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# Multiple anthelmintic resistance in a village sheep flock in Mahendergarh district (Haryana)

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#### Abstract

Sixty sheep with egg per gram of more than or equal to 150 were divided into four groups i.e. S1, S2, S3 and S4 of 15 animals each to assess the prevalence of anthelmintic resistance in gastrointestinal nematodes against commonly used anthelmintics in Mahendergarh district (Haryana). Group S1, S2 and S3 were treated with fenbendazole @ 5 mg/kg b.wt. orally, morantel @ 10 mg/kg b.wt. orally and ivermectin @ 0.2 mg/kg b.wt. subcutaneously, respectively. Group S4 served as untreated control. Faecal egg count was ascertained on day of treatment (0 day) and 12th day post treatment (PT), from sheep of all groups and individual faecal egg counts were determined by the modified McMaster technique. Pooled faecal cultures were made to recover infective larvae on day 0 and 12 PT. Results revealed that fenbendazole reduced the faecal egg counts by 67.19% on 12th day PT with upper and lower confidence levels as 83.26% and 35.69%, respectively, morantel caused 92.11% reduction in faecal egg counts with upper and lower confidence levels as 96.77% and 80.74%, respectively and ivermectin caused 80.44% reduction in faecal egg counts with upper and lower confidence levels as 90.82% and 58.32%, respectively indicating moderate anthelmintic resistance for all three drugs. The post-treatment coproculture showed Haemonchus contortus and Strongyloides spp. larvae. Thus, the present study revealed presence of multiple anthelmintic resistance against fenbendazole, morantel and ivermectin in sheep of Mahendergarh district in Haryana.

Keywords: Anthelmintic resistance, fenbendazole, morantel, ivermectin, sheep

## Introduction

Sheep and goat rearing has been a major source of income especially to the marginal farmers of the country (Pathak and Pal, 2008) [12]. Gastrointestinal parasitic infection is a serious threat to small ruminant production systems. In fact, most of the economic losses caused by internal parasites are due to associated production losses in terms of decreased milk / wool production, poor hair coat or fleece growth, cost of prevention, cost of treatment and the death of infected animals (Gwaze et al., 2009) [7]. Parasitic gastroenteritis caused by many gastrointestinal (GI) nematodes like Haemonchus contortus, Trichostrongylus spp., Oesophagostomum spp., Nematodirus spp. and Strongyloides papillosus and is the major constraint in profitable animal husbandry practice. Among these GI nematodes, H. contortus, is most pathogenic, widely prevalent and important worm in sheep in India which is responsible for high mortality and morbidity (Yadav, 1997) [26]. The degree of parasitism or worm burden greatly depends on the management and hygienic conditions of the area (Singla, 1995) [23]. Control of GIT parasites is mainly achieved by the use of anthelmintic drugs and it will continue to remain, as there seems to be no other alternative for helminth control in small ruminants (Sanyal, 2004) [17]. The extensive use of anthelmintics for control of gastrointestinal nematodes has resulted in development of resistance to one or more of the widely used anthelmintics in many countries. (Maingi et al., 1998) [11]. In addition, multiple resistance to most of the anthelmintics against gastrointestinal nematodes have also been detected in many countries (Paraud et al., 2009) [14]. Thus, regular monitoring of status of anthelmintic resistance is required, atleast once in two years and it is as an integral part of worm control programme (Rialch et al., 2013) [15]. The present study was envisaged to detect the status of anthelmintic resistance to the most commonly used anthelmintic viz. fenbendazole, morantel and ivermectin against gastrointestinal nematodes of sheep in Mahendergarh district of Haryana.

# **Materials and Methods**

During August, 2018, a study was conducted at village Satnali, District Mahendergarh, Haryana to determine the efficacy of anthelmintics against gastrointestinal nematodes of sheep using faecal egg count reduction (FECR) test.

Sixty sheep naturally infected with gastrointestinal nematodes and having eggs per gram (EPG) of faeces > 150 counts prior to treatment were used. The selected animals had not been administered any anthelmintic during the previous two months. These animals were weighed, identified, their EPG estimated and divided into four groups i.e. S1, S2, S3 and S4 of 15 animals each. Group S1, S2 and S3 were treated with fenbendazole (FENAZOL-150® tablets, Pharmaceuticals Ltd., Animal Health Division, Mumbai) @ 5 mg/kg b.wt. orally, morantel (Banminth® Tab., Boehringer Ingelheim India Private Ltd. Mumbai) @ 10 mg/kg b.wt. orally and ivermectin (Trumectin®, Zydus Animal Health Limited, Ahmadabad) @ 0.2 mg/kg b.wt. subcutaneously, respectively. Group S4 served as untreated control.

Faecal egg count of each animal was ascertained on 0 day and 12<sup>th</sup> day post treatment (PT) by the modified McMaster technique to an accuracy of one egg counted representing 50 EPG. Pooled faecal cultures was incubated in petridish at 27 ± 2°C for 7 days and the infective larvae (L<sub>3</sub>) were recovered from each group on day 0 and 12<sup>th</sup> day PT. The infective larvae were identified as per criteria of (Keith, 1953)<sup>[9]</sup>. Faecal egg count reduction percentage and confidence intervals (95%) were determined following the method of the World Association for the Advancement of Veterinary Parasitology (WAAVP) using arithmetic mean egg counts (Coles *et al.*, 1992) <sup>[3]</sup>. The drug was considered fully effective when they reduced the egg counts by more than 95% and lower

confidence limits were higher than 90%. The drug was considered moderately resistant when they reduced the egg counts between 60% to 95% and considered severely resistant when the reduction in egg counts was below 60% along with lower confidence limits below 90%. All the recorded data was statistically analyzed by one way ANOVA test (SPSS software version 2.0).

## **Results**

Faecal egg counts (Mean  $\pm$  S.E.) on 0 and 12<sup>th</sup> day posttreatment (PT), percent reduction in faecal egg counts (FECR%), variance, upper and lower confidence limits (95%) of sheep naturally infected with gastrointestinal nematodes and treated with different anthelmintics at Satnali village, Mahendergarh are given in table 1. Results revealed that fenbendazole @ 5 mg/kg b. wt. (Group S1) reduced the faecal egg counts by 67.19% on 12<sup>th</sup> day PT with upper and lower confidence levels as 83.26% and 35.69%, respectively indicating moderate anthelmintic resistance. Further, morantel @ 10 mg/kg b. wt. (Group S2) caused 92.11% reduction in faecal egg counts with upper and lower confidence levels as 96.77% and 80.74%, respectively, again indicating moderate anthelmintic resistance. Ivermectin @ 0.2 mg/kg b. wt. (Group S3) caused 80.44% reduction in faecal egg counts with 95% upper and lower confidence levels as 90.82% and 58.32%, respectively, also indicating moderate anthemintic resistance.

Table 1: Response to various anthelmintics in sheep naturally infected with gastrointestinal nematodes at Satnali village, Mahendergarh

Group	Anthelmintic	Dose (mg/kg)	No. of sheep treated	Route of administration	Faecal egg counts on days (Mean ± S.E.)		Faecal egg counts reduction on day 12 post treatment		Confidence limits at 95%	
					0	12	%	Variance	Upper	Lower
S1	Fenbendazole	5	15	Oral	1866.67 <sup>a</sup> ± 137.55	$693.33^{b} \pm 217.66$	67.19	0.11	83.26	35.69
S2	Morantel	10	15	Oral	$1560.00^{a} \pm 98.46$	$166.67^{b} \pm 70.82$	92.11	0.19	96.77	80.74
S3	Ivermectin	0.2	15	S/C	$1660.00^{a} \pm 136.56$	$413.33^{\text{b}} \pm 147.32$	80.44	0.14	90.82	58.32
S4	Control		15		$2226.67^{a} \pm 238.54$	$2113.33^{a} \pm 205.37$	0			

Means with same superscripts in column are not significantly different (p<0.05)

The coproculture of pooled faecal cultures of infective third stage larvae in different groups and untreated control on day 0 and 12 (PT) are depicted in Table 2. A total of 100 infective larvae in each group (S1, S2, S3 and S4) were counted. The result showed different genera of GI nematodes of sheep with the predominance of *H. contortus* (83-86%) followed by *Strongyloides* sp. (10-12%), *Trichostrongylus* sp. (4-5%) and

only 1% *Oesophagostomum* spp. larvae in all the treated and untreated control groups on day 0. After 12 days of treatment, there was predominance of *H. contortus* larvae in fenbendazole and morantel treated animals while *Strongyloides* sp. larvae were predominant in ivermectin treated animals

Table 2: Anthelmintic effect on different genera of gastrointestinal nematodes of sheep at Satnali village, Mahendergarh

		Sheep Per cent larval composition on day		
Group	Species			
		0	12	
	Haemonchus spp.	86	90	
S1-Fenbendazole	Trichostrongylus spp.	4	0	
S1-relibelluazole	Oesophagostomum spp.	1	0	
	Strongyloides sp.	11	10	
	Haemonchus spp.	83	91	
S2- Morantel	Trichostrongylus spp.	5	0	
S2- Morantei	Oesophagostomum spp.	0	0	
	Strongyloides sp.	12	9	
	Haemonchus spp.	85	34	
S3- Ivermectin	Trichostrongylus spp.	4	0	
55- Ivermecum	Oesophagostomum spp.	1	0	
	Strongyloides sp.	10	66	
S4- Control	Haemonchus spp.	83	85	
54- Collifol	Trichostrongylus spp.	5	4	

Oesophagostomum spp.	1	1
Strongyloides sp.	11	10

### Discussion

The faecal egg counts on 0 and 12th day post-treatment (PT) and upper and lower confidence limits in sheep of Satnali village, Mahendergarh revealed presence of multiple anthelmintic resistance against fenbendazole, morantel and ivermectin. Fenbendazole belongs to benzimidazole class and its resistance to gastrointestinal nematodes in sheep had been reported by many workers (Vohra et al., 2013; Dolinska et al., 2014; Sharma et al., 2015; Singh et al., 2017) [4, 19, 20, 25]. The repeated administration of the compound predisposes the nematodes to develop resistance. History of use of anthelmintic and government supply in veterinary hospitals revealed that this is the most commonly used and supplied drug. The resistance of morantel against GI nematodes has also been reported by other workers (Singh and Yadav, 1997; Kumar and Singh, 2016) [10, 22]. History revealed that morantel was frequently used after fenbendazole depending upon availability and convenience of owner. The resistance of ivermectin has also been reported by many workers (Howell et al., 2008; Falzon et al., 2013; Pena-Espinoza et al., 2014; Sharma et al., 2015; Kumar and Singh, 2016) [5, 8, 10, 13, 19]. This drug has been used sometimes in the flock. Coles et al. (1999) [2] have reported the development of anthelmintic resistance even when two or three treatment are given annually. The multiple anthelmintic resistance against fenbendazole, morantel and ivermectin was also reported by other workers (Singh and Gupta, 2010; Butter et al., 2012, Garcia et al. 2016) [1, 6, 21].

The coproculture of pooled faecal cultures of infective third stage larvae in different groups on day 12 (PT) showed the predominance of *H. contortus* larvae in fenbendazole and morantel treated animals while *Strongyloides* sp. larvae was predominant in ivermectin treated animals. The presence of *H. contortus* and *Strongyloides* sp. larvae was also reported by workers (Sangwan *et al.*, 2006; Sarika, 2012; Sharma *et al.*, 2015) [16, 18, 19]. In the present study predominance of *Strongyloides* sp. larvae might be due to its reinfection in experimental animals because of its short prepatent period (Soulsby, 1965) [24] and unhygienic management conditions of the farm.

It should always be considered primarily to use an anthelmintic judiciously and the anthelmintic resistance may be estimated at least once in two years. The drugs which show partial resistance should be changed immediately and discontinued for some years so that the larval population resistant to the drug is diluted and the portion of susceptible larval population is increased in the sheep flocks. Due to frequent use of all classes of anthelmintics in sheep of this flock, resistance against all the classes of anthelmintics has developed.

# Conclusion

It may be concluded that the choice of anthelmintic in a flock should be based on the previous history of use of drug, frequency of use of drug and status of anthelmintic resistance. This is the first report of multiple anthelmintic resistance against all commonly used anthelmintics in a sheep flock from Mahendergarh in Haryana.

## References

1. Buttar BS, Rai HS, Singh NK, Haque JM, Rath SS.

- Emergence of anthelmintic resistance in an organized sheep farm in Punjab. Journal of Veterinary Parasitology. 2012; 26(1):69-71.
- Coles GC. Anthelmintic resistance and the control of worms. Journal of Medical Microbiology. 1999; 48:323-325
- Coles GC, Bauer C, Borgsteede FHM, Geerts S, Klei TR, Taylor MA *et al.* World Association for Advancement of Veterinary Parasitology (WAAVP) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. Veterinary Parasitology. 1992; 44:35-44.
- 4. Dolinska M, Ivanisinova O, Konigova A, Varady M. Anthelmintic resistance in sheep gastrointestinal nematodes in Slovakia detected by *in vitro* methods. BMC Veterinary Research. 2014; 10:233. doi: 10.1186/s12917-014-0233-4.
- 5. Falzon LC, Menzies PI, Vanleeuwen J, Jones bitton A, Shakya KP, Avula J *et al*. A survey of farm management practices and their associations with anthelmintic resistance in sheep flocks in Ontario, Canada. Small Ruminant Research. 2013; 114(1):41-45.
- 6. Garcia CMB, Sprenger LK, Ortiz EB, Molento MB. First report of multiple anthelmintic resistance in nematodes of sheep in Colombia. Anais da Academia Brasileira de Ciências. 2016; 88(1):397-402.
- 7. Gwaze FR, Chimonyo M, Dzama K. Prevalence and loads of gastrointestinal parasites of goats in the communal areas of the Eastern Cape Province of South Africa. Small Ruminant Research. 2009; 84:132-134.
- 8. Howell SB, Burke JM, Miller JE, Terrill TH, Valencia E, Williams MJ *et al.* Prevalence of anthelmintic resistance on sheep and goat farms in the south eastern United States. Journal of the American Veterinary Medical Association. 2008; 233(12):1913-1919.
- 9. Keith RK. The differentiation of infective larvae of some nematode parasites of cattle. Australian Journal of Zoology. 1953; 1:223-235.
- 10. Kumar S, Singh S. Detection of multiple anthelmintic resistance against gastrointestinal nematodes in sheep on central Sheep breeding farm, Hisar. Haryana Veterinarian. 2016; 55(2):210-213.
- 11. Maingi N, Bjom H, Dangolla A. The relationship between faecal egg` count reduction and the lethal dose 50% in the egg hatch assay and larval development assay. Veterinary Parasitology. 1998. 77:133-145.
- 12. Pathak AK, Pal S. Seasonal prevalence of gastrointestinal parasites in goats from Durg district of Chhattisgarh. Veterinary World. 2008; 5:136-137.
- Pena-Espinoza BM, Stig M, Thamsborgb, Demelerc J, Enemarka HL. Field efficacy of four anthelmintics and confirmation of drug resistant nematodes by controlled efficacy test and pyrosequencing on a sheep and goat farm in Denmark. Veterinary Parasitology. 2014; 206:208-215.
- 14. Paraud C, Kulo A, Pors I, Chartier C. Resistance of goat nematodes to multiple anthelmintics on a farm in France. Veterinary record. 2009; 164:563-564.
- 15. Rialch A, Vatsya S, Kumar RR. Detection of benzimidazole resistance in gastrointestinal nematodes of sheep and goats of sub-Himalyan region of northern India

- using different tests. Veterinary Parasitology. 2013; 198(3-4):312-318.
- 16. Sangwan AK, Sangwan N, Gupta SK. Anthelmintic resistance in trichostrongyloids of small holder sheep production system in some areas of Haryana. Haryana Veterinarian. 2006; 45(2):76-78.
- 17. Sanyal PK. Road leading to the future of anthelmintic therapy: No longer a bed of Red Roses. In: XV Annual Congress of IAAVP and National Symposium on Application of Molecular Biology in Parasitic Diseases for Rural Development, Pantnagar, 2004, 65-71.
- 18. Sarika. Studies on the status of anthelminthic resistance and efficacy of anthelmintic combinations against gastrointestinal nematodes in sheep. M.V.Sc. Thesis, Lala Lajpat Rai University of Veterinary and Animal Science, Hisar, 2012.
- 19. Sharma R, Singh S, Vohra S. Detection of anthelmintic resistance in gastrointestinal nematodes of sheep on government sheep breeding farm Hisar. Haryana Veterinarian. 2015; 54(2):147-149.
- 20. Singh R, Bal MS, Singla LD, Kaur P. Detection of anthelmintic resistance in sheep and goats against fenbendazole by faecal egg count reduction test. Journal of parasitic diseases. 2017; 41(2):463-466.
- 21. Singh S, Gupta SK. A survey of anthelmintic resistance in gastrointestinal nematodes in sheep of Haryana. Haryana Veterinarian. 2010; 49:25-28.
- 22. Singh S, Yadav CL. A survey of anthelmintic resistance by nematodes on three sheep and goat farm in Hisar (India). Veterinary Research Communications. 1997; 21:447-451.
- 23. Singla LD. A note on sub-clinical gastro-intestinal parasitism in sheep and goats in Ludhiana and Faridkot districts of Punjab. Indian Journal of Veterinary Medicine. 1995; 19:61-62.
- Soulsby EJL. Text Book of Veterinary Clinical Parasitology. Helminths. 1<sup>st</sup> ed. Blackwell Scientific Publications, Oxford, 1965, I.
- 25. Vohra S, Singh S, Poonia JS. Efficacy of individual and combinations of anthelmintic against gastrointestinal nematodes in goats. Haryana Veterinarian. 2013; 52:124-128.
- 26. Yadav CL. Premature ovine births caused by *Haemonchus contortus*. Indian Veterinary Journal. 1997; 74:983-984.