



ISSN (E): 2277- 7695

ISSN (P): 2349-8242

NAAS Rating: 5.03

TPI 2018; 7(6): 376-379

© 2018 TPI

www.thepharmajournal.com

Received: 15-04-2018

Accepted: 18-05-2018

**Anjali Sharma**

Department of Biotechnology  
and Bioinformatics, DAV  
College, Sector-10, Chandigarh,  
India

**Aparna Thakur**

Department of Biotechnology  
and Bioinformatics, DAV  
College, Sector-10, Chandigarh,  
India

**Rupinder Virk**

Department of Biotechnology  
and Bioinformatics, DAV  
College, Sector-10, Chandigarh,  
India

**Rupinder kaur**

Department of Biotechnology  
and Bioinformatics, DAV  
College, Sector-10, Chandigarh,  
India

**Correspondence**

**Rupinder kaur**

Department of Biotechnology  
and Bioinformatics, DAV  
College, Sector-10, Chandigarh,  
India

## Role of tristetrapolin protein in muscle regeneration: an *Insilico* analysis

**Anjali Sharma, Aparna Thakur, Rupinder Virk, Meenu Khurana and  
Rupinder kaur**

### Abstract

Skeletal muscle has a highly complex structure and shaving a numerous roles animal body. A set of inhouse satellite has an interal role in the regeneration pathway forming new myofibers post tissue damage due to injury or a degeneration. On getting a signal of from necrosis in muscle fibres, satellite cells get activated, have an asymmetric division to self-generate, and form activated myoblasts capable of proliferating and reaching the site of damage, and bind to the already present myofibers or create newer ones. It is known that Tristetraprolin, an RNA-binding protein involved in the regulation of cytokine expression has a function of muscle regeneration. Tristetraprolin is present in humans, mice and rats. It is encoded by the Zinc Ring Finger Protein 36 gene. In this study, we have elucidated the role of Tristetraprolin in muscle regeneration by using *insilico* approaches.

**Keywords:** Muscle, regeneration, transcripts, RNA-binding protein

### 1. Introduction

Messenger RNA decay is a important way to regulate the expression of numerous inflammation- and cancer-related genes [1]. These transcripts are tagged for quick breakdown via AU-rich element (ARE) motifs present in the mRNA 3'untranslated region (3'UTR) [2]. Tristetraprolin (TTP), also termed as zinc finger protein 36 homolog ZFP36, is present in humans, mice and rats and encoded by the ZFP36 gene [3].

TTP, an RNA-binding protein performs a pivotal function of regulating the expression of ARE-encompassing mRNAs. Owing to its potential of attaching to AREs it tags the corresponding mRNA for quick breakdown, TTP can decrease the expression of numerous important genes generally overexpressed during inflammation and cancer. Mice deficit in TTP have a complicated syndrome of inflammatory diseases [4]. Regulation of TTP is at many strata via cellular signaling pathways to regulate transcription, mRNA turnover, phosphorylation status, cellular localization, binding to other proteins, and proteosomal degradation, all affecting TTP's potential to enhance ARE-promoted mRNA desaggregation and decay-independent role of TTP.

Regeneration of damaged muscle is a complicated pathway encompassing numerous cell types and the mediation of a many growth factors, cytokines, chemokines, extracellular matrix components and signaling molecules [5]. Skeletal muscle development, repair and function rely on highly regulated expression of many genes. Myogenic precursor cells called satellite cells exist in adult skeletal muscle. Satellite cells refer to adult myogenic stem cells that regenerate damaged muscle. The capability for muscle regeneration needs effective satellite cell expansion post injury, their differentiation to form myoblasts that can reform damaged fibres and their self-renewal to regenerate the muscle stem cell pool for further cycles of injury and recovery. Dormant satellite cells are activated via signals coming from damaged muscle. Sachidanandan *et al.* have shown the quick activation of two genes post to muscle injury; these transcripts encode LPS-inducible CXC chemokine (LIX), a neutrophil chemoattractant, and Tristetraprolin (TTP), an RNA-binding protein involved in the regulation of cytokine expression [5]. LIX and TTP mRNAs have a short life span and are quickly upregulated in reply to muscle damage *in vivo* [5]. Along with generating new cells for repair, induced satellite cells might be a reservoir of signaling molecules implicated in tissue remodeling during regeneration [5]. Tristetraprolin (TTP) binds RNAs that has AU-rich parts and binds enzymes that degrade RNA. The activity of TTP is controlled by phosphorylation. MK2 phosphorylates TTP that further binds 14-3-3.

The binding to 14-3-3 restricts phosphorylated TTP from having access to stress granules and stabilizes mRNA flanked by phosphorylated TTP [6].

This studies suggest that TTP is playing an important role in muscle regeneration. In this paper *insilico* approaches have been used to study the interactions of TTP with other proteins through which it could be playing a role in muscle regeneration.

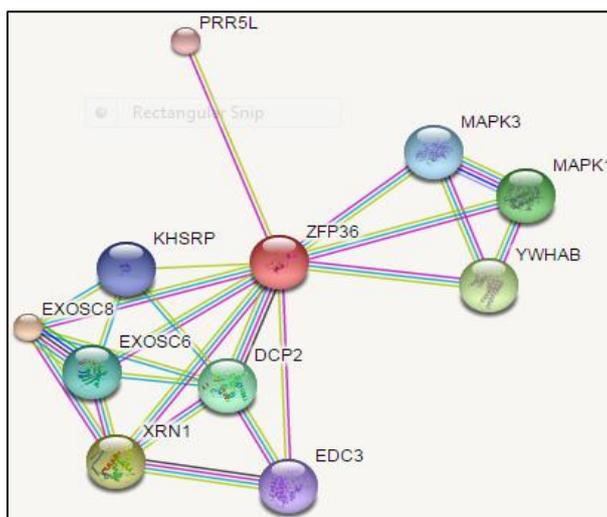
**2. Material and Methods**

**String** – The interactions network of the TTP protein with other protein was found out using the STRING database (<http://string-db.org>) [7]. The sequence used as a query was the human TTP protein Accession no. AAA61240 retrieved from NCBI database. (<http://www.ncbi.nlm.nih.gov/protein>) [8].

**Bio grid** –The interactions of TTP protein were also studied using BIOGRID database (<https://thebiogrid.org/>) [9].

**3. Results and Discussion**

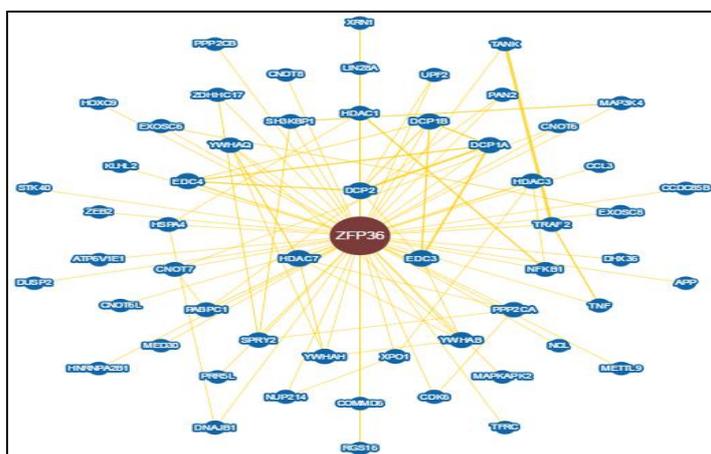
TTP is an RNA-binding protein that plays a role in regulating the expression of ARE-containing mRNAs. It has the ability to bind AREs and target the bound mRNA for rapid degradation. It has been cited in literature that TTP expression increases after muscle injury [5]. In the present study we have used *insilico* approaches to find the role of TTP protein in muscle regeneration after injury. The results of the protein interaction databases STRING (Fig 1 and Table 1) and BIOGRID (Fig 2 and Table 2) showed that TTP interacts with various proteins like chemokines, proteins involved in cytoskeleton organization, phosphorylation etc.



**Fig 1:** Showing the protein interactions of Human TTP protein using the STRING database

**Table 1:** TTP interacting proteins using string database

S. No.	TTP Interacting Protein	Functions
1	EXOSX8	3'-5' exoribonuclease activity and participates in degrading events. It help in the elimination of mRNA with processing defects
2	EXOSX6	Play role in replication dependent histone mRNA degradation, major 5'-3' exoribonuclease
3	EDC3	Enhance mRNA decapping
4	KHSRP	Degrade unstable mRNA by recruiting degrading machinery
5	PRR5L	Modulate phosphorylation of PKCA and regulate actin cyto-skeleton organization
6	MAPK3	Extracellular signal regulated kinase important role in MAPK/ERK cascade this signaling pathway mediate cell growth and differentiation through regulating transcription
7	YWHAB	Adaptor protein implicated in the regulation of specialized signaling pathway
8	RGS16	Regulator of G protein signaling family. Inhibition of G protein subunit by GTP ase.



**Fig 2:** Showing the protein interactions of Human TTP protein using the BIOGRID database

**Table 2:** TTP interacting proteins using bio grid

S. No	TTP Interacting Proteins	Functions
1	APP	Activate a widespread caspases dependent self destruction program
2	ATP6V1E1	Hydrogen exchange ATPase helps in phosphorylation
3	CCDC85B	Hydrogen exchange ATPase helps in phosphorylation
4	CCl3	It is a chemokine which increases rapidly during muscle injury and returns to control levels during muscle regeneration by stimulating myoblast proliferation
5	CNOT6L	involved in deadenylation dependent degradation of CDKMIB mRNA mediates peroliferation and cell survival and prevent cellular senescence
6	CNOT6	Function as poly A nuclease
7	CNOT7,CNOT8	3'-5' exoribonuclease activity forms complex with CCR4-NOT for bulk mRNA decay
8	COMMMD6	Negative regulation of NF kappa B transcription factor activity
9	DCP1A,DCP1B	Main activation of DCP2
10	DCP2	Forms complex with DCP main decoupling enzyme
11	DHX36	RNA helicases associated with AU rich element
12	DNAJB1	Heat shock protein
13	DUSP2	Dual phosphatase, terminate MAPK activity and helps in dephosphorylation
14	EDC3	Enhancer of mRNA decapping
15	EDC4	Involved in deadenylation dependent decay
16	EXOSC6	Required for rapid degradation of ARA containing RNA but not for poly A shortening
17	EXOSC8	Shows 3'-5' exoribonuclease activity and participates in degrading events eliminating mRNA with processing defects
18	HDAC1	Global repressor but regulate specific genes by repressing and activation at specific parts
19	HDAC3	Effective co repressor acts as binding site for myocyte enhancer factor 2
20	HDAC7	Involves in maturation by repressing of myocyte enhancer factor(MEF-1)
21	HNRNPA2B1	Helps in poly A RNA binding
22	HSPA4	it is co enzyme and enzyme regulator which Binds with ATP, selectively and non covalently
23	KLHL2	Helps the factor binds to RNA
24	MAP3K4	Activates by mytogenic stimulation due to environment
25	MAPKAPK2	Medicates phosphoryl of TTP which regulates myo- D by mRNA decay leads in the activation of satellite cell regulation
26	MED30	Facilitates gene expression by various transcription activation
27	METTL9	Function to help in Protein binding
28	NCL	Abundantly expressed acidic phosphoprotein in growing cell
29	NFKB1	Involved in negative regulation of transcription helps in binding to regulatory region
30	NUP214	Mediates nucleocytoplasmic transport

Analyzing the protein interactions from these two databases we have shortlisted a list of proteins like PRR5L, MAPK3,

CCl3, HDACs etc. having a probable role in muscle regeneration as listed in Table 3.

**Table 3:** TTP interactions playing a role in Muscle regeneration.

S. No.	TTP Interacting Protein	Functions
1	PRR5L	Modulate phosphorylation of PKCA and regulate actin cyto -skeleton organization
2	MAPK3	Extracellular signal regulated kinase important role in MAPK/ERK cascade this signaling pathway mediate cell growth and differentiation through regulating transcription
3	YWHAB	Adaptor protein implicated in the regulation of specialized signaling pathway
4	RGS16	Regulator of G protein signaling family. Inhibition of G protein subunit by GTPase.
5	PRR5L	Modulate phosphorylation of PKCA and regulate actin cyto -skeleton organization
6	CCl3	It is a chemokine which increases rapidly during muscle injury and returns to control levels during muscle regeneration by stimulating myoblast proliferation
7	HDAC1	Global repressor but regulate specific genes by repressing and activation at specific parts
8	HDAC3	Effective co repressor acts as binding site for myocyte enhancer factor 2
9	HDAC7	Involves in maturation by repressing of myocyte enhancer factor(MEF-1)

It can be seen that TTP binds either to ARE-containing mRNAs, or to MAP kinases (Table 1). MAP kinases have a role of phosphorylating TTP at the Ser<sup>52</sup> and Ser<sup>178</sup> positions, so prevents its interface with and employment of explicit deadenylases to the target mRNAs [6]. TTP interactions with YWHAB stops its nuclear translocation [10].

#### 4. Conclusions

There are reports in literature citing the role of TTP in muscle cell regeneration. In this study, we have used the protein interaction databases to explain the role of TTP in muscle regeneration. It was seen that it does so by interacting with a

number of proteins involved in cytoskeleton rearrangement, chemokine secretion, cell growth and signaling pathways (Table 3). This study shows that *insilico* approaches can be used for elucidating protein function. In this study protein interactions have been used to characterize the function of TTP in muscle regeneration. These may be further corroborated by using *invitro* approaches.

#### 5. Acknowledgments

D.A.V. College, Sector-10, Chandigarh for providing the support and infrastructure to carry out the study.

## 6. References

1. Sanduja S, Blanco FF, Young LE, Kaza V, Dixon DA. The role of Tristetraprolin in cancer and inflammation. *Frontiers in bioscience: a journal and virtual library*. 2012; 17:174-188.
2. Zubiaga AM, Belasco JG, Greenberg ME. The nonamer UUAUUUAUU is the key AU-rich sequence motif that mediates mRNA degradation. *Molecular and Cellular Biology*. 1995; 15(4):2219-2230.
3. Carballo E, Gilkeson GS, Blakeshear PJ. Bone Marrow Transplantation Reproduces the Tristetraprolin-Deficiency Syndrome in Recombination Activating Gene-2 (2/2) Mice. *J Clin Invest*. 1997; 100(5):986-995.
4. Qiu L-Q, Stumpo DJ, Blakeshear PJ. Myeloid-Specific Tristetraprolin Deficiency in Mice Results in Extreme Lipopolysaccharide Sensitivity in an Otherwise Minimal Phenotype. *Journal of Immunology*. 2012; 188(10):5150-5159.
5. Sachidanandan C, Sambasivan R, Dhawan J. Tristetraprolin and LPS-inducible CXC chemokine are rapidly induced in presumptive satellite cells in response to skeletal muscle injury. *Journal of CellScience*. 2002; 115:2701-2712.
6. Chrestensen CA, Schroeder MJ, Shabanowitz J, Hunt DF, Pelo JW, Worthington MT *et al*. MAPKAP kinase 2 phosphorylates tristetraprolin on *in vivo* sites including Ser178, a site required for 14-3-3 binding. *J Biol Chem*. 2004; 279(11):10176-84.
7. Mering CV, Huynen M, Jaeggi D, Schmidt S, Bork P, Snel B. STRING: a database of predicted functional associations between proteins. *Nucleic Acids Res*. 2003; 31:258-261.
8. Wheeler DL, Church DM, Federhen S, Lash AE, Madden TL, Pontius JU *et al*. Database resources of the National Center for Biotechnology. *Nucleic Acids Res*. 2003; 31(1):28-33.
9. Stark C, Breitkreutz B, Reguly T, Boucher L, Breitkreutz A, Tyers M. BioGrid: A general repository for interacting datasets. *Nucl. Acids Res*. 2006; 34:535-539.
10. Stoecklin G, Tenenbaum SA, Mayo T, Chittur SV, George AD, Baroni TE *et al*. Genome-wide analysis identifies interleukin-10 mRNA as target of tristetraprolin. *J Biol Chem*. 2008; 283(17):11689-99.