



ISSN (E): 2277- 7695

ISSN (P): 2349-8242

NAAS Rating: 5.03

TPI 2018; 7(3): 597-601

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www.thepharmajournal.com

Received: 18-01-2018

Accepted: 20-02-2018

**K Prashanth Kumar**

Ph. D Scholar, Department of Poultry Science College of Veterinary Science, P. V. N. R Telangana Veterinary University, Rajendranagar, Hyderabad, Telangana, India

**V Ravinder Reddy**

Professor & University Head, Department of Poultry Science College of Veterinary Science, P. V. N. R Telangana Veterinary University, Rajendranagar, Hyderabad, Telangana, India

**M Gnana Prakash**

Principal Scientist, Poultry Research Station P. V. N. R Telangana Veterinary University, Rajendranagar, Hyderabad, Telangana, India

**Correspondence**

**K Prashanth Kumar**

Ph. D Scholar, Department of Poultry Science College of Veterinary Science, P. V. N. R Telangana Veterinary University, Rajendranagar, Hyderabad, Telangana, India

## Effect of supplementing pomegranate (*Punica granatum*) peel extract on serum biochemical parameters and immune response in broilers during summer

**K Prashanth Kumar, V Ravinder Reddy and M Gnana Prakash**

### Abstract

This study was conducted to evaluate the effects of pomegranate (*Punica granatum*) peel extract (PPE) on serum biochemistry and immunity of broiler chicken reared during summer. The average minimum and maximum room temperatures recorded from 0-42 day were  $36.8\text{ }^{\circ}\text{C}\pm 0.25$ - $39.7\text{ }^{\circ}\text{C}\pm 0.25$ , respectively, with relative humidity ranging from 47-74%. Day old (n=160) broiler chicks were randomised to 4 dietary treatment groups containing eight replicates with five chicks each and are kept in a closed, ventilated, wire-floor battery caged broiler house under uniform conditions of temperature, humidity, and ventilation. The experimental treatment groups were: 1. Positive control (PC)-Corn soya based diet supplemented with Vit-E 70mg/kg + Selenium 0.15 mg/kg, 2. Negative control (NC)-Corn soya based diet without any antioxidant, 3. PPE50-Corn soya based diet supplemented with 50 mg PPE extract/kg, 4. PPE100-Corn soya based diet supplemented with 100mg PPE extract/kg diet. Inclusion of PPE at 50 mg/kg diet and 100 mg/kg diet decreased ( $P<0.05$ ) the serum cholesterol levels, whereas serum protein levels were unaffected ( $P>0.05$ ) by supplementation of PPE. Antibody titer levels in response to New castle disease vaccine and cutaneous basophilic hypersensitivity (CBH) were not significantly ( $P>0.05$ ) influenced by supplementation of PPE in broiler diets.

**Keywords:** *Punica granatum*, biochemical parameters, broilers

### 1. Introduction

Heat stress is an increasing concern to poultry industry in hot humid climates due to reduced growth performance and increased mortality. Like mammals, Birds are homeotherms, thus they are able to maintain a near-constant body temperature. To achieve a constant body temperature, heat produced by metabolism must equal heat loss. In birds, heat losses are limited by dense coverage of feather coat and by the absence of sweat glands, thus these two characters limit their heat transfer ability from the body that generated during metabolism. The main consequence of heat exposure is a reduction in feed intake in order to reduce metabolic heat production (Gous and Morris, 2005) <sup>[1]</sup>. The optimum temperature for performance is likely to be 19 to 22 °C for laying hens and 18 to 22 °C for growing broilers <sup>[2]</sup>. This reduction is approximately 17 % for every 10 °C increase in ambient temperature above 20 °C <sup>[3]</sup>. This decreased feed intake leads to growth depression. However, the reduction in growth is often greater than the reduction in feed intake, resulting in a lower feed efficiency <sup>[4]</sup>. It has been established that high temperatures affect the development of a specific immune response in the chicken <sup>[5-7]</sup>. Thus, concerning the effects of heat stress on birds, the following findings were reported: 1) decreases in the feed consumption, bodyweight gain, as well as the total white blood cell count and antibody production <sup>[8]</sup> 2) decreases in the number of peripheral blood lymphocytes and induction of an electrolyte imbalance <sup>[9]</sup>; 3) decreases in the blood lymphocytes and spleen weight <sup>[10]</sup>; 4) decrease in macrophage activity <sup>[11]</sup>. Increase in stress induces sympatho-adrenal activity which further leads to release of corticosteroids, hormones that increase protein and lipid catabolism in turn elevating plasma cholesterol concentration <sup>[12]</sup>. Hypercholesterolemia is one of the risk factors for coronary artery disease (CAD) <sup>[13]</sup>. Strategies to protect poultry from heat stress effect include i. e. i) Genetical manipulations such as introduction of dwarf gene, Naked neck chicken and frizzle gene ii) Management practices such as the use of insulation, fans, foggers and evaporative coolers. iii) Dietary manipulations such as low protein diets, antioxidant vitamins-Vit A, vit E, vit C and herbal supplements containing different properties like adoptogenic (*Withania somnifera*, *Ocimum sanctum*),

antioxidant (*Punica granatum*, *Phyllanthus emblica*) have been used to protect tissues from oxidative injury and enhances host antioxidant enzyme systems [14, 15]. Among all nutrient manipulations such as inclusion of antioxidants are most economical, practical and easiest methods to adopt. Heat stress impairs absorption of vitamins A, E and C, and, thus, increases the requirement of these vitamins [16] and reduces plasma and tissue concentrations of minerals such as Se, Fe, Zn and Cu which are related to the antioxidant and immune system [17]. Seville *et al.* [18] recently reviewed the composition of pomrgranate, health benefits, including association with antioxidant and immunomodulatory effects. The objective of this study was to evaluate the efficacy of Pomegranate (*Punica granatum*) peel extract on immunity and serum biochemical parameters in broilers reared during hot summer months.

## 2. Material and Methods

One day old broiler chicks were housed in open sided, well ventilated battery cages in broiler house. A total of 160 male broiler chicks from, 1-42 d old were used. The broiler chickens were constantly observed for health status and behaviour. 160 broiler chicks were randomised to 4 dietary treatment groups containing eight replicates with five chicks

each and are kept in a closed, ventilated, wire-floor battery caged broiler house under uniform conditions of temperature, humidity, and ventilation. Birds were vaccinated against Newcastle (7th and 28th d) and infectious bursal disease (15th d) as per the standard vaccination schedule. Maize and soybean meal-based diets were prepared to contain 3050 and 3,150 kcal ME/kg during the starter (0 to 21 d) and finisher (22 to 42 d of age) phases, with respective crude protein contents of 21.5 and 19.5 g/100 g feed (Table 1). The experimental treatment groups were as follows:

1. Positive control (PC)-Corn soya based diet supplemented with Vit-E 70mg/kg + Selenium 0.15 mg/kg.
2. Negative control (NC)-Corn soya based diet without any antioxidant.
3. PPE50-Corn soya based diet supplemented with 50 mg PPE extract/kg.
4. PPE100-Corn soya based diet supplemented with 100mg PPE extract/kg diet.

Water and mash feed were provided *adlibitum* throughout 42 days trial period. The temperature and relative humidity of shed during trial period were 36.8 °C±0.25-39.7 °C±0.25 and 47-74% respectively.

**Table 1:** Ingredient and nutrient composition of basal diet

Ingredients (%)	Starter (0-21d)	Finisher (22-42d)
Maize	56.9	61.31
Soya bean meal	35.43	30.60
Vegetable oil	3.355	4.287
Salt	0.423	0.422
Dicalcium phosphate	1.602	1.542
Limestone powder	1.343	1.177
DL-methionine	0.278	0.217
L-Lysine	0.241	0.076
AB <sub>2</sub> D <sub>3</sub> K*	0.015	0.015
B-complex**	0.015	0.015
Trace mineral mixture***	0.100	0.100
<b>Nutrient composition</b>		
ME (kcal/kg)	3050	3150
Crude protein (%)	21.5	19.5
Lysine (%)	1.25	0.98
Methionine (%)	0.56	0.48
Calcium (%)	0.9	0.82
Available Phosphorous (%)	0.42	0.40

\*AB<sub>2</sub>D<sub>3</sub>K provided per kg diet: Vitamin A 20000 IU, Vitamin B<sub>2</sub> 25 mg, Vitamin D<sub>3</sub> 3000IU, Vitamin K 2mg.

\*\*Riboflavin 25mg, Vitamin B<sub>1</sub> 1mg, Vitamin B<sub>6</sub> 2mg, Vitamin B<sub>12</sub> 40mg, and Niacin 15mg.

\*\*\*Trace mineral provided per kg diet: Manganese 120mg, Zinc 80mg, Iron 25mg, Copper 10mg, Iodine 1mg.

### 2.1 Immunity

The humoral immunity was determined in birds by measuring antibody titre to Newcastle disease (ND) vaccine (antibody production against ND virus). At 21st and 42nd days of age blood was collected and serum was separated. Haemagglutination inhibition (HI) activity of serum was estimated and the antibody titers (log<sub>2</sub>) were measured following the standard procedure [19].

The cell mediated immune (CMI) response was assessed by measuring cutaneous basophilic hypersensitivity (CBH) to phytohaemagglutinin phosphate (PHA-P). On 40th day of experiment, one bird per replicate was injected with 100 µg of PHA-P suspended in 0.1 ml of phosphate buffer saline (PBS) into the web between third and fourth inter-digital space of right foot, while the left web (control) was injected with 0.1 ml of PBS. The web thickness of both feet was measured by micrometer after 24 h of injection and CBH was calculated by

method of Edelman [20].

### 2.2 Serum Bio-Chemistry

On 41st day of experiment, blood collected from one bird per replicate into a clean sterilized tube and kept in a slanted position at room temperature to facilitate separation of serum. Serum protein and cholesterol were determined by using commercially available diagnostic kits (Erba Mannheim Chemicals).

### 2.3 Statistical analysis

The data were analysed using General Model procedure of Statistical Package for Social Sciences (SPSS) 15<sup>th</sup> version and comparison of means was done using Duncan's multiple range test [21] and significance was considered at  $P < 0.05$ . Data were subjected to statistical analysis under completely randomized design employing one-way analysis of variance [22].

### 3. Results

Table 2 shows the effect of supplementing of pomegranate peel extract on serum biochemistry in broilers reared during summer. Dietary supplementation of PPE100 and PPE50 tended to decrease ( $P<0.05$ ) the cholesterol levels in serum compared to that of NC group. PPE100 group recorded lowest cholesterol levels. However, there was no effect ( $P>0.05$ ) of dietary supplementation of PPE on serum protein levels. Inclusion of PPE at 50 mg/kg and 100 mg/kg had no significant ( $P<0.05$ ) effect on mean antibody titer against Newcastle disease virus and cell mediated immune response to phytohaemagglutinin phosphate (Table 3).

### 4. Discussion

Increase in stress induces sympatho-adrenal activity which further leads to protein and lipid catabolism in turn elevating plasma cholesterol concentration [12]. Low cholesterol in PC compared to NC may be due to Selenium, which inhibitory effect on 3-hydroxy-3-methylglutaryl-coA (HMG-CoA) reductase, a key enzyme in cholesterol biosynthesis [23]. By quenching free radicals vit-E reduces the effects of oxidative stress there by reduces corticosteroids secretion [24]. Similar results were found by Habibian [25] by feeding vit-E at 250 mg/kg & Se at 0.5-1 mg /kg diet. In the present study decreased serum cholesterol levels by feeding PPE in diets was in agreement with Kishawy *et al.* (2016) who reported reduced cholesterol in serum by feeding PPE at 0.05% and 0.1% levels. Similarly results observed by [27, 28] Yaseen *et al.* (2016) and Sarica *et al.* (2016) by feeding PP at 15/kg and PPE at 0.1g/kg, 0.2g/kg respectively. PPE exerts inhibitory effect on pancreatic lipase activity that inhibit fat absorption from intestinal tract and increase faecal excretion of fat [26, 29] this suggests the reduced serum total cholesterol levels of PPE fed groups in present study. Supplementation of PPE did not influence the immune parameters in present study. However, improved immune response by feeding pomegranate seed pulp and polyphenol rich grape seed [30-32]. Improvement in immune responses might be due to presence of linoleic acid, n-3 fatty acids of seeds, which are known to increase immunoglobulin's and weight of immune organs [33, 34].

**Table 2:** Effect of dietary inclusion of pomegranate peel extracts on serum biochemical profile in broiler chicken at 42d of age.

Treatments	Cholesterol (mg/dl)	Total Protein (g/dl)
PC	148.6 <sup>ab</sup>	3.26
NC	164.4 <sup>a</sup>	2.85
PPE50	142.3 <sup>b</sup>	3.08
PPE100	135.9 <sup>b</sup>	3.52
SEM	2.283	0.076
N	8.0	8.00
P-Value	0.012	0.167

A b c d Mean bearing at least one common superscript in a column do not differ significantly ( $P>0.05$ )

In addition, some polyphenols with dihydroxy groups can conjugate metals, preventing metal-catalyzed free radical formation [35]. Essentially it is iron, as well as other redox active metals (e. g. copper and manganese) catalyze the decomposition of hydrogen peroxide into hydroxyl radicals which is one of the most powerful oxidant species, being able to initiate free radical chain reactions by abstracting hydrogen from almost any molecule (Fatty acids i. e linoleic acid). *Ellagitannins* which present in PPE also have immunomodulatory function by stimulating growth of lymphocytes [36].

**Table 3:** Effect of dietary inclusion of pomegranate peel extracts on immune response of broiler chicken

Treatments	NDV titers (log <sub>2</sub> )		*PHA-P response (thickness index)	Spleen weights (% live weight)
	3 <sup>rd</sup> week	6 <sup>th</sup> week	6 <sup>th</sup> week	
PC	3.25	3.75 <sup>a</sup>	164.1	0.092
NC	3.50	2.75 <sup>ab</sup>	144.3	0.078
PPE50	2.88	2.38 <sup>b</sup>	151.2	0.091
PPE100	2.50	3.00 <sup>ab</sup>	155.4	0.097
SEM	0.133	0.132	2.355	0.005
N	8.00	8.00	8.0	8.000
P-Value	0.678	0.103	0.452	0.880

A b c d Mean bearing at least one common superscript in a column do not differ significantly ( $P>0.05$ ).

High ambient temperatures was reported to reduce weights of lymphoid organs [37] in broilers and heat stress during experimental period might be responsible for decreased circulating macrophages and antibodies [8-11, 38, 39]. This reduction could be indirectly due to stimulation of hypothalamic pituitary adrenal (HPA) axis which results in release of corticosterone that inhibits antibody production [40, 41].

We speculate that reduced immunity in the present study might be due to low doses of PPE and high temperatures (36.3 °C-39.7 °C) during experiment. Temperature effect can be correlated with reduced spleen weights in this study.

### 5. Conclusion

The results of present study indicated that supplementation of Pomegranate peel extract at 100mg/kg diet significantly decreased serum cholesterol levels without any deleterious effects on performance and did not show any significant effect on immunity.

### 6. Acknowledgement

The authors are thankful to college of veterinary science, Rajendranagar, PVNR TVU, for providing necessary facilities to carry out the present study.

### 7. References

- Gous RM, Morris TR. Nutritional interventions in alleviating the effects of high temperatures in broiler production. *World's Poultry Science Journal*. 2005; 61(3):463-475.
- Charles DR. Responses to the thermal environment. *In: Poultry Environment Problems, A guide to solutions* (Charles, D. A. and Walker, A. W. Eds.), Nottingham University Press, Nottingham, United Kingdom, 2002, 1-16.
- Austic RE. Feeding poultry in hot and cold climates. *In Stress Physiology in Livestock*, [M. K. Yousef, editor]. Boca Raton: CRC Press. 1985; 3:123-136.
- Howlider MAR, Rose SP. Temperature and growth in broilers. *World's Poultry Science Journal*. 1987; 43:228-233.
- Thaxton PC, Sadler R, Glick B. Immune response of chickens following heat exposure or injection with ACTH. *Poult. Sci*. 1968; 47:264-266.
- Subba Rao DSV, Glick B. Immunosuppressive action of heat in chickens. *Proc. Soc. Exp. Biol. Med*. 1970; 133:445-448.
- Thaxton P, Siegel HS. Depression of secondary immunity

- by high environmental temperature. *Poult. Sci.* 1972; 51:1519-1526.
8. Mashaly MM, Hendricks 3<sup>rd</sup> GL, Kalama MA, Gehad AE, Abbas AO, Patterson PH. Effect of heat stress on production parameters and immune responses of commercial laying hens. *Poultry Science.* 2004; 83(6):889-894.
  9. Borges SA, Fischer da Silva AV, Majorca A, Hooge DM, Cummings KR. Physiological responses of broiler chickens to heat stress and dietary electrolyte balance (sodium plus potassium minus chloride, milliequivalents per kilogram). *Poultry science.* 2004; 83(9):1551-1558.
  10. Trout JM, Mashaly MM. The effects of adrenocorticotrophic hormone and heat stress on the distribution of lymphocyte populations in immature male chickens. *Poultry science.* 1994; 73(11):1694-1698.
  11. Smith MO. Effects of different levels of zinc on the performance and immunocompetence of broilers under heat stress. *Poultry science.* 2003; 82(10):1580-1588.
  12. Sahin N, Onderci M, Sahin K, Gursu MF, Smith MO. Ascorbic acid and melatonin reduces heat induced performance inhibition and oxidative stress in Japanese quails. *Br. Poult. Sci.* 2004; 45:116-122.
  13. Gambhir DS, Gambhir JK, Sudha R. Dyslipidemia and coronary heart disease: management issues from Indian perspective, 2000.
  14. Davis L, Kuttan G. Immunomodulatory activity of *Withania somnifera*. *Journal of Ethnopharmacology.* 2000; 71(1):193-200.
  15. Saravanan S, Sri Kumar R, Manikandan S, Parthasarathy NJ, Devi RS. Hypolipidemic effect of triphala in experimentally induced hypercholesteremic rats. *Yakugaku Zasshi.* 2007; 127(2):385-388.
  16. Naziroglu M, Sahin K, Simsek H, Aydilek N, Ertas ON. The effects of food withdrawal and darkening on lipid peroxidation of laying hens in high ambient temperatures. *DTW. Deutsche tierärztliche Wochenschrift.* 2000; 107(5):199-202
  17. Combs GF, Combs SB. The nutritional biochemistry of selenium. *Annual review of nutrition.* 1984; 4(1):257-280.
  18. Sevilla AA, Pastor AS, Barrachina AC. Pomegranate and pomegranate juices. *Alimentacion, Equipos y Tecnologia.* 2008; 234:36-39.
  19. Edelman AS, Sacher PI, Robinson ME, Hochwalad GM, Thorbecke GJ. Primary and Secondary Swelling response to Phytohaemagglutinin as a measure of immune competence in chickens. *Avian Diseases.* 1986; 30:105-111.
  20. Wegmann TG, Smithies O. A simple hemagglutination system requiring small amounts of red cells and antibodies. *Transfusion.* 1966; 6(1):67-73.
  21. Snedecor GWC, William G. *Statistical Methods/George W. Snedecor and William G. Cochran (No. QA276. 12. S6313 1989.)*, 1989.
  22. Duncan DB. Multiple range and multiple F tests. *Biometrics.* 1955; 11(1):1-42.
  23. Nassir F, Moundras C, Bayle D, Serougne C, Gueux E, Rock E, Mazur A. Effect of selenium deficiency on hepatic lipid and lipoprotein metabolism in the rat. *British Journal of Nutrition.* 1997; 78(3):493-500.
  24. Sahin K, Sahin N, Yaralioglu S. Effects of vitamin C and vitamin E on lipid peroxidation, blood serum metabolites, and mineral concentrations of laying hens reared at high ambient temperature. *Biological trace element research.* 2002; 85(1):35-45.
  25. Habibian M, Ghazi S, Moeini MM, Abdolmohammadi A. Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermoneutral or heat stress conditions. *International journal of biometeorology.* 2014; 58(5):741-752.
  26. Kishawy AT, Omar AE, Gomaa AM. Growth performance and immunity of broilers fed rancid oil diets that supplemented with pomegranate peel extract and sage oil. *Japanese Journal of Veterinary Research.* 2016; 64(2):S31-S38.
  27. Yaseen AT, El-Kholy MESH, El-Razik WMA, Soliman MH. Effect of Using Pomegranate Peel Extract as Feed Additive on Performance, Serum Lipids and Immunity of Broiler Chicks. *Zagazig Veterinary Journal.* 2016, 42(1).
  28. Sarica S, Urkmez D. The use of grape seed-olive leaf-and pomegranate peel-extracts as alternative natural antimicrobial feed additives in broiler diets. *European Poultry Science,* 2016, 80.
  29. Lei F, Zhang XN, Wang W, Xing DM, Xie WD, Su H *et al.* Evidence of anti-obesity effects of the pomegranate leaf extract in high-fat diet induced obese mice. *International Journal of Obesity.* 2007, 31(6).
  30. Yamasaki M, Kitagawa T, Koyanagi N, Chujo H, Maeda H, Kohno-Murase J *et al.* Dietary effect of pomegranate seed oil on immune function and lipid metabolism in mice. *Nutrition.* 2006; 22(1):54-59.
  31. Hao R, Li Q, Zhao J, Li H, Wang W, Gao J. Effects of grape seed procyanidins on growth performance, immune function and antioxidant capacity in weaned piglets. *Livestock Science.* 2015; 178:237-242.
  32. Ahmed ST, Yang CJ. Effects of Dietary Punica granatum L. By-products on Performance, Immunity, Intestinal and Fecal Microbiology, and Odorous Gas Emissions from Excreta in Broilers. *The Journal of Poultry Science.* 2017; 54(2):157-166.
  33. Wang YW, Field CJ, Sim JS. Dietary polyunsaturated fatty acids alter lymphocyte subset proportion and proliferation, serum immunoglobulin G concentration, and immune tissue development in chicks. *Poultry science.* 2000; 79(12):1741-1748.
  34. Yamasaki M, Kitagawa T, Chujo H, Koyanagi N, Nishida E, Nakaya M *et al.* Physiological difference between free and triglyceride-type conjugated linoleic acid on the immune function of C57BL/6N mice. *Journal of agricultural and food chemistry.* 2004; 52(11):3644-3648.
  35. Guo Q, Zhao B, Li M, Shen S, Xin W. Studies on protective mechanisms of four components of green tea polyphenols against lipid peroxidation in synaptosomes. *Biochim. Biophys. Acta.* 1996; 1304:210-222.
  36. Fraga CG. Plant polyphenols: how to translate their *in vitro* antioxidant actions to *in vivo* conditions. *IUBMB life.* 2007; 59(4-5):308-315.
  37. Hassan AM, Hussein MM, Osman MM. Effect of chronic heat stress on broiler chick's performance and immune system. *SCVMJ.* 2007; 12:55-68.
  38. Quinteiro-Filho WM, Ribeiro A, Ferraz-de-Paula V, Pinheiro ML, Sakai M, Sá LRM *et al.* Heat stress impairs performance parameters, induces intestinal injury, and decreases macrophage activity in broiler chickens. *Poultry Science.* 2010; 89(9):1905-1914.

39. Ghazi SH, Habibian M, Moeini MM, Abdolmohammadi AR. Effects of different levels of organic and inorganic chromium on growth performance and immunocompetence of broilers under heat stress. *Biological trace element research*. 2012; 146(3):309-317.
40. Gross WB. Effect of short-term exposure of chickens to corticosterone on resistance to challenge exposure with *Escherichia coli* and antibody response to sheep erythrocytes. *American journal of veterinary research*. 1992; 53(3):291-293.
41. Star L, Decuypere E, Parmentier HK, Kemp B. Effect of single or combined climatic and hygienic stress in four layer lines: 2. Endocrine and oxidative stress responses. *Poultry science*. 2008; 87(6):1031-1038.