



ISSN (E): 2277- 7695
 ISSN (P): 2349-8242
 NAAS Rating: 5.03
 TPI 2019; 8(11): 278-280
 © 2019 TPI
 www.thepharmajournal.com
 Received: 19-09-2019
 Accepted: 21-10-2019

Mylaram Jeevana Latha
 Assistant Professor & Head,
 Department of Veterinary
 Pathology, College of Veterinary
 Science, Mamnoon, Warangal
 District, PVNRTVU, Telangana,
 India

M Lakshman
 Associate Professor, I/C RUSKA
 Labs, Department of Veterinary
 Pathology, College of Veterinary
 Science, PVNRTVU,
 Rajendranagar, Hyderabad,
 Telangana, India

A Anand Kumar
 Associate Dean, College of
 Veterinary Science, SVVU,
 Proddutur, YSR Kadapa,
 Andhra Pradesh, India

A Gopal Reddy
 Professor and Head, Department
 of Pharmacology and
 Toxicology, College of Veterinary
 Science, PVNRTVU,
 Rajendranagar, Hyderabad,
 Telangana, India

D Pramod Kumar
 Professor and Head, Department
 of Anatomy and Histology,
 College of Veterinary Science,
 PVNRTVU, Rajendranagar,
 Hyderabad, Telangana, India

Corresponding Author:
Mylaram Jeevana Latha
 Assistant Professor & Head,
 Department of Veterinary
 Pathology, College of Veterinary
 Science, Mamnoon, Warangal
 District, PVNRTVU, Telangana,
 India

A radiographic study of embryos on hexavalent chromium toxicity in female wistar rats

Mylaram Jeevana Latha, M Lakshman, A Anand Kumar, A Gopal Reddy and D Pramod Kumar

Abstract

Hexavalent chromium salts have been recognized as occupational health hazard for more than 160 years. It is a potent toxic agent. It has been found to be carcinogenic in human and animals. Hexavalent chromium is generally considered 1000 times more toxic than trivalent chromium. The experiment was conducted to study the embryonal toxicity in female wistar rats by inducing hexavalent chromium at the rate of 500 parts per million in the form of potassium dichromate in drinking water orally for three months and also to pregnant animals, with the protective and ameliorating agents like vitamin C and *emblica officinalis* (*E. officinalis*) respectively. Foetuses were collected by sacrificing pregnant rats from all six groups for the purpose of radiographic study. The fetuses from all groups were subjected to radiographic study. The embryos from the toxin group revealed mild subdermal haemorrhages over abdomen region, ill developed fetuses when compared to the morphological features of control, protective and ameliorative group embryos.

Keywords: Chromium toxicity, embryos, amelioration, radiographic morphological study

Introduction

Chromium is a naturally occurring element found in animals, plants, rocks, soil, volcanic dust and gases (Nejla Soudani *et al.*, 2010) [1]. Hexavalent chromium is an important reproductive and developmental toxicant as per office of environmental Health Hazard Assessment (OEHHA) and the Development and Reproductive toxicant Identification Committee (DARTIC) (Toxicology profile for chromium, United States Environmental Protection Agency, USEPA, 2001) [2]. The effect of hexavalent chromium on multiple pathways could be mitigated by Vitamin C (Sakhila *et al.*, 2011) [3]. Plant parts of *emblica officinalis* showed antidiabetic, hypolipidemic, antibacterial, antioxidant, hepatoprotective and gastro protective properties (Krishna veni and Mrunalini, 2010) [4].

Materials and Methods

All rats were handled in accordance with the guidelines of Institutional Animal Ethics Committee (IAEC) (No 1/4/14, date 27.11.2014). The dams were sacrificed and the fetuses were collected to record the lesions. X-ray Radiographic studies of fetuses were carried out at Super Speciality Hospital, Narayanaguda, Hyderabad as per the standard protocol.

Experimental design

Group	No of Rats	Type of treatment / diet
I (Control)	20	Basal diet
II (Toxin control)	20	Basal diet + Potassium dichromate 500 parts per million (ppm) in drinking water orally for 3 months
III (Vit C control)	20	Basal diet + vitamin C @ 100 milligram (mg)/kg body weight (kg.b.wt) orally for 3 months.
IV (<i>Emblica officinalis</i> control)	20	<i>Emblica officinalis</i> powder given @ 2 % in feed for 3 Months
V (Chromium VI + Vitamin C control)	20	Basal diet + Potassium dichromate 500 ppm in drinking water orally for 3 months + vitamin C @ 100 mg/kg b.wt orally for 3 months.
VI (Chromium VI + <i>Emblica officinalis</i>)	20	Basal diet +potassium dichromate 500ppm in drinking water orally for 3 months+ <i>Emblica officinalis</i> powder given @ 2% in feed for 3 months.

Results

The radiographic morphological study of fetuses/embryos through x-ray studies in the toxin group showed significantly mild sub dermal haemorrhages in the form of patches over abdomen region (Fig: 1). The fetuses were ill developed and very small in size, which is very prominent feature (Fig: 2) observed in treated embryos when compared to morphological features of other control, protective and ameliorative group embryos (III, IV, V and VI).

The toxin group fetuses were also showed abnormality in skeletal development like incomplete ossification in nasal, cranium, abdominal or caudal bones and absence of ossification in the sacral vertebrae which clearly depicted through in x-ray radiographs (Fig: 3) in comparison with the skeletal development of other control, protective and ameliorative group of embryos (Fig: 4). The morphological features and skeletal development is apparently normal in protective and ameliorative groups.

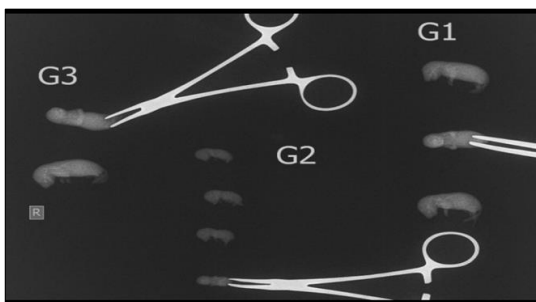


Fig 1: Note the morphological features of the fetuses showing defects in the abdomen and thigh region in the toxin group (group II) rats

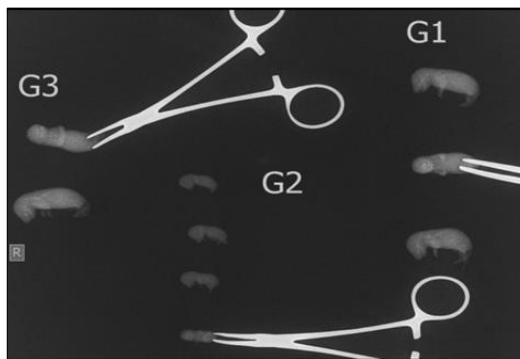


Fig 2: Note the apparently normal morphological features of the fetuses in the control group (Group I, III) rats, abnormal and very small sized fetuses of group II through radiographic studies (30 days)

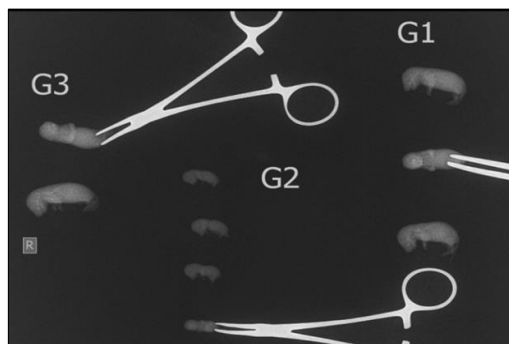


Fig 3: Note the abnormality in skeletal development of fetuses showing incomplete ossification in nasal, cranium, abdominal or caudal bones in the toxin group (group II) rats through radiographic studies (30 days)

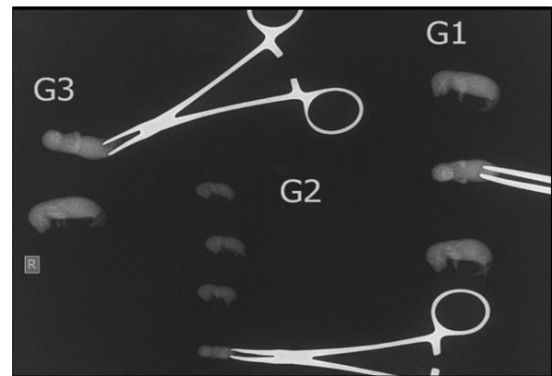


Fig 4: Note the very small size of the fetuses and a characteristic and pronounced ill development in toxin group compared to control group (I, III) of rats (30 days)

Discussion

The defects in the embryos may be due to impaired development of embryos during gestation (Kanojia *et al.*, 1996) [5], interrupted uterine blood flow in hexavalent chromium treated rats (New and Coppola, 1977 and Garis, 1989) [6, 7], foeto toxic effects with significant reduction of implantation rate, number of fetuses, an increase in the resorption number, pre implantation and post implantation (Junaid *et al.*, 1996a and Kanojia *et al.*, 1996) [8, 5] and decrease in the crown rump due to embryo and foetotoxic effects (Neila Morouani *et al.*, 2010) [9].

These adverse effects were comparatively minimum by supplementing vitamin C and *emblica officinalis* because of its ameliorative role. Vitamin C acts as a biological antioxidant by donating an electron to free radical, thereby interrupting the radical chain reaction in biological membranes and protects from the deleterious effects like fetal abnormalities in rats (Kesinger and Stevens, 2009) [10]. Absence of ossification in the sacral vertebrae found through x-rays analysis and the same was reported by Neila Morouani *et al.*, 2010 [9] during the organogenesis period.

References

1. Nejla Soudani, Hanen Bonaziz, Mediha Sefi, Yassine Chtoneou, Tahia Boudawara, Najiba Zeghal. Toxic effects of chromium (VI) by Maternal Ingestion of Liver function of Female Rats and their sucking pups, 2010.
2. US EPA. Toxicology profile for chromium. Inc. [Last cited in 2001], 2001. Available from: <http://www.epa.gov>.
3. Sakhila K Banu, Jone A Stanley, Jettoon Lee, Sam D Stephen, Joe A, Arosh JA, *et al.* Hexavalent chromium induced apoptosis of granulose cells involves selective sub cellular translocation of BCL-2 members, ERK1/2 and P53, Toxicol Appl Pharmacol. 2011; 251(3):253-266.
4. Krishnaveni M, Mirunalini S. Therapeutic potential of *Phyllanthus emblica* (amla): The ayurvedic wonder. J Basic Clin Physiol Pharmacol. 2010; 21:93-105.
5. Kanojia RK, Junaid M, Murthy RC. Chromium induced teratogenicity in female rat. Toxicol Lett. 1996; 89(3):207-13.
6. New D, Coppola P. Development of a placental blood circulation in rat embryos *in vitro*. J. Embryol. Exp. Morph. 1977; 37:227-35.
7. Garris D. Intrauterine growth of the guinea pig fetal placental unit throughout pregnancy: regulation by utero-placental blood flow. Teratology. 1989; 29:93-9.

8. Junaid M, Murthy RC, Saxena DK. Embryotoxicity of orally administered chromium in mice: exposure during the period of organogenesis. *Toxicol. Lett.* 1996a; 84:143-8.
9. Neila Marouani, Olfa Tebourbi, Moncef Mokni, Mohammed Tahar Yacoubi, Mohsen Sakly, Moncef Benkhalifa, *et al.* Embryotoxicity and fetotoxicity following intraperitoneal administrations of hexavalent chromium to pregnant rats. *Zygote*, 2010, 229-235.
10. Kesinger N, Stevens J. Covalent interaction of ascorbic acid with natural products phytochemistry. 2009; 70(17/18):1930-1939.