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Role of Kisspeptin in livestock reproduction

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Abstract

The discovery of "Kisspeptin" has opened new vista to understand the regulatory mechanisms of ovarian steroids on control of GnRH secretion. Kisspeptin (Kp) appears to be involved in most (if not all transitional steps) of reproductive life, such as the onset of puberty, initiation of the breeding season and the dynamic changes of gonadotropin secretion throughout the oestrous cycle. Exogenous administration of Kp induces the secretion of gonadotropin (LH) in many species, mainly through stimulation of GnRH secretion including mares, pigs, bovines, ewes, canines, women and ultimately responsible for oocyte maturation and ovulation. Kp stimulates gonadotropin release most potently during the preovulatory phase of the menstrual cycle in women. Infusion of Kp caused synchronization of LH surge and ovulation in progesterone primed cyclic ewes and anoestrus ewes, respectively by activating hypothalamo-pituitary-gonadal axis. In this context, it has been suggested that Kp may be used as new tool to design new protocols for reproductive technologies in mammals.

Keywords: kisspeptin, livestock, reproduction, ovulation

Introduction

The recent advent of a small molecule "Kisspeptin" has opened new vista to understand the regulatory mechanisms of ovarian steroids on control of GnRH secretion (Caraty and Franceschini, 2008) [4]. Kisspeptin (Kp) appears to be involved in most (if not all transitional steps) of reproductive life, such as the onset of puberty, initiation of the breeding season and the dynamic changes of gonadotropin secretion throughout the oestrous cycle (Clarke *et al.*, 2015) [10].

Exogenous administration of Kp induces the secretion of gonadotropin (LH) in many species, mainly through stimulation of GnRH secretion including mares (Wilborn, 2008) [48], pigs (Lents *et al.*, 2008) [21, 22], bovines (Whitlock *et al.*, 2011; Naniwa *et al.*, 2013) [45, 28], ewes (Wang *et al.*, 2012) [43, 44], canines (Albers-Wolthers *et al.*, 2014) [1], women (Jayasena *et al.*, 2014) [17] and ultimately responsible for oocyte maturation and ovulation. It has been observed that, Kp stimulates gonadotropin release most potently during the preovulatory phase of the menstrual cycle in women (Dhillon *et al.*, 2007) [13]. Infusion of Kp caused synchronization of LH surge and ovulation in progesterone primed cyclic ewes and anoestrus ewes, respectively by activating hypothalamo-pituitary-gonadal axis (Caraty *et al.*, 2007) [5, 6]. In this context, it has been suggested that Kp may be used as new tool to design new protocols for reproductive technologies in mammals (Caraty and Franceschini, 2008) [4].

Genes encoding Kp and its receptors (KISS1 and KISS1R) have been documented in the ovaries of rat (Terao *et al.*, 2004) [42], hamsters (Shahed and Young, 2009) [37, 38], pigs and goats (Inoue *et al.*, 2009) [16], primate and human ovaries (Gaytan *et al.*, 2009; Cejudo Roman *et al.*, 2012) [15, 8], this indicate that Kp may also have direct gonadal effects and interact with metabolic pathways (Clarke *et al.*, 2015) [10]. Presence of Kp at high concentrations in porcine follicular fluid than serum revealed the involvement of Kp during follicular development and suggests an intrafollicular or systemic origin of action (Saadeldin *et al.*, 2012) [35]. However, the local role of Kp in controlling the ovulation process has not yet fully understood. Very few *in vitro* studies have been conducted to see the effect of Kp on IVM and fertilization of oocytes in farm animals. The expression of genes encoding Kp and its receptors were detected during IVM period in both oocytes and cumulus cells of pig (Saadeldin *et al.*, 2012) [35] and bovine (Ming *et al.*, 2015) [26] and also suggested that supplementation of Kp in IVM media improved the maturation efficiency of oocytes in pig and bovines. It has been suggested that, Kp has continuous and direct action on oocytes and cumulus cells in an autocrine-paracrine fashion.

Effect of Kisspeptin on reproduction

Kisspeptins (Kp) are peptide products of the KISS1 gene, which was discovered by Lee *et al.*, in 1996 [20] as a metastasis-suppressing gene in malignant melanoma cells (Lee *et al.* 1996) [20]. Kp is now best known as a multifunctional peptide because of its role in cancer, the cardiovascular system and reproduction (Mead *et al.*, 2007) [24]. The reproductive dimension of the KISS1/KISS1R system was discovered in late 2003, when KISS1R (previously known as GPR54) gene mutations were first reported in humans and mice suffering hypogonadotropic hypogonadism (De Roux *et al.*, 2003 and Seminara *et al.*, 2003) [11, 36]. From that moment onwards, Kp became known as a master regulator of the reproductive axis in a hierarchical manner (Tena-Sempere, 2006) [41] and among the most potent elicitors of GnRH-gonadotrophin secretion in a variety of mammalian species, including humans (Roseweir and Millar, 2009 and Pineda *et al.*, 2010) [33, 34, 25]. It works primarily on the hypothalamus to activate GnRH neurons; in addition, neuroanatomical studies have allowed identification of a discrete population of Kp neurons in different hypothalamic areas (Smith *et al.*, 2005; Kauffman *et al.*, 2007) [39, 40]. Kp acts primarily at central levels to regulate ovarian function is well defined, the possibility of additional effects at other sites of the hypothalamic–pituitary– ovarian axis cannot be ruled out (Roa and Tena-Sempere, 2007) [32]. Bilban *et al.* (2004) [2] identified Kp as an endocrine– paracrine regulator of trophoblast invasion.

Expression of Kisspeptin on genes

In rodents, the expression of KISS1 and KISS1R genes were detected in the ovary (Terao *et al.*, 2004) [42]. In addition, Kp and KISS1R immunoreactivity were recently demonstrated in ovarian tissue sections from cyclic rats, where KISS1, but not KISS1R, gene expression was shown to fluctuate in a cycle dependent manner under the regulation of pituitary LH (Castellano *et al.*, 2006) [7]. Moreover, ovarian expression of KISS1 and KISS1R genes was recently documented in fish (Nocillado *et al.*, 2007) [29], hamsters (Shahed and Young, 2009) [37, 38], pigs and goats (Inoue *et al.*, 2009) [16] and primates (Gaytan *et al.*, 2009) [15].

Role of Kisspeptin on ovulation

Kp stimulates ovulation in rats (Matsui *et al.*, 2004), ewes (Caraty *et al.*, 2007) [5, 6] and mares (Briant *et al.*, 2006). Furthermore, inhibitors of cyclooxygenase-2 known to disturb follicular rupture, ovulation and inhibited KISS1 gene expression selectively in the rat ovary (Gaytan *et al.*, 2009) [15]. Together, these reports suggest a local role of Kp is directly controlling the ovulation process, but a clear picture of its local function has not yet fully emerged. Roseweir *et al.* (2009) [33, 34] discovered a potent Kp antagonist (p234) that revealed the critical roles of Kp in both female puberty and the preovulatory gonadotrophin surge (Pineda *et al.*, 2010 and Millar *et al.*, 2010) [30, 25]. Kp enhanced the sensitivity of reproductive steroids to somatotrophic axis and it acts as integrator of LH and Growth Hormone (GH) release (Whitlock *et al.*, 2015) [47]. Kp caused increased LH secretion in bovines (Naniwa *et al.*, 2013; Whitlock *et al.*, 2008) [28, 46], prepubertal heifers (Kadokawa *et al.*, 2008) [18], ewes (Redmond *et al.*, 2011 and Wang *et al.*, 2012) [31, 43, 44], acyclic mares (Wilborn, 2008) [48], swine (Lents *et al.*, 2008) [21, 22] and dogs (Cheng *et al.*, 2011; Elderen, 2014) [9]. Single injection of Kp-54 can induce egg maturation in women with

sub fertility in dose dependent manner (Jayasena *et al.*, 2014) [17].

In vivo effects of Kisspeptin in different livestock species

Bovines

Kadokawa *et al.* (2008) [18] observed that, Kp stimulates the secretion of LH and GH in prepubertal heifers and there is a possibility for important links among Kisspeptin, the reproductive axis, and also the somatotrophic axis. Whitlock *et al.* (2011) [45] reported that, lactation status and stage of lactation did not change the sensitivity of the GH system to Kp. However, an effect of stage of lactation on Kp stimulated LH secretion was detected in the dairy cows. Reproductive steroids enhance the sensitivity of the somatotrophic axis to physiologically relevant doses of Kp and it is an integrator of LH and GH release in bovines (Whitlock *et al.*, 2008) [46]. Administration of full-length Kp causes LH secretion, which is sustained for a few hours, and it is capable of stimulating follicular development and/or ovulation (Naniwa *et al.*, 2013) [28].

Sheep

Kp is a potent stimulator of gonadotropin secretion in sheep. Continuous infusion of Kp can synchronize LH surges in progesterone-primed cyclical ewes and cause ovulation in seasonally acyclic ewes (Caraty *et al.* (2007) [5, 6]. Kp stimulates pulse-like release of LH within 15 min following intra venous injections, and increases the frequency and amplitude of LH pulses and estradiol in prepubertal ewe lambs (Redmond *et al.*, 2011 and Wang *et al.*, 2012) [31, 43, 44]. Whitlock *et al.* (2015) [47] reported that, Kp stimulates the gonadotropic axis of ruminants *in vivo*.

Swine

Lents *et al.* (2008) [21, 22] indicated that, peripheral administration of Kisspeptin increased serum concentrations of LH but not FSH or GH. Thus, Kp can activate gonadotropic hormones but not somatotrophic hormone secretion in prepubertal gilts. But it plays a role in the mechanism involved in initiating puberty in swine.

Dogs

Cheng *et al.* (2011) [9] conducted a study in dogs and observed that, Kp induces LH secretion effectively and also an elevation in blood LH concentrations in neutered dogs. Canine Kp elicits robust gonadotrophin and oestradiol responses in anoestrous bitches, suggesting that canine KISS1/KISS1R are cogent targets for modulating reproduction in dogs (Albers-Wolthers *et al.*, 2014) [1].

Conclusions

The discovery of "Kisspeptin" has opened new vista to understand the regulatory mechanisms of ovarian steroids on control of GnRH secretion. Kisspeptin (Kp) appears to be involved in most (if not all transitional steps) of reproductive life, such as the onset of puberty, initiation of the breeding season and the dynamic changes of gonadotropin secretion throughout the oestrous cycle. Exogenous administration of Kp induces the secretion of gonadotropin (LH) in many species, mainly through stimulation of GnRH secretion including mares, pigs, bovines, ewes, canines, women and ultimately responsible for oocyte maturation and ovulation. Kp stimulates gonadotropin release most potently during the preovulatory phase of the menstrual cycle in women. Infusion

of Kp caused synchronization of LH surge and ovulation in progesterone primed cyclic ewes and anoestrus ewes, respectively by activating hypothalamo-pituitary-gonadal axis. In this context, it has been suggested that Kp may be used as new tool to design new protocols for reproductive technologies in mammals.

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