



ISSN (E): 2277- 7695  
ISSN (P): 2349-8242  
NAAS Rating: 5.03  
TPI 2019; 8(7): 141-146  
© 2019 TPI  
www.thepharmajournal.com  
Received: 10-05-2019  
Accepted: 12-06-2019

**Arti Arya**  
Department of Biotechnology,  
D.A.V. College, Sector 10,  
Chandigarh, India

## FTIR analysis: Biodegradation of complex hydrocarbons using *Coriolus versicolor*

**Arti Arya**

### Abstract

Industrial Pollution has gross adverse effects on biotic and abiotic components of environment. Organopollutants arising from petrochemical industries mainly constitute polyaromatic hydrocarbons, which are toxic and mutagenic. Control and treatment strategies include Old-school physicochemical treatments, which are cost intensive and may produce harmful byproducts. Offering high efficiency and low cost, bioremediation represents a significant approach for environmental clean up. These strategies involve microorganisms and utilize their metabolic capability to degrade organic compounds. In this study, degradation of complex hydrocarbons was studied using fungus, *Coriolus versicolor* in Shake flask experiment, using methylene blue as indicator dye. Results of dye decolorization assay and FTIR analysis indicated that *Coriolus versicolor* was capable of effectively degrading or removing the test compounds (Petroleum ether, DPA, PVP, Picric acid and Pthallic acid) used in the present study. The extent of decolorization recorded at wavelength 510nm, was taken to indicate fungal growth and FTIR analysis of culture supernatants showing altered peak profile with test compounds indicated possible breakdown to simpler structures.

**Keywords:** Fourier transform infra red (FTIR), poly aromatic hydrocarbons, *Coriolus versicolor*, industrial pollutants

### Introduction

Bioremediation is the use of living organisms, primarily microorganisms, fungi and plants, to degrade the environmental pollutants into less toxic forms. Living organisms through reactions that are a part of their normal metabolism transform complex compounds. Biodegradation of a compound is often a result of the actions of multiple organisms<sup>[1]</sup>. For bioremediation to be effective, conditions have to be manipulated to permit microbial growth and activity. Complex hydrocarbons like PAHs are most widespread organic pollutant owing to their physical and chemical properties and are associated with recalcitrance, bioaccumulation and toxicity. Phenolic compounds are ubiquitous in aqueous streams of industrial processes including: petroleum refining, coking and coal conversion, chemical plants, foundries and pulp and paper plants<sup>[2,3]</sup>. The presence of these compounds in drinking and irrigation water represents a health and environmental hazard. Although conventional methods of dephenolisation do exist, they are often expensive, deliver incomplete purification, can form hazardous byproducts and are often applicable to only to a limited concentration range.

White-rot fungus has the ability to degrade a wide variety of structurally diverse organic compounds, including a number of environmentally persistent organ pollutants<sup>[4]</sup>. The unique biodegradative abilities of fungus appear to be dependent upon its lignin-degrading system. White-rot fungi (*Phanerochaete chrysosporium*, *Pleurotus ostreatus*, *Coriolus versicolor*) have been demonstrated to oxidise PAH's rapidly with their extracellular ligninolytic enzyme systems and therefore have raised interest in the use of these organisms for bioremediation of contaminated soils<sup>[5]</sup>.

In this context, the present study reports degradation of Test aromatic hydrocarbons by the fungus *Coriolus versicolor* in shake flask experiment using dye decolorization and FTIR analysis as parameters.

### Material and Methods

**Revival of culture:** Lyophilized culture of the fungus (MTCC-138) was procured from Institute of Microbial Technology (IMTECH), Chandigarh. The fungus *Coriolus versicolor* was revived and maintained as a slant culture using standard protocol.

**Correspondence**  
**Arti Arya**  
Department of Biotechnology,  
D.A.V. College, Sector 10,  
Chandigarh, India

**Preparation of Broth:** Potato nutrient broth was used. Broth was inoculated with the fungus culture [6].

**Preparation of fungal mat by spread plate method:** Potato dextrose agar (PDA) media was used. Spreading was done using standard protocol [6].

**Morphological Confirmation of Fungal Strain:** A portion of mycelia mat from the plate was used for morphological conformation using Lactophenol cotton blue stain and observed under compound microscope at 10X and 40 X magnifications [7].

**Experimental Design for Biodegradation Studies (shake flask experiment):**

**Test Group:** In autoclaved nutrient broth (100ml), added 3-4 discs of fungal mat and respective amounts of organic compound (picric acid/petroleum ether-5mg/100 ml ;PVP/DPA/Pthallic acid-2mg/100ml), 1ml of methylene blue dye and 1ml of tween 80. Flasks were kept at 27 °C in orbital shaker at 120 r.p.m for 7 days. 2ml of media was taken out daily, centrifuged and O.D. of supernatant was measured at 510nm. Graphs were plotted between absorbance at  $\lambda$  max 510nm and time. Aliquots from the test groups were stored at -20°C and then used for FTIR analysis [8].

**Control group:** Parallel control flasks of media containing organic compounds incubated without fungus were taken.

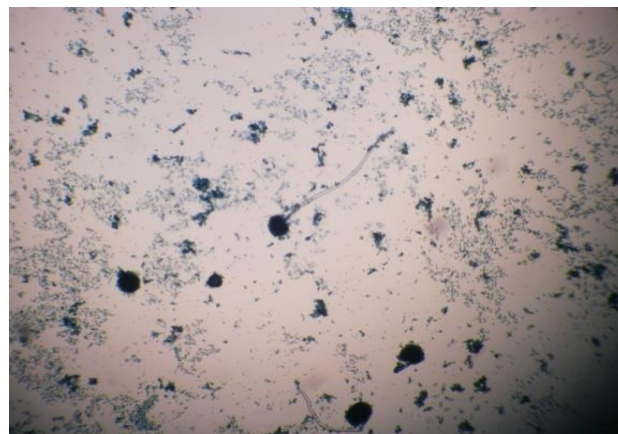
**FTIR analysis:** Aliquots of samples from shake flask experiments that were lyophilized to a fine powder were analyzed by FTIR spectroscopy (Perkin Elmer-spectrum RX-IFTIR) with range  $4000\text{cm}^{-1}$  to  $250\text{cm}^{-1}$  at Central Instrumentation Laboratory of Panjab University, Chandigarh [9].

**Statistical analysis:** All values were recorded and represented as mean  $\pm$  standard deviation (S.D) of 5-6 independent observations. For comparison of data for two groups (test and negative control) of biodegradation studies, student's t- test was performed.

**Result and Discussions**

**Morphological Confirmation of Fungal Strain after Revival of Culture**

*Coriolus versicolor* belongs to the class Basidiomycetes with the top surface of the growth shows typical concentric zones of different colors, 1–3 mm thick and leathery texture. Cap like growth with rust-brown or dark brown and sometimes blackish zones could be observed. The fungal hyphae were stained with lactophenol cotton blue. Hyphae were observed under 100X magnification giving blue appearance (Figure 1). Nucleus of hyphae was darkly stained with long tail. Similar kinds of studies by Wilson *et al*, 1993 [10] and some others [11] have reported the use of *Coriolus versicolor* in biodegradation and confirmed the morphology.

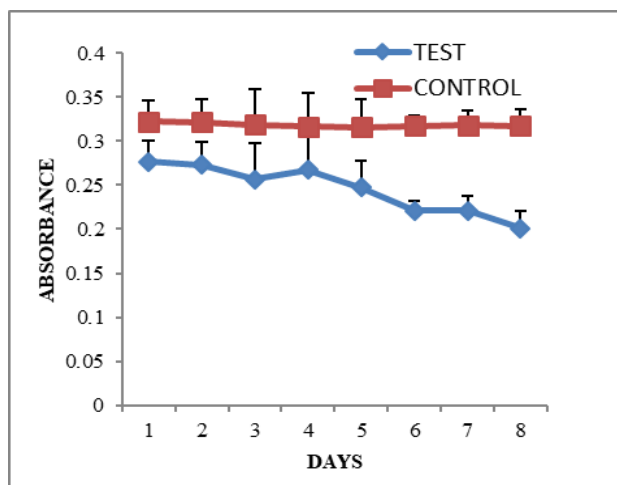


**Fig 1:** Fungal strain under 100X

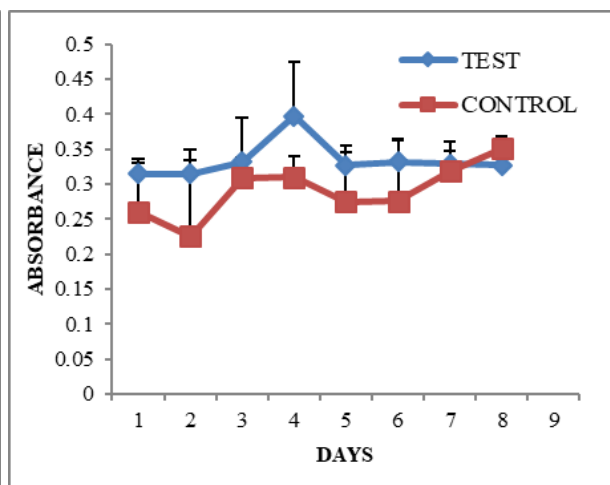
**Degradation of organic compounds**

**Dye Decolorization in presence of test compounds (PVP, DPA, Picric acid, Pthallic acid and Petroleum ether):**

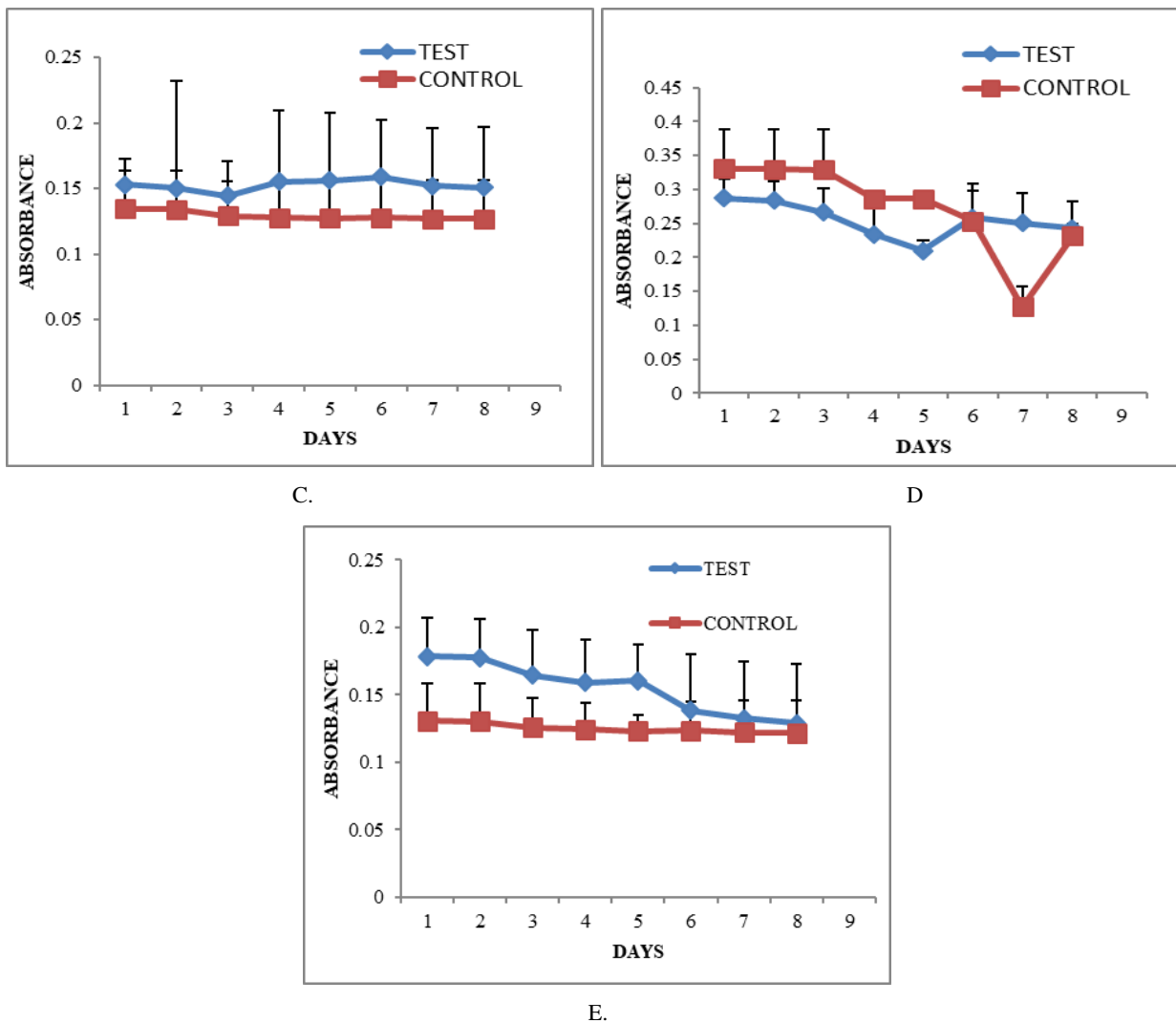
Degradation of test compounds was studied in a culture flask experiment by inoculating the flask with 3-4discs of fully-grown fungal mat (72hours). Methylene blue dye (indicator) and Tween20 (surfactant) was also added to the culture flask.



A.



B.



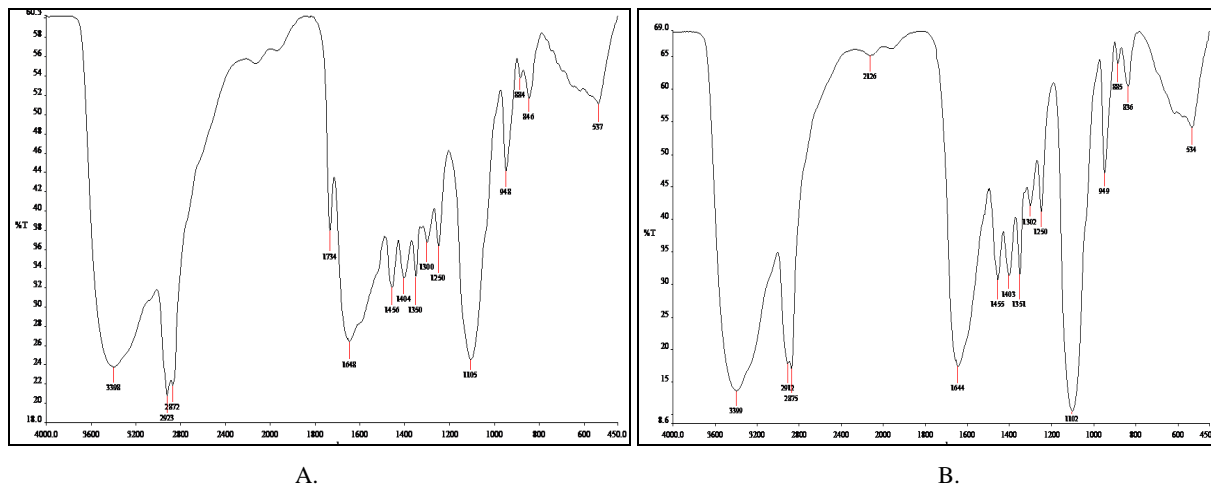
**Fig 2:** Characteristic absorbance vs. time graph showing degradation of (a) polyvinyl pyrrolidone (b) diphenylamine (c) picric acid (d) phthalic acid (e) petroleum ether indicated by decolorization of dye.

The medium turned turbid with time indicating fungal growth and decolorization of dye was observed. There was a significant visual difference in the decolorization of the dye but it was not significantly reflected in the absorbance when observed spectrophotometrically at the specific absorbance maxima (max510nm) (Figure 2). On 4<sup>th</sup> day, the absorbance value increased probably indicating that the fungus secreted some compounds (enzymes and metabolites). The absorbance value was found to decline on day 7 indicating that the fungus was allowed growth in the presence of organopollutants left over in culture media as also shown by decolorization of dye. Hence probably on day 7 the organic compound was reduced

to non toxic levels or was broken to simpler compounds further indicating that the levels was no longer inhibitory to the growth of fungi. Biodegradation of petroleum ether/PVP were reported in various studies [12, 13], where hyphae penetration was taken to indicate degradation.

**FTIR analysis**

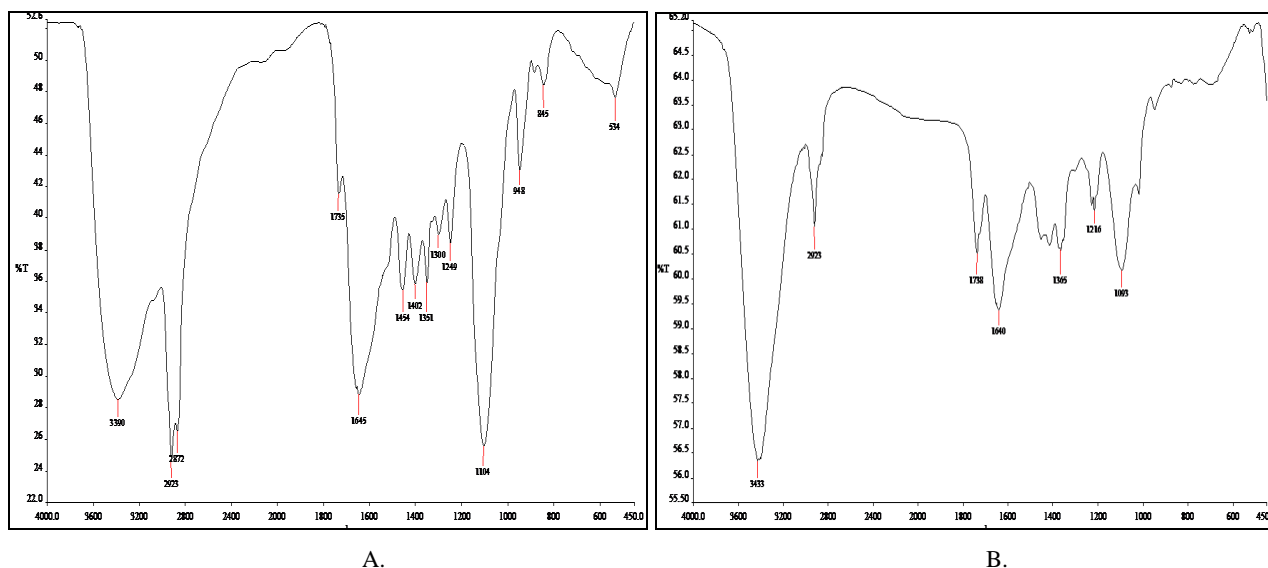
The FTIR studies were performed to analyze proposed transformation or degradation of various organopollutants in the presence of *Coriolus versicolor*.



**Fig 3:** FTIR spectrum of DPA (a) control (day1) and (b) test (day7)

FTIR spectrum of DPA control (day1) showed characteristic peaks at  $3408\text{cm}^{-1}$  caused by heterocyclic aromatic amines and at  $1644\text{cm}^{-1}$  because of secondary amine N-H bond. A C-N stretch was observed at  $1455\text{cm}^{-1}$  representing primary amine whereas test group (Day 1) revealed peaks at  $3398\text{cm}^{-1}$ ,  $2921\text{cm}^{-1}$ ,  $1633\text{cm}^{-1}$  and  $1101\text{cm}^{-1}$ . Control group i.e. without fungus exhibited low % transmittance (high absorbance) values for some of the peaks indicative of compound (Figure 3). The % transmittance increased i.e. decreased absorbance was observed in test group (Day 1) where fungus was incorporated indicating physical removal of compound by fungal mycelia. DPA degradation studies analyzed by FTIR have been reported by Mecozzi, *et al* 2009

[9]. FTIR of un-degraded PVP control (Day 1) revealed prominent peak at  $3390\text{cm}^{-1}$  representing N-H stretch. While test group (Day 7) revealed peaks at  $3343\text{cm}^{-1}$ ,  $2493\text{cm}^{-1}$ ,  $1738\text{cm}^{-1}$ ,  $1640\text{cm}^{-1}$ ,  $1365\text{cm}^{-1}$ ,  $1216\text{cm}^{-1}$ ,  $1093\text{cm}^{-1}$  (Figure 4). Test group (Day 7) revealed peak at  $3389\text{cm}^{-1}$  which may be caused by normal polymeric -OH stretch. At  $2872\text{cm}^{-1}$  and  $2921\text{cm}^{-1}$  methylene C-H with symmetrical and asymmetrical stretch was observed. In test group (Day 7) C-O stretch was observed at  $1640\text{cm}^{-1}$ . Bhardwaj and Khan (2004) reported FTIR analysis of Polyvinyl chloride in which it was analyzed to be not broken down into simpler compounds [13].



**Fig 4:** FTIR spectrum of PVP (a) test (day1) and (b) test (day7)

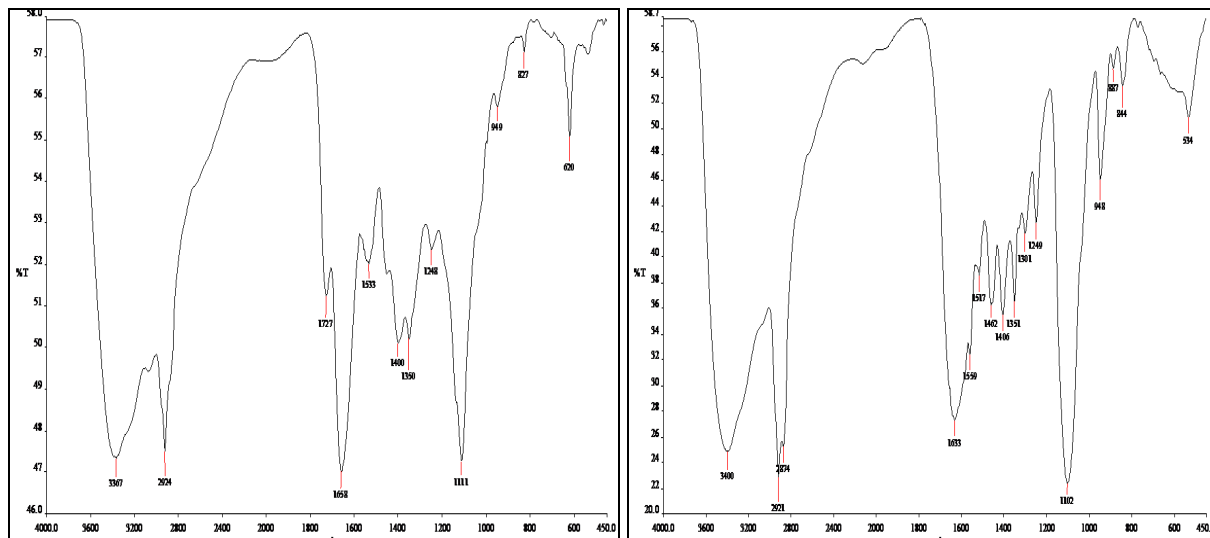


Fig 5: FTIR spectrum of Picric acid (a) control (day1) and (b) test (day7)

FTIR of undegraded Picric acid control (Day1) revealed prominent peak at  $3367\text{ cm}^{-1}$  representing primary amines. N-H stretch with strong appearance. A -CH stretch (asymmetric/symmetric) was observed at  $\sim 2830\text{ cm}^{-1}$  and  $2305\text{ cm}^{-1}$  aromatic hydrocarbon-H in plane ring stretch was observed (Figure 5). The test group (Day1) revealed

prominent peaks at  $3408\text{ cm}^{-1}$ ,  $2923\text{ cm}^{-1}$ ,  $1732\text{ cm}^{-1}$ ,  $1660\text{ cm}^{-1}$  and  $1535\text{ cm}^{-1}$  with low % transmittance i.e. high absorbance compared to peaks of Day 7 test group. The absorption at Day 7 are found to be  $\sim 1480\text{ cm}^{-1}$  and  $1180\text{ cm}^{-1}$  which are due to aromatic C=C absorption stretching vibrations compared with Day 1 having values  $\sim 1580\text{ cm}^{-1}$  and  $1500\text{ cm}^{-1}$ .

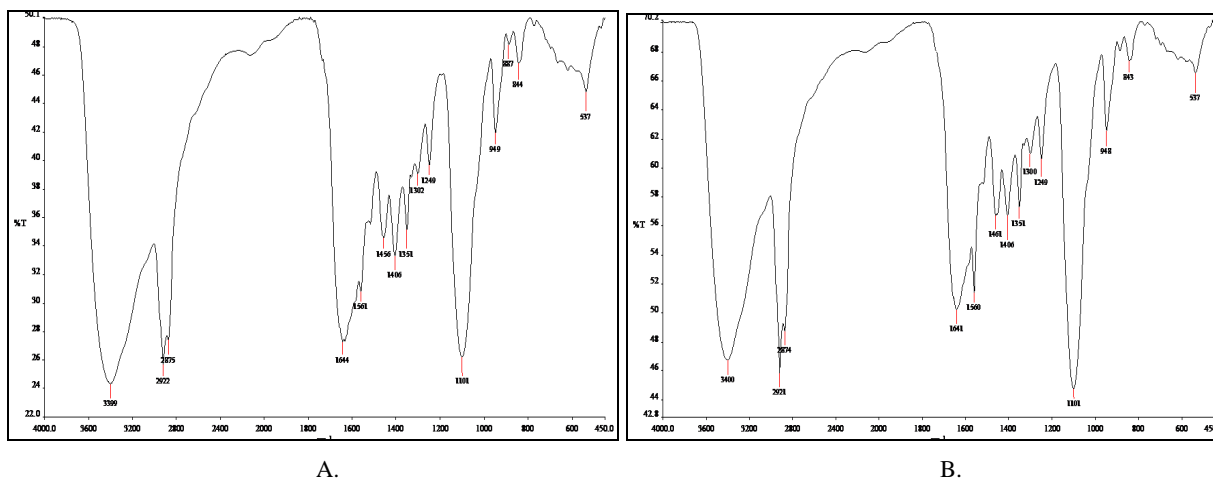


Fig 6: FTIR spectrum of phthalic acid (a) control (day1) and (b) test (day7)

FTIR spectrum of Phthalic acid as present in culture media at the start of degradation experiment showed prominent peaks at  $3434\text{ cm}^{-1}$ ,  $2926\text{ cm}^{-1}$ ,  $2100\text{ cm}^{-1}$ ,  $1642\text{ cm}^{-1}$ ,  $1101\text{ cm}^{-1}$  in case of Test group (Day1) while Test group (Day7) showed characteristic peaks at  $3400\text{ cm}^{-1}$ ,  $2921\text{ cm}^{-1}$ ,  $1641\text{ cm}^{-1}$ ,  $1101\text{ cm}^{-1}$ . Test group (Day 1) showed a peak at  $1457\text{ cm}^{-1}$

representing C-C stretch. A bending vibration of Carboxylic acid was observed at  $1300\text{ cm}^{-1}$  on Day 7 of Test group (Figure 6). Bending mode of phenolic group was observed from  $1304\text{ cm}^{-1}$  -  $1352\text{ cm}^{-1}$  in case of Test group (Day1). Similar studies were also reported by Bhat *et al*(2011) who studied the analysis at different concentrations and pH [14].

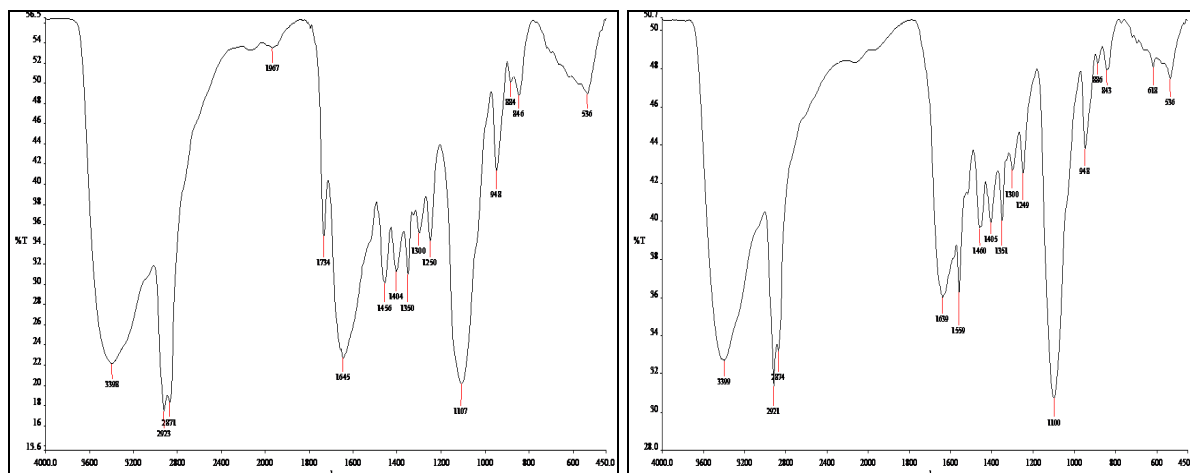


Fig 7: FTIR spectrum of petroleum ether (a) control (day1) and (b) test (day7)

FTIR of Petroleum ether control (Day1) revealed prominent peak at  $2923\text{ cm}^{-1}$  representing primary amine, N-H with strong appearance. A C-H stretch (symmetric/ asymmetric) was observed at  $2871\text{ cm}^{-1}$  and  $1645\text{ cm}^{-1}$  aromatic hydrocarbon, C-H in a plane ring stretch was observed (Figure 7). The test group (Day1) revealed prominent peaks at  $3399\text{ cm}^{-1}$ ,  $2921\text{ cm}^{-1}$ ,  $1645\text{ cm}^{-1}$ ,  $1104\text{ cm}^{-1}$  with high % transmittance i.e. low absorbance as compared to peaks of test group (Day7) which appeared at low % Transmittance. Similar studies have also been reported by Olfat & Karimi (2005) in which degradation of crude oil was observed in presence of *Coriolus versicolor* [12]. The vibrational spectrum of a molecule is considered a unique physical property and is characteristics of the molecules as such; the IR spectrum can be used as a fingerprint for identification, by comparison of spectrum of unknown with previously recorded reference spectra [15].

The present study indicated that fungus *Coriolus versicolor* has the potential to degrade organic compounds as indicated by dye decolorization using spectrophotometric analysis. The fungal growth turned the media turbid with time and decolorization of dye was observed. Decrease in absorbance value with the growth of fungi indicated reduction of organic compound to non-toxic levels or break down to simpler compounds. Supporting evidence for biodegradation of compounds came from FTIR analysis of various groups, where a change in the peak intensities and patterns was reflective of bioconversion/removal. Moreover, FTIR spectroscopy can be a very useful tool in performing preliminary tests in order to predict remediation performance to select an appropriate approach for clean-up technologies.

## References

1. Aparna C, Saritha P, Himabindu V, Alok B, Anjaneyulu Y. Evaluation of bioremediation effectiveness on sediments contaminated with industrial wastes. *International Journal of Environmental Sciences*. 2010; 1(4):607-620.
2. Van Schie PM, Young LY. Isolation and characterization of phenol- degrading denitrifying bacteria. *Applied and Environmental Microbiology*. 1998; 64(7):2432-2438.
3. Aitken MD, Massey IJ, Chen T, Heck PE. Characterization of reaction products from the enzyme catalyzed oxidation of phenolic pollutants. *Water Research*. 1994; 28(9):1879-1889.
4. John A, Bumpus SDA. Biodegradation of environmental pollutants by the white rot fungus *Phanerochaete chrysosporium*. Involvement of the lignin degrading system, 2005.
5. Canet R, Lopez-Real JM and Beck AJ. Overview of Polycyclic Aromatic Hydrocarbon Biodegradation by White-rot Fungi. *Land Contamination & Reclamation* 1999; 7(3):191-197.
6. Ruhul SM, Sarker NC, Kamal QH. Optimization of in-vitro culture conditions for mycelia growth of turkey tail mushroom. *National mushroom development and extension centre, Dhaka, Bangladesh*. 2008; 2920:63-71.
7. Harrigan MG, McCance ME. *Laboratory methods in food and dairy microbiology*. London: academic press. 1976; 42:33-200.
8. Bending GD, Friloux M, Walker A. Degradation of contrasting pesticides by white rot fungi and its relationship with lignolytic potential. *Horticulture research international*. Wellesbourne, 2002.
9. Mecozzi M, Moscato F, Pietroletti M, Quarto F, Oteri F, Cicero AM. Applications of FTIR spectroscopy in environmental studies supported by two dimensional correlation analyses. *Global NEST Journal*. 2009; 11(4):593-600.
10. Wilson SC, Jones KC. Bioremediation of soil contaminated with polynuclear aromatic hydrocarbon (PAHs): A Review. *Environmental Pollution* 1993; 81:229-249.
11. Cohen MJ, Gabriele PD, Wilson BW, Bean RM, Franz JA, Thomas BL *et al*. Submitted to *Energy and Fuels* 1982, 80-84.
12. Olfat AM, Karmi AN. Degradation of crude oil and beech wood by *Coriolus versicolor*. *Pakistan journal of biological sciences*. 2005; 8(10):1453-1456.
13. Bhardwaj S, Khan AK. Decolorization of pulp and paper mill effluent using polyvinyl chloride. *Indian Journal of Chemical Technology*. 2004; 11:607-611.
14. Bhat MM, Shankar S, Shikha, Yunus M, Shukla RN. Remediation of Hydrocarbon Contaminated Soil through Microbial Degradation- FTIR based prediction. *Advances in Applied Science Research*. 2011; 2(2):321-326.
15. Dent G, Charmer JM. *Industrial analysis with vibrational spectroscopy*, Royal Society of Chemistry, Cambridge, 1997.