



ISSN (E): 2277- 7695  
ISSN (P): 2349-8242  
NAAS Rating: 5.03  
TPI 2018; 7(12): 175-181  
© 2018 TPI  
www.thepharmajournal.com  
Received: 02-10-2018  
Accepted: 05-11-2018

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## An updated review on pharmacological studies of *Rumex nepalensis*

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### Abstract

Medicinal plants are necessary part of human society since the civilization started. Medicinal plants are the boon of nature to treat a number of ailments of human beings. Plants contain bioactive molecules and thus can provide lead structures for the development of alternative medicines to available toxic commercial drugs with better effectively and increased safety. Rumex, a genus of polygonaceae family, is very prevalent worldwide. There are about 200 species of this genus, many of which are beneficial and used as vegetables and for their medicinal properties. Root, seed, leaf, fresh plant juice, aerial parts etc. are the parts generally used. *Rumex nepalensis* Spreng. (Polygonaceae) commonly known as Nepal Dock has wide-spectrum therapeutic potencies and is extensively used for centuries in traditional medicine systems. The leaves of this plant are edible and a rich source of natural antioxidants. They act as a possible food supplement and are largely used in pharmaceutical industry. Extracts and metabolites from this plant exhibit many pharmacological activities. Due to its remarkable biological activities, it has the potential to act as a rich source of drug against life threatening diseases. However, more studies are needed to scientifically validate the traditional uses of this plant, beside isolating and identifying their active principles and characterizing the mechanisms of action. We present herein a decisive account of its botany, ecology, traditional uses, phytoconstituents profile and major pharmacological activities reported in recent years and therefore will provide a source of information on this plant for further studies.

**Keywords:** Medicinal plants, polygonaceae, *Rumex nepalensis*

### Introduction

The evolution of mankind has happened along with their surrounding herbs and medicinal plants from ancient period of time. Approximately 270,000 plant species have been discovered by human, while there is a possible of existence of close to 400,000 species in mother earth [1]. No synthetic substitute is currently available for about 121 major plant based drug molecules, comprising 45 from tropical and 76 from subtropical areas. A recent study by World Health Organization claimed 80% dependency of world population on ethno medicines in some extent [2]. Medicinal plants are nature's gift for answering a limitless range of fatal diseases among human beings, therefore medicinal plants are getting more consideration currently than ever, especially in the line of medicine and pharmacology. The bioactive phytochemical constituents of the plant are being explored worldwide for their broad-spectrum medicinal potencies. Medicinal plants are explored as a source to isolate pure active principles or in the form of phyto complex, where there is a synergistic combination of active ingredients and other substances like enzymes, resins, essential oils, tannins to facilitate their actions. The health-promoting properties of medicinal plants are usually derived from the interaction of all the substances naturally present in the phyto complex. However, the emphasis on the use of total herbs as medicines and food supplements is gradually replacing the techniques to isolate the biologically active novel compounds and molecules as leading drug molecules [3]. The *Rumex* species, belonging in the Polygonaceae family, comprise about 200 species widely distributed around the World. The name *Rumex* originated from the Latin word for dart, alluding to the shape of the leaves [4]. There have been numerous ethno botanical and ethno pharmacological literature reports dealing with the occurrence and traditional uses of *Rumex* species [5-7]. *Rumex Nepalensis* Spreng. (*R. Nepalensis*) is a perennial, ascending herb [8], commonly known by the name Nepal dock and vernacular names are in English-Sheep sorrel, Nepal duck, Hindi-Jangli palak, Amla, Amlora, Bhilmora, Malori, Sanskrit-Amlavetasa, Kashmiri-Aliphiri, Bengali-Pahari palang, Pakistani-Shalkhay, Hoola, Nepali-Halhale sag, Ban haldi, Halya, Halye, Uttarakhandi- Kathura, Tamil-Sukkankeerai, Manipuri-Torongkhongchak [3].

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Though *R. Nepalensis* is an agricultural weed but this wild plant is not really unwanted in the arena of traditional herbal medicines [9-11]. Several studies have confirmed striking medicinal benefits of this plant. The juvenile leaves of this plant are cooked as vegetables which gives an acidic-lemon flavor to dishes [12]. The young shoot is also locally eaten as a cooked vegetable [13]. This plant is also used as a colouring agent (dye) [14]. Green colour from the leaves of plants is often used in sweet preparations [15]. Phytochemical screening shows that the *R. Nepalensis* contains various constituents viz., sterols, amino acids, quercetin, alkaloid, phenolic components, flavonoids, triterpenoids, stilbene glycosides, tannic acid, saponins, resveratrol, anthraquinone glycosides, anthraquinone, vitamin C, some cardiac glycoside, naphthalene's and many more [8,12,16]. The foundations of modern drugs are based on these natural compounds. In North Western Himalaya, *R. Nepalensis* is a high value medicinal herb due to its high anthraquinone content [17]. *R. Nepalensis* has shown purgative, antioxidant, antifungal, antibacterial, antihistaminic, anticholinergic, antibradykinin ant prostaglandin, antipyretic, anti-inflammatory, antialgal, insecticidal [8, 18-20], analgesic and CNS depressant properties. The plant is also reported to possess skeletal muscle relaxant activity [21]. The present review provides a survey of the current state of knowledge of the Ethnopharmacology, Phytochemistry, pharmacological activities, toxicity and safety of *R. Nepalensis*, as well as their traditional uses which have been supported by pharmacological investigations in order to identify their relevance as food and potential therapeutic applications and to show further directions of research.

#### Dissemination of *Rumex nepalensis*

*R. Nepalensis* grows in parts of China, Afghanistan, India, Indonesia, Japan, Myanmar, Nigeria, Nepal, Pakistan, Tajikistan, Vietnam, South-west Asia, Turkey, Bhutan and South Africa. It grows abundantly in many parts of India. It is widely distributed throughout Himalayas from Bhutan to Kashmir. It is a fairly common plant of higher altitudes and grows between 900-4000 m on moist as well as dry slopes, under shades, and even in plains [8], Western Ghats, Nilgiri, Pulney hills, Nainital hills and Palni hills at altitudes between 1200-4 300 m. *R. nepalensis* shows flowering from April to May; fruiting from June to July [21-23]. A plant of the warm temperate to tropical zones, usually at higher elevations in the tropics. Succeeds in most soils but prefers a deep fertile moderately heavy soil that is humus-rich, moisture-retentive but well drained and a position in full-sun or part shade [24].

#### Morphological Description

Herbs are perennial with long tap roots, erect stems (50- 100 cm tall) which are branched, glabrous, grooved, green or pale brown in color. Basal leaves: petiole 4-10 cm; leaf blade broadly ovate, 10-15 × 4-8 cm, both surfaces glabrous or abaxially minutely papillate along veins, base cordate, margin entire, apex acute; cauline leaves shortly petiolate, ovate-lanceolate; ocrea fugacious, membranous, inflorescence paniculate. Flowers bisexual. Pedicel articulates below middle. Outer tepals elliptic, ca. 1.5 mm; inner tepals enlarged in fruit; valves broadly ovate, 5-6 mm, valves all or 1 or 2 with tubercles, base truncate, each margin with 7 or 8 teeth, apex acute; teeth 1.5-3 mm, apex hooked or straight. Achenes brown, shiny, ovoid, sharply trigonous, ca. 3 mm, base truncate, apex acute [25]. Part of this plant which is used in its

propagation is seed. Vashistha *et al.* [26] studied the phenological observations of this plant and found that the growth initiation occurs in May, senescence occurs in October, wherein flowering (reddish) occurs in between June and July flower, and fruiting takes place from August to September [27].

#### Anatomical studies

##### Leaf

In leaves epidermis is single layered. Irregular epidermal cells with undulating walls are restricted to abaxial surface. Thick and pitted walls have been noted only along with the presence of crystalliferous cells in epidermis [28]. The cells are spherical. Mesophyll is elongated. Collenchyma and sclerenchyma is absent. Endodermis is single layered. Pericycle is oval and single layered. Xylem is oval and phloem is spherical, whereas pith is absent [29].

##### Stomata

Stomata are distributed on both leaf surfaces [28]. Stomata in the upper epidermis of this plant are paracytic and anisocytic, and in the lower epidermis are anisocytic. The percentage of the open and close stomata in the upper epidermis of *R. nepalensis* is 42 and 58, and that of the lower epidermis is 30 and 70 [30].

##### Trichomes

Glandular trichomes could be considered facultative salt glands and they may be part of apparatus of dispersion of extreme radiation. Non glandular trichomes are totally absent, while glandular trichomes are peltate and 1-4 celled centrally [28].

##### Roots

Epidermis in roots is single layered. Polygonal shape parenchyma is compactly packed. Collenchyma and sclerenchyma is absent. Endodermis is single layered. The cells of endodermis are elongated. Pericycle is single layered having spherical shape. Xylem is oval shaped, phloem and pith is spherical. Stone cells are absent [29].

##### Stem

Epidermis in stem of *R. nepalensis* is single layered and shape is oval. Collenchyma is present which is spherical in shape. Sclerenchyma is absent. Single layered endosperm is present. The cells are oval. Pericycle is single layered and spherical in shape. Xylem is oval shaped, phloem and pith is spherical. Mean length and width of the cell in *R. nepalensis* is 52.1 µm and 16.9 µm, respectively. Stone cell is absent in stem [29].

##### Petiole

Epidermis is single layered. The cells are oval in shape. Parenchyma is compactly packed. It is spherical. Collenchyma is present and is spherical in shape. Mean length and width of the cell 50.1 µm and 23.9 µm. Sclerenchyma is absent. Endodermis is single layered. The cells are oval in shape. Pericycle is single layered and spherical. Xylem is oval in shape. Phloem is spherical. Pith is spherical in shape in *R. Nepalensis* [29].

##### Pollen

Pollen grains usually tricolpate and tetracolpate, circular in shape. The size of pollen grain (polar axis × equatorial diameter) reported is (24×22) µm. The pollen is radially

symmetrical and isopolar. Under scanning electron microscope, tectum can be seen as perforate punctate in *R. Nepalensis* [31].

### Plant Development and Flexibility

*R. Nepalensis* is herb that grows in fertile areas that is rich in nitrogen. The plant is palatable to cattle and is high in fibre and nutrition [32]. Reports revealed that enzymatic activities of this plant related to carbon metabolism such as aspartate aminotransferase (EC 2.6.1.1), ribulose-1,5-bisphosphate carboxylase / oxygenase (EC 4.1.1.39), phosphoenolpyruvate carboxylase (EC 4.1.1.31) and glutamine synthetase (EC 6.3.1.2) increased with altitude viz. 1 300, 2 250, and 3 250 m. The elevated oxygenase activity of ribulose-1,5-bisphosphate carboxylase/ oxygenase in *R. Nepalensis* supports its role in protection against photo oxidative damage. These enzymatic alterations also provide adaptive advantage to plant in order to conserve carbon and nitrogen at high elevation [33]. When *R. nepalensis* is exposed to CO<sub>2</sub> to assess its effect on their growth, it is found that elevated CO<sub>2</sub> (EC) has been reported to enhance vegetative growth and biomass accumulation through enhanced photosynthetic activity in annual C<sub>3</sub> plants. Therefore, elevated CO<sub>2</sub> significantly stimulates the growth and biomass through increased plant height, leaf number and area [34].

### Uses in Traditional Medicine

*R. Nepalensis* has served as the basis of traditional medicine systems in India, Nigeria, China and Indonesia. *R. Nepalensis* is used for various therapeutic purposes and is well known in Indian traditional medicine. Plant of *R. Nepalensis* is consumed fried in olive oil or sautéed with butter or lard or are used as filling for pie [35]. The leaves of this plant are diuretic, astringent and demulcent. It also soothes the irritation caused by *Urtica dioica* L. [36]. This plant is used for treatment of scurvy as it is rich in vitamin C [37] and also for treatment of rheumatism [21]. Infusion of leaves is purgative [38], the juice of the leaves is applied externally to relieve headache and also used for its astringent qualities [39]. Its leaf extract has antiseptic properties and is used to stop bleeding. It is also used against allergy caused by leaves of *Acacia nilotica* (L.) Willd ex Delile [40] and also for the treatment of syphilitic and colic ulcers [8]. Leaf extract is applied to cure skin sores. Aqueous extract is used as wash for reducing body pain. Leaf powder mixed with butter is used to treat scabies [41, 42]. Infusion of leave is used in dysmenorrhoea and stomach ache [43, 44]. Crushed leaf extract is applied externally on cuts, boils, blisters and wounds as an anti-allergic [42, 45, 46]. This plant is also used to treat skin infections in Jimma [47]. *R. Nepalensis* acts as a favourite source of fodder for cows, horses and sheep [9, 48]. Leaves are crushed and solution is made and used as pesticide to kill pests. Also leaves are crushed and paste is made with milk, churned curd, or with the urine of cow, and applied on the area around the snake bite on the body [46]. The fresh young leaves of *R. Nepalensis* are rubbed over the affected areas after injury from stinging nettles [49]. Root juice of *R. Nepalensis* is orally given on empty stomach as an effective cure for jaundice [50, 51]. The roots of plant are traditionally used for the treatment of pain, inflammation [52], bleeding, constipation and tinea in Chinese folk medicine [8]. The pounded root is given to animals in case of diarrhoea [43] and dysentery [41, 42]. The root of *R. Nepalensis* is also used as an astringent [36], purgative [19, 53]. A decoction is applied to dislocated bones and to reduce body pain. A paste of the root is applied to swollen gums, pimples

and ringworm [39, 52, 54] and applied externally to relieve headache [8, 55, 56]. Methanolic root extract is applied in joint pain, paralysis and significantly possesses hypotensive effect and also shows property of muscle relaxant and tranquilizer activity [57]. Traditional medicine practitioners of Bale used *R. nepalensis* to treat diarrhoea, blackleg and swelling. In Ethiopia this plant is used to treat colic in livestock and as an antidote for poisoning as well as a laxative [58]. The roots of the plant have been used in folklore medicine to relieve mental tension and disturbance [49]. Root is crushed and the juice is applied on the scalp to prevent hair loss [46]. Half spoon of the grounded floral parts and root extract is used to cure joint pain. It also cures body ache. Roots grounded powder is applied on burned body part to avoid infection and for immediate healing [59]. The roots of plant are also used in traditional Chinese medicine for the treatment of emostasis [60]. Crushed fresh root and leaf with water is taken orally to treat tonsillitis [61]. In South Africa strong decoction of leaf in tablespoon doses 3 times daily to treat bilharziasis [62].

### Phytochemical constituents

*R. Nepalensis* has been reported to contain phytochemicals like phenols, flavonoids, anthraquinones, naphthalenes, saponins, cardiac glycosides, stilbene glycosides, triterpenoids, anthraquinone glycosides, tannic acid and sterols, tannins, steroids, reducing sugar, saponin and sitosterols. Investigation of the *n*-butanolic extract of the roots of *R. nepalensis* yielded two *seco* anthraquinone glucosides, nepalensides A and B and the *seco* nor derivative aloesin. The *seco*-anthraquinones are probably formed by the decomposition and oxidation of the anthraquinones chrysophanein and pulmatin [60]. Emodin, Chrysophanol, physcion, citreoresin, endocrocic, emodin-8-*O*- $\beta$ -D-Glucopyranoside, chrysophanol - 8 - *O* -  $\beta$  D Glucopyranoside, chrysophanol -8-*O*- $\beta$ -D-(6'-*O*-acetyl) glucopyranoside and emodin-8-*O*- $\beta$ -D-(6'-*O*-acetyl) glucopyranoside, were isolated from the roots of *R. Nepalensis* [63]. From the roots of *R. nepalensis*, aloesin, rumexoside, orientalosite and torachryson were isolated by [60]. Later, Gautam *et al.* reported nepodin and its glucoside from the plant [64]. Liang *et al.* identified nepodin-8-*O*- $\beta$ -D-glucopyranoside, torachryson, torachryson-8-*O*- $\beta$ -D-glucopyranoside and two naphthalene acylglucosides, rumexneposides A and B and Epicatechin and epicatechin-3-*O*-gallate were then detected in the roots of the plant [60]. An investigation of the EtOAc fraction of *R. nepalensis* roots and the ethanolic extract of *R. hastatus* roots also resulted in the isolation of resveratrol [65]. From the roots of *R. nepalensis*, (3,5-dimethoxy-4-hydroxyphenol)-1-*O*- $\beta$ -D-(6-*O*-galloyl) glucose, orcinol-glucoside, a lignan derivative, lyoniresinol 3 $\alpha$ -*O*- $\beta$ -D-glucopyranoside was isolated [60]. It is reported that anthraquinones have several pharmacological properties like antifungal, anti-inflammatory, antioxidant and anticancer, whereas naphthalene derivatives possess anti-inflammatory and antioxidant activities [3].

### Pharmacological properties

Researchers reported the different pharmacological activities of *R. nepalensis*, which are discussed in the following sections.

### Antioxidant activities

Several studies showed the antioxidant activity of *R. nepalensis*. Water, ethyl acetate, ethanol, methanol, acetone extracts of this plant has been shown to have antioxidant

properties. *In vitro* assays such as 2,2'-azinobis (3-ethylbenzothiazoline-6-sulphonic acid (ABTS), 2,2-diphenyl-1-picrylhydrazyl (DPPH), superoxide, hydroxyl and nitric oxide radical scavenging were employed to evaluate free radical scavenging and antioxidant potential of methanol, water and acetone extracts [66]. The components of ethanolic, chloroform and ethyl acetate extracts are inhibitors of DPPH radical [67], and also are reported to scavenge the NO radical, inhibit the lipid-peroxidation and also chelate the metal ions [68]. Bhattacharya *et al.* [69] evaluated antioxidant potentials of *in vitro* propagated plants of *R. nepalensis*, with highest activities in plants obtained through indirect shoot organogenesis. The antioxidant effects (DPPH and ABTS) of the chloroform and ethyl acetate extracts of *R. nepalensis* root were evaluated. Both fractions contained phenolic compounds, but their level was higher in the ethyl acetate extract (27.71%) than in the chloroform extract (8.20%). Trolox (IC<sub>50</sub> = 15.7 µM in the case of DPPH, and 16.2 µM in the case of ABTS) and ascorbic acid (IC<sub>50</sub> = 22.4 µM in the case of DPPH, and 25.5 µM in the case of ABTS) were used as positive controls. These extracts contained anthraquinones and naphthalenes too. The higher radical scavenging activity could be due to the phenolic content (in the case of the ethyl acetate extract) and the presence of nepodin (in the case of the chloroform extract) [64].

#### Antitumour activity

A cytotoxic assay of emodin, chrysophanol, physcion, citreosin, emodin-8-*O*-β-D-glucopyranoside, aloesin nepalensides A and B, rumexneposides A and B and (3,5-dimethoxy-4-hydroxyphenol)-1-*O*-β-D-(6-*O*-galloyl)-glucose ect, isolated from *R. nepalensis* and *R. hastatus* was performed against A549, H522 (lung cancer), MCF-7, MCF-10A and SKBR3 cancer cell lines by using the MTT method, with cisplatin as positive control. Compounds exhibited marked activities [64]. A recent study on cytotoxicity of methanolic root extracts of *R. nepalensis* against *Artemia salina* has been reported. It was reported that 1 000 µg/mL concentration of methanolic root extracts showed significant cytotoxic activity against *Artemia salina* and the phytotoxicity activity against *Lemna minor* [56].

#### Anti-inflammatory activities

We can associate inflammation with several acute and chronic diseases which have been a matter of concern for mankind. The ethanolic root extract of *R. nepalensis* showed activity against carrageenan induced rat paw edema, comparative to the standard anti-inflammatory drug diclofenac [69]. Anti-inflammatory activity of chloroform and ethyl acetate root extracts against ear edema was evaluated in a 12-*O*-tetradecanoylphorbol-13-acetate (TPA)-induced acute inflammation mouse model and found significant reduction in ear edema [65]. HPLC analysis of root revealed the presence of nepodin and chrysophanol [71] which showed significant cyclooxygenase inhibitory activity. Aqueous and alcoholic leaf extract was reported to reduce size of the wheal produced by bradykinin, histamine, carbachol and acetylcholine, which indicate that this plant has antibradykinin, antihistaminic and anticholinergic activity [6].

#### Antimicrobial activity

##### Antibacterial activities

In an investigation, methanolic leaf extract of *R. nepalensis* showed potential activity against pathogenic bacterial strains

such as *Escherichia coli*, *Bacillus subtilis*, *Pseudomonas aeruginosa* and *Bacillus cereus* [18]. Antibacterial activities of methanolic root extracts of this plant against *Pseudomonas aeruginosa*, *Salmonella typhi*, *Enterobacter aerogenes*, *Citrobacter freundii* were studied by Hussain *et al.* Highest activity was observed against *E. coli* and *Staphylococcus aureus* [71]. The leaves stem and root extracts were also investigated for their activity against *Proteus vulgaris*, *Salmonella sp.* (MTCC), *Rhodococci sp.*, *Bacillus stearothermophilus* [39], *Streptococcus mutans* [42], *Streptococcus pyogenes* [47] and *S. aureus* [67]. Ghosh *et al.* reported antibacterial property of methanol extract against *Bacillus subtilis*, *S. aureus*, *Vibrio cholerae*, *E. coli* and *Shigella dysenteriae*. Aloe-emodin is reported to possess antibacterial activity [72]. The compounds isolated from *R. nepalensis* were investigated against *Mycobacterium tuberculosis* exhibited potent inhibitory activity. Isoniazid was used as positive control. Moreover, torachryson displayed significant inhibitory activity against the *p*-aminobenzoic acid pathway [63].

##### Antiviral activity

Methanol root extract of *R. nepalensis* exhibited inhibitory action against RNA polymerase of hepatitis C virus. Extracts inhibited HCV-RdRp by 77.9% at a concentration of 50 µg/ml, and extract contained a high percentage of tannin [73]. Leaves showed anti-HIV activity [74]. Extract obtained from the leaves of *R. bequaertii* (syn. *Rumex nepalensis* Spreng.) (SI > 11; EC<sub>50</sub> = 17.7 µg/mL; CC<sub>50</sub> > 200.0; 89% protection against HIV-induced cytopathic effect) [75].

##### Antifungal activity

It is reported that the methanolic leaf extract showed potential activity against pathogenic fungi such as *Candida albicans* [23], *Aspergillus niger*, *Aspergillus flavus* [18]. Methanolic root extract showed higher activity against *Aspergillus niger* and moderate activity against *Aspergillus flavus* and *Alternaria solani* [76]. It is also reported that the ethanolic extracts of root of *R. nepalensis* were significantly active against the fungal pathogens such as, *Avicularia versicolor*, *Fusarium moniliforme*, *Fusarium semitectum*, *Fusarium solani*, *Pythium sp.*, *Rhizopus sp.*, *Sporotrichum sp.*, *Thermomyces sp.* [44].

##### Antidiabetic activity

Yang *et al.* showed that anthraquinone derivatives (chrysophanol, emodin, physcion, nepalenside A and nepalenside B) from *R. nepalensis* could significantly inhibit the secretion of IL-6 at 10 µM concentration. It is investigated that compounds (chrysophanol, emodin, physcion, and nepalenside A) inhibit production of extracellular matrix that could considerably decrease fibronectin and collagen IV production at 10 µM concentration and this concentration is not cytotoxic. This suggests that anthraquinone derivatives are valuable assets to optimize anti-diabetic nephropathy drug [77].

##### Psychopharmacological activity

The methanol extract of *R. nepalensis* was assessed for different psychopharmacological activities in rats and mice. The pharmacological results indicated that the methanol extract of *R. Nepalensis* appears to have an influence on alterations in general behavioural profiles, including alertness, awareness, spontaneous activity, touch, pain and sound responses. The extract significantly potentiated the duration of phenobarbital sodium-induced sleeping time in mice at 200

and 400 mg/kg, suggesting probable tranquillizing and CNS depressant action. Possible effects were examined on other test systems too, e.g. the exploratory behavioural pattern and muscle relaxant activity. Finally, it was concluded that the methanol extract of *R. nepalensis* possessed most of the pharmacological activities characteristic of minor tranquillizers [78].

#### Purgative activity

The methanol extract of *R. nepalensis* root was investigated for its purgative effect in rats. Bisacodyl (3.5 mg/kg) was used as a standard. At oral doses of 100–400 mg/kg, the extract exhibited significant and dose-dependent purgative activity by increasing the intestinal peristalsis and gastrointestinal motility [53].

#### Anti-algal activity

Yi *et al.* screened root extract of *R. nepalensis* for anti-algal activity against the cyanobacterium *Microcystis aeruginosa*. Coexistence culture system assay reported that remarkable inhibition of the algae with inhibitory rate of 24.4% [57]. This data suggest that this plant has significant anti-algal activity.

#### Insecticidal activity

The methanolic root extracts of *R. nepalensis* show significant insecticidal activity against *Sitophilus oryzae*. *R. nepalensis* methanolic extract also showed high mortality rate against *Rhyzopertha dominica*, *Callosobruchus analis* and *Trogoderma granarium* [71].

#### Wound healing activity

Leaf extract of *R. nepalensis* was mixed with Vaseline or butter and applied to the wounds. Antibacterial and antipyretic activity of *R. nepalensis* further justifies its use in traditional medicine to cure wounds [45, 47, 79, 80].

#### Conclusion and Future Perspectives

The aim of this review was to enlighten the valuable application of this unique and valuable plant species. It carries high nutritional and medicinal values for humans and animals. The literature was analyzed to congregate the phytochemical and pharmacological information on *R. nepalensis*, which reaffirmed that this plant is a good source of phytocomplexes and medicinally important pure compounds for treatment of various diseases. *R. nepalensis* demonstrated various medicinal, pharmacological and phytoremediation activities which gives immense importance to this herb. However, further clinical trials should be performed to verify efficacy and any side effects or toxicity of purified plant extracts. It is essential to conduct in-depth and comprehensive pharmacological studies at molecular level to investigate unexploited potential of this plant. For these reasons, wide pharmacological and chemical studies, together with human metabolism, might be the focus of future studies. Besides, the isolation of pure compounds with pharmacological activities and deciphering the underlying mechanisms holds significance in contemporary and future research. This plant could also be improved, through the use of conventional breeding techniques and genetic engineering approaches for metal tolerance, or the metabolism of organic chemicals. Therefore, there is huge room for research in these directions.

#### References

1. Medicinal plants and their importance. [www.cms.herbalgram.org](http://www.cms.herbalgram.org).

2. Food, health, natural medicine, research, uncategorized.30/12/2011. Medicinal plants and their importance as alternative medicines. [www.mdnasirahmed.wordpress.com](http://www.mdnasirahmed.wordpress.com).
3. Shaikh S, Shriram V, Srivastav A, Barve P, Kumar V. A critical review on Nepal Dock (*Rumex nepalensis*): A tropical herb with immense medicinal importance. *Asian Pacific Journal of Tropical Medicine*. 2018; 11(7):405-414.
4. Saleh NAM, El-Hadidi MN, Arafa RFM. Flavonoids and anthraquinones of some Egyptian *Rumex* species (Polygonaceae). *Biochemical Systematics and Ecology*. 1993; 21:301-303.
5. Pardo-de-Santayana M, Tardío J, Morales R. The gathering and consumption of wild edible plants in the Campoo (Cantabria, Spain). *International Journal of Food Science and Nutrition*. 2005; 56:529-542.
6. Giday M, Asfaw Z, Woldu Z. Medicinal plants of the Meinit ethnic group of Ethiopia: An Ethnobotanical study. *Journal of Ethnopharmacology*. 2009; 124:513-521.
7. Cakilcioglu U, Türkoglu I. An ethnobotanical survey of medicinal plants in Sivrice (Elazig-Turkey). *Journal of Ethnopharmacology*. 2010; 132:165-175.
8. Farooq U, Pandith SA, Saggoo MI, Lattoo SK. Altitudinal variability in anthraquinone constituents from novel cytotypes of *Rumex nepalensis* Spreng. A high value medicinal herb of North Western Himalayas. *Industrial Crops and Products*. 2013; 50(10):112-117.
9. Ahmad KS, Kayani WK, Hameed M, Ahmad F, Nawaz T. Floristic diversity and ethnobotany of Senhsa, District Kotli, Azad Jammu & Kashmir (Pakistan). *Pakistan Journal of Botany*. 2012; 44:195-201.
10. Yi YL, Lei Y, Yin YB, Zhang HY, Wang GX. The anti-algal activity of 40 medicinal plants against *Microcystis aeruginosa*. *Journal of Applied Phycology* 2012; 24(4):847-856.
11. Jain A, Jain A. *Tridax procumbens* (L.): A weed with immense medicinal importance: A review. *International Journal of Pharma and Bio Sciences*. 2012; 3(1):544-552.
12. Anusuya NA, Gomathi RA, Manian SE, Sivaram VE, Menon AN. Evaluation of *basella rubra* L., *Rumex nepalensis* spreng and *commelina benghalensis* L. for antioxidant activity. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2012; 4(3):714-720.
13. Kensa M. Floristic study in a Vembanur wetland, Kanyakumari District, Tamilnadu, South India. *Plant Sciences Feed* 2011; 1:194-199.
14. Farooque NA, Majila BS, Kala CP. Indigenous knowledge systems and sustainable management of natural resources in a high altitude society in Kumaun Himalaya, India. *Journal of Human Ecology*. 2004; 16(1):33-42.
15. Gaur RD. Traditional dye yielding plants of Uttarakhand, India. *Natural Product Radiance*. 2008; 7(2):154-165.
16. Liang HX, Dai HQ, Fu HA, Dong XP, Adebayo AH, Zhang LX, *et al.* Bioactive compounds from *Rumex* plants. *Phytochemistry Letters*. 2010; 3(4):181-184.
17. Wahid SF, Osman CP, Ismail NH. Distinguishing isomeric anthraquinone by LC-MS. *Global Journal of Pharmacology*. 2013; 7(4):479-485.
18. Kumar SU, Joseph L, George MA, Bharti VI. Antimicrobial activity of methanolic extracts of *Rumex*

- nepalensis leaves. International Journal of Pharmacy and Pharmaceutical Sciences. 2011; 3(4):240-242.
19. Solanki R, Dalsania S. Evaluation of CNS action of *Rumex nepalensis* Spreng. (Polyginaceae) using mice as experimental animal. International Journal of Research in Pharmaceutical and Biomedical Sciences. 2012; 3:1750-1752.
  20. Aggarwal PK, Kumar LA, Garg SK, Mathur VS. Effect of *Rumex nepalensis* extracts on histamine, acetylcholine, carbachol, bradykinin, and PGs evoked skin reactions in rabbits. Annals of Allergy. 1986; 56(2):177-182.
  21. Kumar S, Joseph L, George M, Kaur L, Bharti V. Skeletal muscle relaxant activity of methanolic extract of *Rumex nepalensis* in albino rats. Journal of Chemical and Pharmaceutical Research. 2011; 3(3):725-728.
  22. Himi H, Iwatsubo Y, Naruhashi N. Chromosome numbers of 11 species in Japanese *Rumex* subg. *Rumex* (Polygonaceae). Journal of Phytogeography and Taxonomy. 1999; 47(2):121-130.
  23. Yadav S, Kumar S, Jain P, Pundir RK, Jadon S, Sharma A, *et al.* Antimicrobial activity of different extracts of roots of *Rumex nepalensis* Spreng. Indian Journal of Natural Product Resource 2011; 2(1):65-69.
  24. Anthony Julian Huxley. The new Royal Horticultural Society dictionary of gardening, Vol 4 Macmillan Press, Cornell University, 1992.
  25. Anjen Li, Alisa E. Grabovskaya-Borodina, Sergei L. Mosyakin, *Rumex Linnaeus*. Flora of china. 2003; 5:333-341.
  26. Vashistha RK, Rawat N, Chaturvedi AK, Nautiyal BP, Prasad P, Nautiyal MC. An exploration on the phenology of different growth forms of an alpine expanse of North-West Himalaya, India. New York Science Journal 2009; 2:29-41.
  27. Shrestha D. Indigenous vegetables of Nepal for biodiversity and food security. Biodiversity and Conservation. 2013; 5(3):98-108.
  28. Yasmin G, Khan MA, Shaheen N, Hayat MQ. Micro morphological investigation of foliar anatomy of *Fagopyrum* Mill and *Rumex* L. of Polygonaceae. Pakistan Journal of Botany. 2010; 42(1):47-57.
  29. Hameed I, Hussain F, Dastagir G. Anatomical studies of some medicinal plants of Family Polygonaceae. Pakistan Journal of Botany. 2010; 42(5):2975-2983.
  30. Hameed I, Hussain F, Dastagir G. Stomatal studies of some selected medicinal plants of Polygonaceae. Pakistan Journal of Botany. 2008; 40(6):2273-2280.
  31. Yasmin G, Khan MA, Shaheen N, Hayat MQ, Ali S, Abbas S. Taxonomic implications of pollen morphology of seven species of *Rumex* L., from Pakistan. Pakistan Journal of Botany. 2010; 42(3):1435-1442.
  32. Bhattarai KR, Vetaas OR, Grytnes JA. Relationship between plant species richness and biomass in arid sub-alpine grassland of the central Himalayas, Nepal. Folia Geobot. 2004; 39(1):57-71.
  33. Kumar N, Vats SK, Kumar S, Ahuja PS. Altitude-related changes in activities of carbon metabolism enzymes in *Rumex nepalensis*. Photosynthetic. 2008; 46(4):611-614.
  34. Chaturvedi AK, Prasad P, Nautiyal MC. Impact of elevated CO<sub>2</sub> on growth, morphology and dry matter partitioning in alpine growth forms of north western Himalayas. Indian Journal of Plant Physiology 2013; 18(2):118-124.
  35. Dreon AL, Paoletti MG. The wild food (plants and insects) in Western Friuli local knowledge (Friuli-Venezia Giulia, North Eastern Italy). Contributions to Natural History. 2009; 12:461-488.
  36. Iqbal I, Hamayun M. Studies on the traditional uses of plants of Malam Jabba valley, District Swat, Pakistan. Ethnobotanical Leaflets. 2004; 1:15.
  37. Kala CP, Farooquee NA, Dhar U. Prioritization of medicinal plants on the basis of available knowledge, existing practices and use value status in Uttaranchal, India. Biodiversity and Conservation. 2004; 13(2):453-469.
  38. Rao KN, Sunitha C, David B, Sandhya S, Mahesh V. A study on the nutraceuticals from the genus *Rumex*. Hygeia Journal of Drug and Medicine. 2011; 3(1):76-88.
  39. Mungole A, Chaturvedi A. Determination of antibacterial activity of two medicinally important Indian Taxa. Der Pharma Chemica. 2011; 3(1):83-89.
  40. Gangwar KK, Deepali GR, Gangwar RS. Ethnomedicinal plant diversity in Kumaun Himalaya of Uttarakhand. Indian National Science Academy. 2010; 8(5):66-78.
  41. Tauchen J, Doskocil I, Caffi C, Lulekal E, Marsik P, Havlik J, *et al.* *In vitro* antioxidant and anti-proliferative activity of Ethiopian medicinal plant extracts. Indian Crops Products. 2015; 74:671-679.
  42. Uniyal SK, Singh KN, Jamwal P, Lal B. Traditional use of medicinal plants among the tribal communities of Chhota Bhangal, Western Himalaya. Journal of Ethnobiology and Ethnomedicine. 2006; 2(1):1.
  43. Weckerle CS, Huber FK, Yongping Y, Weibang S. Plant knowledge of the Shuhi in the Hengduan Mountains, southwest China. Economic Botany. 2006; 60(1):3-23.
  44. Sharma RS, Mishra V, Singh R, Seth N, Babu CR. Antifungal activity of some Himalayan medicinal plants and cultivated ornamental species. Fitoterapia. 2008; 79(7):589-591.
  45. Joshi AR, Joshi K. Documentation of wetland plant diversity with indigenous uses in Nepal: A case study of some wetlands of two Valleys (Kathmandu and Pokhara). Ethnobot. 2009; 21:11-17.
  46. Rana PK, Kumar P, Singhal VK, Rana JC. Uses of local plant biodiversity among the tribal communities of Pangi Valley of district Chamba in cold desert Himalaya, India. Scientific World Journal. 2014; 753289:1-15.
  47. Kothai S, Befirdu G. Ethno botany and antimicrobial activities of medicinal plants used for skin infections in Jimma, Ethiopia. Discovery Pharma. 2012; 2:5-11.
  48. Cheng S. Heavy metal pollution in China: origin, pattern and control. Environmental Science and Pollution Research. 2003; 10(3):192-198.
  49. Vasas A, Orbán-Gyapai O, Hohmann J. The genus *rumex*: review of traditional uses, phytochemistry and pharmacology. Journal of Ethnopharmacology. 2015; 175:198-228.
  50. Rajan S, Sethuraman M, Baburaj DS. Plants from the traditional medical system of the Nilgiri tribes. Ancient Science of Life. 1997; 16(4):360.
  51. Sharma P, Rani S, Ojha SN, Sood SK, Rana JC. Indian herbal medicine as hepatoprotective and hepatocurative: a review of scientific evidence. Life Sciences. 2014; 49:61-115.
  52. Gaire BP, Subedi L. Medicinal plant diversity and their pharmacological aspects of Nepal Himalayas. Pharmacognosy Journal. 2011; 3(25):6-17.

53. Ghosh L, Gayen JR, Murugesan T, Sinha S, Pal M, Saha BP. Evaluation of purgative activity of roots of *Rumex nepalensis*. *Fitoterapia*. 2003; 74(4):372-374.
54. Rawat VS, Jalal JS. Sustainable utilization of medicinal plants by local community of Uttarkashi District of Garhwal, Himalaya, India. *European Journal of Medicinal Plants*. 2011; 1(2):18.
55. Hameed I, Dastagir G, Hussain F. Nutritional and elemental analyses of some selected medicinal plants of the family Polygonaceae. *Pakistan Journal of Botany* 2008; 40(6):2493-2502.
56. Hussain F, Hameed I, Dastagir G, Khan I, Ahmad B. Cytotoxicity and phytotoxicity of some selected medicinal plants of the family Polygonaceae. *African Journal of Biotechnology*. 2010; 9(5):770-774.
57. Kunwar RM, Burlakoti C, Chowdhary CL, Bussmann RW. Medicinal plants in farwest Nepal: Indigenous uses and pharmacological validity. *Medicinal and Aromatic Plant Science and Biotechnology*. 2010; 4(1):28-42.
58. Yineger H, Kelbessa E, Bekele T, Lulekal E. Ethno veterinary medicinal plants at bale mountains national park, Ethiopia. *Journal of Ethnopharmacology* 2007; 112(1):55-70.
59. Singh KN. Traditional knowledge on Ethnobotanical uses of plant biodiversity: a detailed study from the Indian western Himalaya. *Biodiversity Research and Conservation*. 2012; 28(1):63-77.
60. Mei R, Liang H, Wang J, Zeng L, Lu Q, Cheng Y, *et al.* New secoanthraquinone glucosides from *Rumex nepalensis*. *Planta Medica*. 2009; 75(10):1162-1164.
61. Teklehaymanot T. Ethnobotanical study of knowledge and medicinal plants use by the people in Dek Island in Ethiopia. *Journal of Ethnopharmacology*. 2009; 124(1):69-78.
62. Watt JM, Breyer-Brandwijk MG. The medicinal and poisonous plants of Southern Africa. Livingston, Edinburgh, 1932.
63. Liang HX, Dai HQ, Fu HA, Dong XP, Adebayo AH, Zhang LX, *et al.* Bioactive compounds from *Rumex* plants. *Phytochemistry Letters*. 2010; 3:181-184.
64. Gautam R, Karkhile KV, Bhutani KK, Jachak SM. Anti-inflammatory, cyclooxygenase (COX)-2, COX-1 inhibitory and free radical scavenging effects of *Rumex nepalensis*. *Planta Medica*. 2010; 76:1564-1569.
65. Zhang LS, Li Z, Mei RQ, Liu GM, Long CL, *et al.* Hastatusides A and B: two new phenolic glucosides from *Rumex hastatus*. *Helvetica Chimica Acta*. 2009; 92:774-778.
66. Mungole AJ, Chaturvedi A. Determination of antioxidant activity of *Hibiscus sabdariffa* L. and *Rumex nepalensis* Spreng. *International Journal of pharma and bio sciences* 2011; 2(1):120-127.
67. Shrestha R, Timilsina N. Antioxidant and antimicrobial activity and GC-MS analysis of extract of *Rumex nepalensis* Spreng. *The Pharma Innovation*. 2017; 6(9):155-158.
68. Devkota SR, Paudel KR, Sharma K, Baral A, Chhetri SB, Parajuli P, *et al.* Investigation of antioxidant and anti-inflammatory activity of roots of *Rumex nepalensis*. *World Journal of Pharmaceutical Sciences*. 2015; 4(3):582-594.
69. Bhattacharyya P, Kumaria S, Bose B, Paul P, Tandon P. Evaluation of genetic stability and analysis of phytomedicinal potential in micro propagated plants of *Rumex nepalensis*: A medicinally important source of pharmaceutical biomolecules. *Journal of Applied Research on Medicinal and Aromatic Plants*. 2017; 6:80-91.
70. Gautam R, Srivastava A, Jachak SM. Simultaneous determination of naphthalene and anthraquinone derivatives in *Rumex nepalensis* Spreng roots by HPLC: comparison of different extraction methods and validation. *Phytochemical Analysis*. 2011; 22(2):153-157.
71. Hussain F, Ahmad B, Hameed I, Dastagir G, Sanaullah P, Azam S. Antibacterial, antifungal and insecticidal activities of some selected medicinal plants of polygonaceae. *African Journal of Biotechnology*. 2010; 9(31):5032-5036.
72. Ghosh L, Gayen JR, Sinha S, Pal S, Pal M, Saha BP. Antibacterial efficacy of *Rumex nepalensis* Spreng roots. *Phytotherapie Research*. 2003; 17(5):558-559.
73. Jo M, Nakamura N, Kakiuchi N, Komatsu K, Qui MH, Shimotohno K, *et al.* Inhibitory effect of Yunnan traditional medicines on hepatitis C viral polymerase. *Journal of Natural Medicine*. 2006; 60(3):217-224.
74. Maroyi A. Alternative medicines for HIV/AIDS in resource-poor settings: Insight from traditional medicines use in Sub-Saharan Africa. *Tropical Journal of Pharmaceutical Research*. 2014; 13(9):1527-1536.
75. Cos P, Hermans N, De Bruyne T, Apers S, Sindambiwe JB, Witvrouw M, *et al.* Antiviral activity of rwandan medicinal plants against human immunodeficiency virus type-1 (HIV-1). *Phytomedicine*. 2002; 9:62-68.
76. Kumari P, Misra SK, Sharma N. Herbals as antimicrobials: A review. *Journal of Ayurveda Medical Sciences*. 2016; 2(1):31-35.
77. Yang Y, Yan YM, Wei W, Luo J, Zhang LS, Zhou XJ, *et al.* Anthraquinone derivatives from *Rumex* plants and endophytic *Aspergillus fumigatus* and their effects on diabetic nephropathy. *Bioorganic & Medicinal Chemistry Letters*. 2013; 23(13):3905-3909.
78. Ghosh L, Arunachalam G, Murugesan T, Pal M, Saha BP. Studies on the psychopharmacological activities of *Rumex nepalensis* Spreng. Root extract in rats and mice. *Phytomedicine*. 2002; 9(3):202-206.
79. Begum S, Abd ElIslam NM, Adnan M, Tariq A, Yasmin A, Hameed R. Ethno medicines of highly utilized plants in the temperate Himalayan region. *African Journal of Traditional, Complementary and Alternative medicines* 2014; 11(3):132-142.
80. Giday M, Teklehaymanot T, Animut A, Mekonnen Y. Medicinal plants of the Shinasha, Agew-awi and Amhara peoples in northwest Ethiopia. *Journal of Ethnopharmacology*. 2007; 110(3):516-525.