C, Miller N, Paganga G. Antioxidant properties of phenolic compounds. *Trends in Plant Science*. 1996; 2(4):152-159.
 58. Saeed M, Naveed M, Arain M, Arif M, Abd El-Hack ME, Alagawany M *et al*. Quercetin: Nutritional and beneficial effects in poultry. *World's Poultry Science Journal*. 2017; 70:23-28.
 59. Salah Eldin TA, Hamady GAA, Abdel-Moneim MA, Farroh KY, El-Reffaei WHM. Nutritional evaluation of Selenium-methionine nanocomposite as a novel dietary supplement for laying hens. *Journal of Animal Health and Production*. 2015; 3(3):64-72. <https://doi.org/10.14737/journal.jahp/2015/3.3.64.72>
 60. Scholz S, Williamson G. Interactions affecting the bioavailability of dietary polyphenols *in vivo*. *International Journal of Vitamin and Nutrition Research*. 2007; 77:224-235.
 61. Serafini M, Peluso I, Raguzzini A. Session 1: antioxidants and the immune system. Flavonoids as anti-inflammatory agents. *Proceedings of the Nutrition Society*. 2012; 69:273-278.
 62. Shizuma T. Relieving effect of quercetin on ischemia/reperfusion-induced liver damage in an animal model. *Journal of Nutrition, Health & Food Science*. 2014; 2(1):1-3.
 63. Sikder K, Das N, Kesh BS, Dey S. Quercetin and β -sitosterol prevent high fat diet induced dyslipidemia and hepatotoxicity in swine albino mice. *Indian Journal of experimental Biology*. 2014; 52:60-66.
 64. Sohaib M, Butt MS, Shabbir MA, Shahid M. Lipid stability, antioxidant potential and fatty acid composition of broilers breast meat as influenced by quercetin in combination with α -tocopherol enriched diets. *Lipids in Health and Disease*. 2015; 14:61-65.
 65. Starcevic KL, Krstulovic D, Brozic M, Mauric Z, Stojevic Z, Mikulec M *et al*. Production performance, meat composition and oxidative susceptibility in broiler chicken fed with different phenolic compounds. *Journal of the Science of Food and Agriculture*. 2015; 95:1172-1178.

66. Stewart LK, Soileau JL, Ribnicky D, Wang ZQ, Raskin I, Poulev A *et al.* Quercetin transiently increases energy expenditure but persistently decreases circulating markers of inflammation in C57BL/6J mice fed a high-fat diet. *Metabolism*. 2008; 57:S39-S46.
67. Surai PF, Fisinin VI. Vitagenes in poultry production: Part 3. Vitagene concept development. *World's Poultry Science Journal*. 2016; 72:793-804.
68. Surai PF. Polyphenol compounds in the chicken/animal diet: from the past to the future. *Journal of Animal Physiology and Animal Nutrition*. 2013; 98:19-31.
69. Suzuki T, Hara H. Role of flavonoids in intestinal tight junction regulation. *Journal of Nutritional Biochemistry*. 2011; 22:401-408
70. Vieira EK, Bona, Cangeri Di Naso FS, Porawski M, Tieppo J, Marroni NP. Quercetin treatment ameliorates systemic oxidative stress in cirrhotic rats. *International Scholarly Research Network*, 2011. Article ID 604071.
71. Viveros A, Chamorro S, Pizarro M, Arija I, Centeno C, Brenes A *et al.* Effects of dietary polyphenol-rich grape products on intestinal microflora and gut morphology in broiler chicks. *Poultry Science*. 2011; 90:566-578.
72. Wilms LC, Hollman PC, Boots AW, Kleinjans JC. Protection by quercetin and quercetin-rich fruit juice against induction of oxidative DNA damage and formation of BPDE-DNA adducts in human lymphocytes. *Mutation Research*. 2005; 582:155-162.
73. Yi L, Jin X, Chen CY, Fu YJ, Zhang T, Chang H *et al.* Chemical structures of 4-oxoflavonoids in relation to inhibition of oxidized low-density lipoprotein (LDL)-induced vascular endothelial dysfunction. *International Journal of Molecular Sciences*. 2011; 12:5471.
74. Zengpeng L, Kun X, Guang L, Dan L, Yuming G. Dietary genistein alleviates lipid metabolism disorder and inflammatory response in laying hens with fatty liver syndrome. *Frontiers in Physiology*. 2018; 9:1493.
75. Zou W, Liu W, Yang B, Wu L, Yang J, Zou T *et al.* Quercetin protects against perfluorooctanoic acid induced liver injury by attenuating oxidative stress and inflammatory response in mice. *International Immunopharmacol*. 2015; 28:129-135.
76. Zou Y, Wei HK, Xiang QH, Wang J, Zhou YF, Peng J *et al.* Protective effect of quercetin on pig intestinal integrity after transport stress is associated with regulation oxidative status and inflammation. *Journal of Veterinary Medical Science*. 2016; 78(9):1487-1494.