



ISSN (E): 2277- 7695

ISSN (P): 2349-8242

NAAS Rating 2017: 5.03

TPI 2017; 6(6): 213-221

© 2017 TPI

www.thepharmajournal.com

Received: 23-04-2017

Accepted: 25-05-2017

Katakam Prakash

Department of biotechnology,
College of Science & Technology,
Acharya Nagarjuna University,
Guntur, Andhra Pradesh, India

Muralidhar Nama

Department of biotechnology,
College of Science & Technology,
Acharya Nagarjuna University,
Guntur, Andhra Pradesh, India

A PKPD interaction between oral hypoglycemic drug-rosiglitazone and *trigonella foenum-graecum* & histopathological study

Katakam Prakash and Muralidhar Nama

Abstract

Trigonella foenum-graecum (Fenugreek plant) is a well-known traditionally used medicated herb, possesses different therapeutical activities. Fenugreek leaves have been used as traditional herbal medicines not only for hyperglycemia but also in hyperlipidemia, cellulitis and gastrointestinal disorders. Preliminary animal and human trials suggested the possible antihyperglycemic activity and antihyperlipidemic activity of Oral fenugreek leaf extract. *T. foenum-graecum* leaves have also previously been shown to have antihyperglycemic and hypocholesterolemic effects on Type I and Type II Diabetes mellitus patients and experimental induced diabetic animals. However, the research so far on the hypoglycemic effect of fenugreek couldn't establish the optimum dose-level for experimental subjects. Hence, the research studies are required to study the pharmacodynamic and pharmacokinetic properties in order to determine the effect of fenugreek herb on the hyperglycemic patients who are taking the therapy with synthetic drugs. This study was taken up to discover the influence of *Trigonella foenum-graecum* on the pharmacokinetics and pharmacodynamics of Rosiglitazone in rats. Results have proven the negative (decrease) effect of *Trigonella foenum-graecum* on pharmacokinetics but positive (increase) effect on pharmacodynamics of Rosiglitazone.

Keywords: *Trigonella foenum-graecum*, Rosiglitazone, hypoglycemic effect

1. Introduction

Herbs are often administered in combination with therapeutic drugs, raising the potential of herb-drug interactions. An extensive review of the literature identified reported herb-drug interactions with clinical significance, many of which are from case reports and limited clinical observations. Cases have been published reporting enhanced anticoagulation and bleeding when patients on long-term warfarin therapy also took *Salvia miltiorrhiza* (danshen). *Allium sativum* (garlic) decreased the area under the plasma concentration-time curve (AUC) and maximum plasma concentration of saquinavir, but not ritonavir and paracetamol (acetaminophen), in volunteers. *A. sativum* increased the clotting time and international normalized ratio of warfarin and caused hypoglycaemia when taken with chlorpropamide. A typical example is St. John's wort, widely used for depressive disorders, which is a potent inducer of CYP3A4. It is often evident that diabetic patients often consume herbal preparations along with routinely prescribed anti-diabetic agents. The Indian subcontinent has many natural remedies like Ayurveda, Yunani and Siddha. Based on these systems we can able to find new lead molecules upon further research may lead to complete drug. Positive results from clinical trials of these remedies require further investigations along with extensive clinical trials. Most of the plant compounds use as medicine in different diseases is secondary metabolites; they have no role in plant metabolism but has a significant role in defective mechanism of plant. Basic metabolic process of these compounds is almost similar in plants and animals [1-7].

Fenugreek (Scientific name-*Trigonella foenum graecum*) is the medicinal herb belongs to the family Leguminose. This is the common part of man's diet. These fenugreek green leaves and dried seeds are used for preparation of different food items at the same time it is used for medicinal use that is the old therapeutic practice of human's history of medical system. This is used to increase the flavor and colour of food items, and also modifies the quality of food. Fenugreek's seeds have therapeutic applications like anti-hypercholesterolemia, induce lactation, antimicrobial, gastric stimulant, for loss of appetite, anti-hyperglycemic action, galactagogue, hepatoprotective action and antineoplasm. These medicinal applications on physiological actions including the anti-hyperglycemic and antihypercholesterolemic actions of fenugreek

Correspondence Author;

Katakam Prakash

Department of biotechnology,
College of Science & Technology,
Acharya Nagarjuna University,
Guntur, Andhra Pradesh, India

leaves and seeds are mainly attributable to the intrinsic dietary fiber constituents which has been promising the nutraceutical values [8-16].

Rosiglitazone, a member of the thiazolidinedione class of antidiabetic agents, improves glycemic control by improving insulin sensitivity. Rosiglitazone is a highly selective and potent agonist for the Peroxisome proliferator-activated receptor- γ (PPAR γ). In humans, PPAR receptors are found in key target tissues for insulin action such as adipose tissue, skeletal muscle, and liver. Activation of PPAR γ nuclear receptors regulates the transcription of insulin-responsive genes involved in the control of glucose production, transport, and utilization. In addition, PPAR γ -responsive genes also participate in the regulation of fatty acid metabolism. Insulin resistance is a common feature characterizing the pathogenesis of type 2 diabetes. The anti-diabetic activity of rosiglitazone has been demonstrated in animal models of type 2 diabetes in which hyperglycemia and/or impaired glucose tolerance is a consequence of insulin resistance in target tissues. Rosiglitazone reduces blood glucose concentrations and reduces hyperinsulinemia in the ob/ob obese mouse, db/db diabetic mouse, and fa/fa fatty Zucker rat [17-25].

There is scope for the potential herb-interactions between *Trigonella foenum graecum* and Rosiglitazone. This can cause few adverse reactions as a result, it precipitates potentially life-threatening effects. Hence, the study need to be subjected to pharmacological studies in order to discover their effect on the patients who are taking the treatment with synthetic drugs.

2. Materials and Methods

Drugs and Chemicals

Adult male wistar-rats weight between 150 \pm 20grams (Mahavear enterprises Hyderabad, Telangana.) were used in this experimental study. These animals were acclimatized to standard laboratory's conditions of suitable temperature (27 $^{\circ}$ C \pm 1 $^{\circ}$ C) and maintained on 12:12 hours light: dark cycle in animal's house. They were maintained in elevated rat's wire cages and provided with regular rat's chow (Standard pellets contains diet), distilled water *ad-libitum* for 14 days. These Experimental protocols were in conducted according with IAEC/CPCSEA.

Extraction of *T. foenum-graecum* leaves

Collection of Plant material

T. foenum graecum's leaves were collected from the vegetable market of Hyderabad (Telangana). The healthy leaves were washed by using distilled water and the surface water drops were removed by using air drying. The fresh leaves are dried in hot-air oven at 40 $^{\circ}$ C for 48 h and powered and are ready for the extraction process.

Procedure for aqueous extraction

50 g of dried leaf powder of *T. foenum-graecum* is subjected to maceration with the 100 ml sterile distilled H₂O in the blender for 10 minutes. Then the resultant macerate was filtered through the double layered muslin cloth and centrifuged at 4000 rpm for 30 minutes. The supernatant was filter through the Whatmann filter paper No.1 and heat sterilized at 120 $^{\circ}$ C Per 30 minutes. The extract preparation was stored aseptically in the brown colored bottle at 4 $^{\circ}$ C until future use.

Pretreatment

Albino rats were selected for this study (180-250gm), These animals are supplied by the NIN, Hyderabad, Telangana,

animals are maintained under the suitable conditions in animal house. [IAEC number]. The rats are kept in the animal cages and high fatty food and water are supplied in the form of carbohydrates: proteins: fat in 42:18:40 for 14 days.

Induction of Hyperglycemia in Rats by streptozotocin {60mg/kg}

After 15 days of feeding with highly fatty food the rats were fasted for a Period of 18hrs before the induction of hyperglycemia & singledose administration of the 60 mg/kg of Streptozocin (SigmaAldrich; St. Louis; MO; USA) were injected intra-peritonally (freshly dissolve in the normal saline solution). After STZ administration, the animals are free accessed with food (pellet diet) & water. moderate polydipsia and marked polyuria are observed in diabetic hyperglycemic rats. After three days i.e. after 72hrs of injection, fasting blood glucose concentration were determined by following glucose levels by using commercial glucose estimation kits with UV-Visible Spectrophotometer at 505nm based on the oxidase/peroxidase GOD/POD method. If any rats showing the fasting blood glucose level more than 150 mg/dL were consider the hyperglycaemic-rats and selected for the different grouping in the experimental design.

Experimental design: Study design of glibenclamide

The hyperglycemic rats are divided in to 6 groups 6 animals in each.

Group I: Diabetic Control Group (0.5% Sodium. Carboxy Methyl Cellulose Suspension per Oral)

Group II: *T. Foenum-Graecum* (100 Mg/Kg, Per Oral)

Group III: *T. Foenum-Graecum* (500 Mg/Kg. per Oral)

Group IV: Combination of Rosiglitazone (4mg/Kg. per Oral) + *T. Foenum-Graecum* (500 Mg/Kg per Oral).

Group V: Combination of Rosiglitazone (8 mg/Kg. per Oral) + *T. Foenum-Graecum* (500 Mg/Kg per Oral).

Group VI: Rosiglitazone (8 mg/Kg. Per Oral) [26-29].

Pharmacokinetics study in hyperglycemic rat model

Single dose Study

These pharmacokinetic studies are carried out in hyperglycaemic rats (weight b/n 180grams and 250grams). These animals were housed in animal's wire cages with free access to diet and water *ad-libitum*. The overnight fasting rats were dividing in to 6 different groups (n=6) and the follow the treatment was mention in the study design. Blood samples were collected at predetermined intervals of 0hr, 1hr, 2hr, 4hr, 8hr, 12hr and 24hr in the hinto microcentrifugal tubes containing Na⁺ citrate from retro-orbital pucture under di ethyl ether anaesthesia. The blood samples are subject to centrifugation at 3000rpm Per 10minutes and plasma was stored at -20 $^{\circ}$ C for analysis and estimation of kinetic parameters as AUC 0 - ∞ , C_{max} ka, ke CL/F, T_{max}, V/F, AUC 0-t & t_{1/2}.

Multiple dose study

The hyperglycemic rats are dividing into 6 different treatment groups same as mention in study design and daily treatment is carried for 21 days. Samples of blood are collected from different rat's groups on 0th, 7th, 14th, 21st day immediatly after drug treatment. Samples of blood are collected in to microcentrifugal tubes containing Na⁺citrate from retro-orbital puncture under anaesthesia. These blood samples were subjected to centrifuged at 3000rpm Per 10 minuts and plasma was stores at -20 $^{\circ}$ C for analysis and estimation of kinetic parameters as AUC 0 - ∞ , V/F, ka, C_{max}, CL/F, T_{max}, ke, AUC 0-t & t_{1/2}.

Pharmacodynamics study in the hyperglycaemic rats

Single dose study

In this study, treatment was given to all groups of animals as per experimental design. Pharmacodynamic parameters like urea, glucose and cholesterol levels were estimated at the interval of 0, 1, 2, 4, 8, 12 and 24 hours by UV spectrophotometer.

Multiple dose study

In this study, daily treatment was given to all groups of animals for 3 weeks as per experimental design. Pharmacodynamic parameters like urea, cholesterol and glucose levels were estimated at the time interval of 0, 7, 14 and 21 days by UV spectrophotometer.

Statistical Application

ANOVA followed by Dunnett test is performed for comparison between different groups of animals. P value fewer than 5% ($P < 0.05$) was considered statistically significant. All clinical data are expressed in the form of Mean \pm Sd. Pharmacokinetics data was calculated by using pk solver software and statistical analysis and graphical representations were done by *INSTANT graph pad* software.

Histopathological Study

After estimation of last blood glucose level, the animals were sacrificed and histopathological studies to estimate the inflammation and necrosis related changes in pancreas. The pancreatic tissues were stained using H&E stains and observed under resolution 100 \times .

3. Results

Table 1: Blood Glucose levels at 0th, 1st, 2nd, 4th, 8th, 12th and 24th Hour after Oral administration of *T. foenum-graecum*, Saxagliptin and combination of Rosiglitazone + *T. foenum-graecum* in diabetic rats (n=6)

Time (Hours)	Treatment (Single dose study)					
	Diabetic Control	<i>T. foenum-graecum</i> (Dose in mg / kg)		Rosiglitazone (Dose in mg / kg)	Rosiglitazone + <i>T. foenum-graecum</i> (Dose in mg / kg)	
	Vehicle	100	500	8	4 + 500	8 + 500
	Blood glucose level (mg/dl)					
0	404.5 \pm 9.1	415.08 \pm 5.2	382.5 \pm 2.3	402.1 \pm 2.11	419.12 \pm 3.16	388.02 \pm 2.52
1	459.1 \pm 4.18	418.3 \pm 3.16**	352.4 \pm 3.6**	369.8 \pm 3.16**	364.2 \pm 8.3**	359.22 \pm 3.61**
2	453.8 \pm 4.16	355.2 \pm 9.01**	344.3 \pm 5.1**	352.3 \pm 2.18**	349.1 \pm 6.13**	342.15 \pm 4.12**
4	419.16 \pm 5.1	318.13 \pm 10.2**	355.1 \pm 2.1**	348.2 \pm 1.91**	348.1 \pm 5.23**	328.15 \pm 2.91**
8	421.6 \pm 5.3	291.18 \pm 1.8**	268.2 \pm 2.3**	255.18 \pm 4.1**	248.91 \pm 7.13**	239.03 \pm 4.16**
12	414.93 \pm 4.1	316.2 \pm 6.4**	283.3 \pm 5.3**	251.14 \pm 2.95**	235.21 \pm 5.14**	229.16 \pm 4.62**
24	415.8 \pm 6.3	323.8 \pm 3.1**	300.02 \pm 1.5**	258.14 \pm 3.05**	249.02 \pm 10.18**	234.15 \pm 8.13**

Values are given as mean \pm Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

N - Number of animals used.

Table 2: Blood Cholesterol levels at 0th, 1st, 2nd, 4th, 8th, 12th and 24th Hour after Oral administration of *T. foenum-graecum*, Rosiglitazone and combination of Rosiglitazone + *T. foenum-graecum* in diabetic rats (n=6)

Time (Hours)	Treatment (Single dose study)				
	Diabetic Control	<i>T. foenum-graecum</i> (Dose in mg / kg)		Rosiglitazone (Dose in mg / kg)	Rosiglitazone + <i>T. foenum-graecum</i> (Dose in mg / kg)
	Vehicle	100	500	8	4 + 500
	Blood cholesterol level (mg / dl)				
0	196.11 \pm 8.2	209.15 \pm 3.9	208.51 \pm 9.2	210.16 \pm 8.3	199.15 \pm 9.1
1	203.63 \pm 2.6	202.18 \pm 7.0	199.13 \pm 10.1	189.01 \pm 6.3	179.81 \pm 8.2
2	205.33 \pm 8.2	188.6 \pm 1.3**	184.21 \pm 7.2**	179.01 \pm 2.6**	170.9 \pm 3.8**
4	204.42 \pm 6.2	181.6 \pm 1.8**	183.6 \pm 3.5**	166.8 \pm 2.31**	158.9 \pm 2.36**
8	203.51 \pm 3.9	141.8 \pm 3.1**	138.12 \pm 8.1**	139.14 \pm 6.3**	139.13 \pm 6.13**
12	210.9 \pm 6.19	149.15 \pm 3.6**	154.92 \pm 3.5**	138.03 \pm 8.12**	132.16 \pm 5.12**
24	218.88 \pm 3.2	175.8 \pm 4.2**	169.03 \pm 2.2**	145.82 \pm 4.8**	131.26 \pm 4.8**

Values are given as mean \pm Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

N - Number of animals used.

Table 3: Blood Urea levels at 0th, 1st, 2nd, 4th, 8th, 12th and 24th Hour after Oral administration of *T. foenum-graecum*, Rosiglitazone and combination of Rosiglitazone + *T. foenum-graecum* in diabetic rats (n=6)

Time (Hours)	Treatment (Single dose study)					
	Diabetic Control	<i>T. foenum-graecum</i> (Dose in mg/kg)		Rosiglitazone (Dose in mg/kg)	Rosiglitazone + <i>T. foenum-graecum</i> (Dose in mg/kg)	
	Vehicle	100	500	8 + 500	4+500	8 + 500
	Blood urea level (mg/dl)					
0	196.1 \pm 3.0	201.9 \pm 1.9	200.2 \pm 2.08	206.15 \pm 9.1	194.13 \pm 8.13	193.41 \pm 6.26
1	199.8 \pm 3.7	203.8 \pm 6.3	192.16 \pm 11.6	190.15 \pm 1.93	183.26 \pm 4.16	183.15 \pm 8.10
2	203.26 \pm 4.13	181.25 \pm 1.8**	181.28 \pm 4.8**	174.17 \pm 2.31**	171.82 \pm 3.71**	169.02 \pm 3.5**

4	204.91±8.2	176.81±3.1**	172.81±3.4**	164.82±3.83**	153.41±5.6**	154.62±3.34**
8	205.62±4.5	145.61±4.8**	148.92±3.1**	143.42±8.15**	133.18±6.15**	134.15±8.2**
12	205.20±3.3	155.8±4.2**	153.25±6.4**	133.84±5.6**	132.80±3.62**	130.91±3.42**
24	209.03±3.5	179.31±3.5**	167.83±2.03**	146.82±8.1**	133.91±3.05**	134.52±6.3**

Values are given as mean± Standard deviation.

* Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

N - Number of animals used.

Table 4: Blood Glucose levels at 0th, 7th, 14th and 21st day after Oral administration of *T. foenum-graecum*, rosiglitazone and combination of rosiglitazone and *T. foenum-graecum* in diabetic rats (n=6)

Time (Day)	Treatment (Multiple dose study)					
	Diabetic Control	<i>T. foenum-graecum</i> (Dose in mg/kg)		Rosiglitazone (Dose in mg/kg)	Rosiglitazone + <i>T. foenum-graecum</i> (Dose in mg/kg)	
	Vehicle	100	500	8	4 + 500	8 + 500
	Blood glucose level (mg/dl)					
0	404.09±1.9	410.11±3.6	379.09±4.2	398.66±3.15	381.52±6.32	176.21±4.23
7	389.16±1.2	233.4±2.5**	226.81±3.3**	218.38±5.31**	207.82±1.2**	199.1±7.2**
14	380.15±2.15	178.14±2.5**	150.91±1.5**	141.12±7.14**	133.49±4.51**	125.28±7.02**
21	384.16±6.4	124.51±3.2**	124.52±3.6**	113.82±4.2**	108.48±3.69**	98.01±4.2**

Values are given as mean± Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

N - Number of animals used.

Table 5: Blood Cholesterol levels at 0th, 7th, 14th and 21st day after Oral administration of *T. foenum-graecum*, Rosiglitazone and combination of rosiglitazone and *T. foenum-graecum* in diabetic rats (n=6)

Time (Day)	Treatment (Multiple dose study)					
	Diabetic Control	<i>T. foenum-graecum</i> (Dose in mg / kg)		Rosiglitazone (Dose in mg / kg)	Rosiglitazone + <i>T. foenum-graecum</i> (Dose in mg / kg)	
	Vehicle	100	5	8 + 500	4	8 + 500
	Blood cholesterol level (mg / dl)					
0	188.16±3.4	184.21±3.2	172.19±8.2	174.51±4.28	168.91±4.31	166.72±8.13
7	190.56±1.8	101.83±6.3**	98.07±1.5**	99.02±7.03**	91.36±3.2**	88.14±3.25**
14	181.36±3.42	83.49±7.53*	81.35±6.2**	72.16±6.81**	68.14±3.28**	61.25±5.13**
21	188.23±1.92	69.08±1.2**	68.24±3.0**	63.82±4.06**	56.72±6.51**	51.32±3.72**

Values are given as mean ± Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

n - Number of animals used.

Table 6: Blood Urea levels at 0th, 7th, 14th and 21st day after Oral administration of *T. foenum-graecum*, Rosiglitazone and combination of rosiglitazone and *T. foenum-graecum* in diabetic rats (n=6)

Time (Day)	Treatment (Multiple dose study)					
	Diabetic Control	<i>T. foenum-graecum</i> (Dose in mg/kg)		Rosiglitazone (Dose in mg/kg)	Rosiglitazone + <i>T. foenum-graecum</i> (Dose in mg/kg)	
	Vehicle	100	500	8	4 + 500	8 + 500
	BLOOD UREA LEVEL (mg/dl)					
0	68.44±1.5	65.68±2.4	66.71±3.41	63.41±3.8	58.22±1.91	56.72±3.51
7	73.11±6.3	38.15±3.16**	35.62±3.4**	31.25±2.43**	28.91±6.2**	28.62±2.91**
14	76.28±7.13	31.48±2.81**	29.01±5.25**	27.81±2.63**	24.62±8.9**	18.14±3.2**
21	78.01±3.2	28.03±4.3**	28.25±4.6**	23.48±3.1**	19.53±3.4**	16.92±1.33**

Values are given as mean ± Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

n - Number of animals used.

Table 7: Mean plasma rosiglitazone concentrations (µg/ml) (Single dose study)

Time (Hours)	Diabetic Control	Rosiglitazone 8 mg / kg	Rosiglitazone + <i>T. foenum-graecum</i>	
			4 mg / kg + 500 mg / kg	8 mg / kg + 500 mg / kg
1	0	3.19±0.04	2.72±0.04	2.96±0.04
2	0	6.42±0.02	5.14±0.03	6.02±0.03
4	0	6.03±0.04	4.85±0.03	5.32±0.04
8	0	5.42±0.04	4.53±0.03	5.11±0.02
12	0	4.58±0.03	3.51±0.02	4.03±0.02
24	0	2.51±0.02	1.89±0.03	2.19±0.01

Table 8: Effect of *T. foenum-graecum* on Pharmacokinetic parameters of Single dose administration of rosiglitazone in diabetic rats (n = 6)

Pharmacokinetic parameter	Unit for Pharmacokinetic parameter	Rosiglitazone 8 mg / kg	Rosiglitazone + <i>T. foenum-graecum</i>	
			4 mg / kg + 500 mg / kg	8 mg / kg + 500 mg / kg
ka	h ⁻¹	0.8084±0.12	0.7216±0.18*	0.7812±0.16
ke	h ⁻¹	0.7591±0.22	0.7003±0.16	0.7124±0.16
t _{1/2}	h	3.51±0.03	3.19±0.02	3.48±0.06
V/F	(mg / kg) / (µg / ml)	2.15±0.04	2.00±0.01	2.08±0.03**
CL/F	(mg / kg) / (µg / ml) / h	2.80±0.03	2.84±0.04*	2.95±0.03*
Tmax	h	3.08±0.03	2.91±0.02	3.02±0.02*
Cmax	Mg / ml	5.98±0.03	5.01±0.04**	5.38±0.05**
AUC 0-t	Mg / ml*h	224.16±0.4	209.36±0.3**	215.34±0.2**
AUC 0 - ∞	Mg / ml*h	289.03±0.6	232.53±0.2**	248.18±0.4**

Values are given as mean ± Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

*Statistical significance $p < 0.05$ (compared with the control group)

N - Number of animals used.

Table 9: Mean plasma rosiglitazone concentrations (µg/ml) (Multiple dose study)

Time (Days)	Diabetic Control	Rosiglitazone 8 mg / kg	Rosiglitazone + <i>T. foenum-graecum</i>	
			4 mg / kg + 500 mg / kg	8 mg / kg + 500 mg / kg
1	0	3.18±0.03	2.91±0.02	3.02±0.02
7	0	5.38±0.05	4.89±0.03	5.02±0.02
14	0	4.32±0.04	3.81±0.02	4.13±0.03
21	0	3.92±0.04	3.16±0.03	3.63±0.03

Table 10: Effect of *T. foenum-graecum* on Pharmacokinetic parameters of Multiple dose administration of rosiglitazone in diabetic rats (n=6)

Pharmacokinetic parameter	Unit for Pharmacokinetic parameter	Rosiglitazone 8 mg / kg	Rosiglitazone + <i>T. foenum-graecum</i>	
			4 mg / kg + 500 mg / kg	8 mg / kg + 500 mg / kg
ka	h ⁻¹	0.8066±0.09	0.7202±0.21*	0.7801±0.11
ke	h ⁻¹	0.7588±0.31	0.7019±0.13	0.7161±0.24
t _{1/2}	h	3.53±0.01	3.22±0.01	3.41±0.02
V/F	(mg/kg)/(µg/ml)	2.21±0.04	2.01±0.01	2.09±0.03**
CL/F	(mg/kg)/(µg/ml)/h	2.69±0.03	2.72±0.04*	2.86±0.03*
Tmax	h	3.16±0.03	2.81±0.02	3.08±0.02*
Cmax	µg/ml	5.84±0.03	5.12±0.04**	5.28±0.03**
AUC 0-t	µg/ml*h	239.25±0.4	215.51±0.3**	231.18±0.2**
AUC 0 - ∞	µg/ml*h	291.22±0.2	242.44±0.2**	271.22±0.4**

Values are given as mean± Standard deviation.

**Statistical significance $p < 0.01$ (compared with the control group)

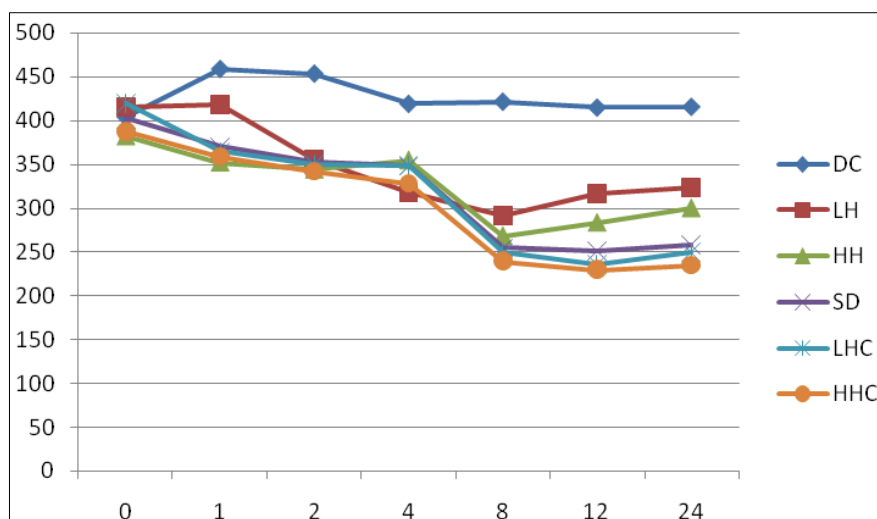
*Statistical significance $p < 0.05$ (compared with the control group)

N - Number of animals used.

4. Discussion

The combination of high dose of rosiglitazone (8 mg / kg) with 500mg/kg *T. foenum-graecum* showed maximum hypoglycemic action, decrease in serum cholesterol and urea

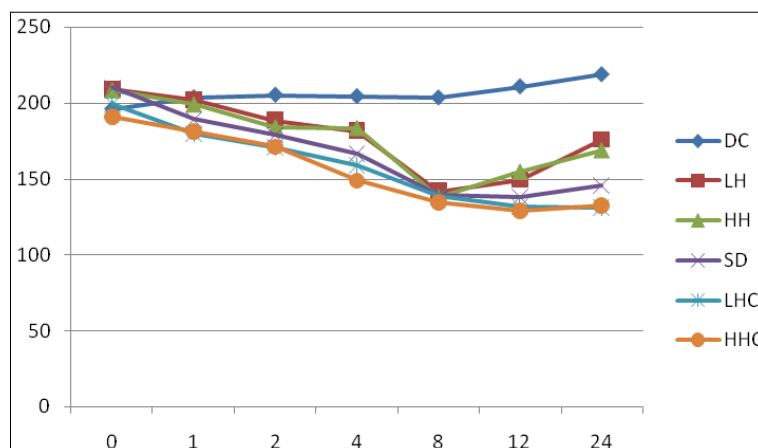
levels. The effect produced by combination of Rosiglitazone (4 mg / kg) with *T. foenum-graecum* was greater than the hypoglycaemic action produced by *T. foenum-graecum* (500 mg / kg) alone and Rosiglitazone (8 mg/kg).



Group 1: Dc, Group II – LH, Group III – HH, Group IV – Sd, Group V – LHC, Group VI – HHC

Figure I: Blood Glucose levels at 0th, 1st, 2nd, 4th, 8th, 12th and 24th Hour after Oral administration of *T. foenum-graecum*,

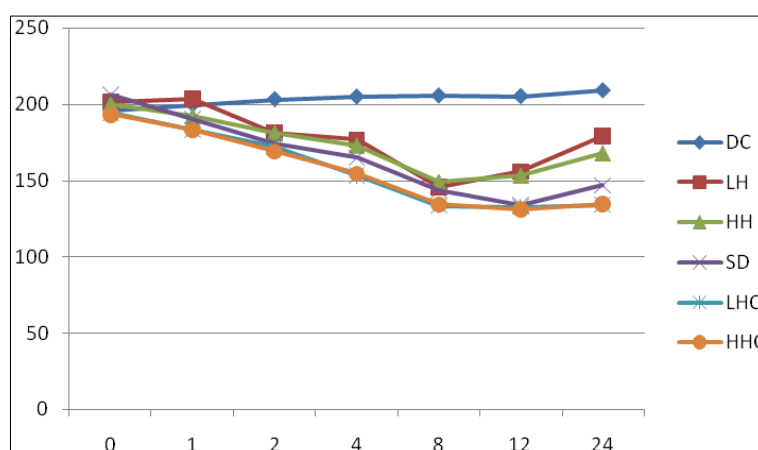
Saxagliptin and combination of Rosiglitazone + *T. foenum-graecum* in diabetic rats (n=6).



Group 2: Dc, Group II - LH, Group III – HH, Group IV – Sd, Group V – LHC, Group VI – HHC

Blood Cholesterol levels at 0th, 1st, 2nd, 4th, 8th, 12th and 24th Hour after Oral administration of *T. foenum-graecum*,

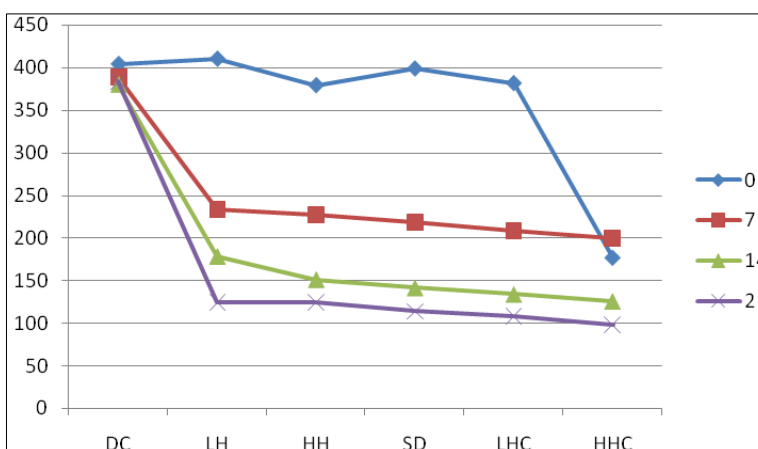
Rosiglitazone and combination of Rosiglitazone + *T. foenum-graecum* in diabetic rats (n = 6).



Group 3: Dc, Group II – LH, Group III – HH, Group IV – Sd, Group V – LHC, Group VI – HHC

Blood Urea levels at 0th, 1st, 2nd, 4th, 8th, 12th and 24th Hour after Oral administration of *T. foenum-graecum*,

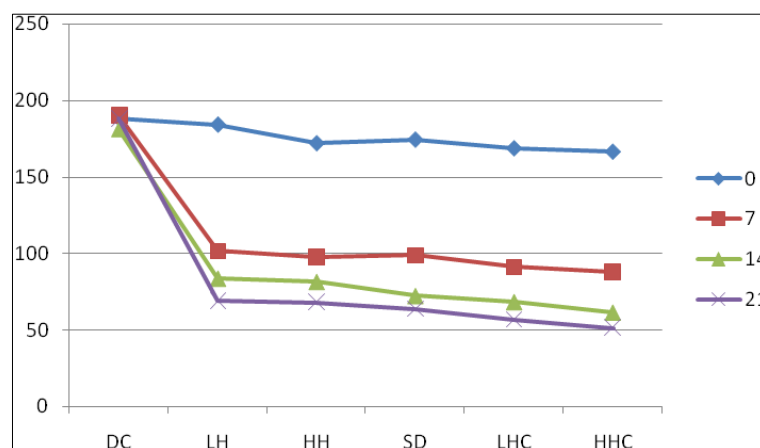
Rosiglitazone and combination of Rosiglitazone + *T. foenum-graecum* in diabetic rats (n=6)



Group3: Dc, Group II – LH, Group III – HH, Group IV – Sd, Group V – LHC, Group VI – HHC

Blood Glucose levels at 0th, 7th, 14th and 21st day after Oral administration of *T. foenum-graecum*, Rosiglitazone and

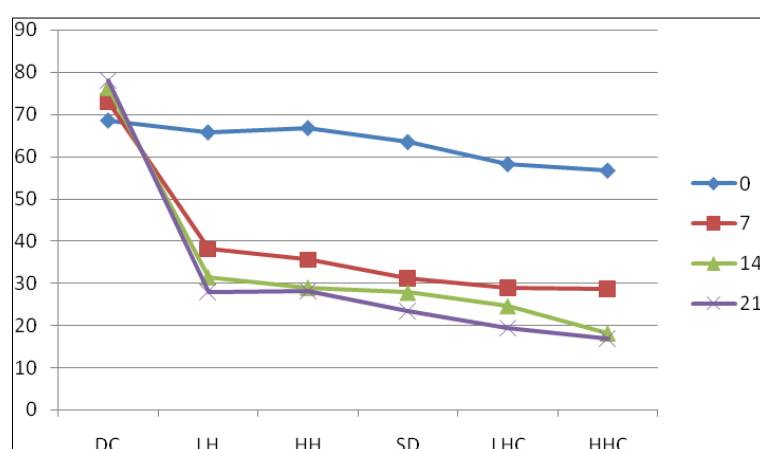
combination of Rosiglitazone and *T. foenum-graecum* in diabetic rats (n=6)



Group 4: Dc, Group II – LH, Group III – HH, Group IV – Sd, Group V – LHC, Group VI – HHC

Blood Cholesterol levels at 0th, 7th, 14th and 21st day after Oral administration of *T. foenum-graecum*, Rosiglitazone and

combination of Rosiglitazone and *T. foenum-graecum* diabetic rats (n=6).



Group 5: Dc, Group II – LH, Group III – HH, Group IV – Sd, Group V – LHC, Group VI – HHC

Blood Urea levels at 0th, 7th, 14th and 21st day after Oral administration of *T. foenum-graecum*, Rosiglitazone and combination of Rosiglitazone and *T. foenum-graecum* in diabetic rats (n=6).

The Single dose study shows that, 80.27% decrease in AUC (0 - ∞) in 500mg/kg of *T. foenum-graecum* and 4 mg/kg of Rosiglitazone, 85.81% decrease AUC (0 - ∞) in 500mg/kg of *T. foenum-graecum* and 8 mg/kg of Rosiglitazone when compared with the 8mg/kg of Rosiglitazone group.

C max was decreased by 83.77% in 500mg/kg of *Trigonella foenum-graecum* and 4mg/kg of Rosiglitazone, 89.96% in 500mg/kg of *T. foenum-graecum* and 8mg/kg of Rosiglitazone in single dose study when compared with the 8mg/kg of Rosiglitazone group.

Significant decrease in absorption rate constant Ka by about 89.26% in Lower dose of 500mg/kg of *T. foenum-graecum* and 4mg/kg of Rosiglitazone, 96.63% in 500mg/kg of *T. foenum-graecum* and 8mg/kg of Rosiglitazone when compared with the 8 mg/kg of Rosiglitazone group.

Significantly increase in clearance 1.42% in 500mg/kg of *T. foenum-graecum* and 4 mg/kg of Rosiglitazone. 5.35% in 500mg/kg of *T. foenum-graecum* and 8 mg/kg of Rosiglitazone compared to 8 mg/kg Rosiglitazone when compared with the 8 mg/kg of Rosiglitazone group.

The multiple dose study shows that, 83.16% decrease in AUC (0 - ∞) in 500mg/kg of *T. foenum-graecum* and 4 mg/kg of Rosiglitazone. 93.12% decrease AUC (0 - ∞) in 500mg/kg of

T. foenum-graecum and 8 mg/kg of Rosiglitazone when compared with the 8 mg/kg of Rosiglitazone group.

C max was decreased by 87.67% in 500mg/kg of *T. foenum-graecum* and 4 mg/kg of Rosiglitazone, 90.41% in 500mg/kg of *T. foenum-graecum* and 8 mg/kg of Rosiglitazone in multiple dose study when compared with the 8 mg/kg of Rosiglitazone group.

Significant decrease in absorption rate constant Ka by about 89.28% in Lower dose of 500mg/kg of *T. foenum-graecum* and 4 mg/kg of Rosiglitazone, 96.71% in 500mg/kg of *T. foenum-graecum* and 8 mg/kg of Rosiglitazone when compared with the 8 mg/kg of Rosiglitazone group.

Significantly increase in clearance 1.01% in 500mg/kg of *T. foenum-graecum* and 4 mg/kg of Rosiglitazone. 1.06% in 500mg/kg of *T. foenum-graecum* and 8 mg/kg of Rosiglitazone compared to 8 mg/kg Rosiglitazone group.

The exact reason behind the reduction in pharmacokinetic parameters was unknown but, it was understood that the combination of Rosiglitazone extract with Rosiglitazone in fact reduces exposure of the synergic drugs without reducing the pharmacodynamic activity. The proposed combination allows a safe therapy with less adverse effects.

Histological study

The histological study shows that the combination therapy (Rosiglitazone + *T. foenum-graecum*) involved in the increase the number of islets and recovered the partially damaged B

cells in pancreas when compare to the Individual treatment.

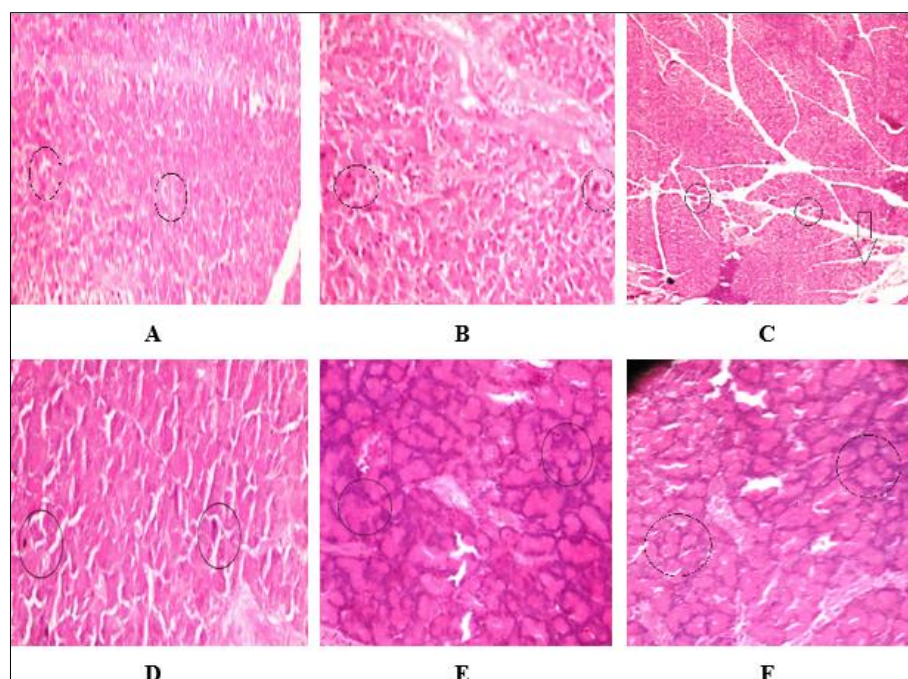


Fig 1: H&S Staining of Pancreatic islets of Diabetic Control, *T. foenum-graecum* alone Rosiglitazone alone and combination of *T. foenum-graecum* & Rosiglitazone treated Diabetic Rats. 6A. diabetic control, 6B. 100mg of *T. foenum-graecum* 6C. 500mg *T. foenum-graecum*, 6D. 8mg of Rosiglitazone. 6E. 500mg of *T. foenum-graecum* + 4mg of Rosiglitazone, 6F. 500mg of *T. foenum-graecum* + 8mg of Rosiglitazone.

Slide A shows that pancreatic cells were damaged due to development of diabetes from STZ. Figure 6B shows that few pancreatic cells were damaged due to Rosiglitazone. Figures 6C, 6D, 6E, 6F shows that B cells are regenerated in pancreatic tissue.

Normal β -cells were observed in low and high doses of Rosiglitazone and *T. foenum-graecum*. (Slides: 6D&6F). In the Rosiglitazone group more damaged β -cells as compared with the 500mg of *T. foenum-graecum* + 8mg of Rosiglitazone and 500mg of *T. foenum-graecum* + 4mg of Rosiglitazone (Figures: 6B, 6C & 6E).

Histopathological studies revealed that the volume of islet cells in pancreas was significantly more in drug treated animals compared to the Diabetic control. The islet cells were shrunk and lytic cellular changes were observed in Diabetic control, Individual treatment had improved it but combination groups with a higher dose of Rosiglitazone showed the return of islets close to original cytoarchitecture. In combination group, islets were big and cells were clear with good vascular pattern. The results of combination group with a high dose of Rosiglitazone produced increment to the volume of islets in pancreas compared to individual treatment. In this study, *T. foenum-graecum* was decrease the absorption and increase the clearance of Rosiglitazone. Hence care must be taken when the combination is taken by diabetic patients.

5. Conclusion

The interaction appears to be pharmacokinetic interaction at absorption, elimination. *T. foenum-graecum* inhibits the absorption of Rosiglitazone that results in a significant decrease in the bioavailability of the later and combination group with a lower dose of Rosiglitazone and increment to the volume of islets in pancreas is observed in combination group when compare to individual treatment. Since the interaction was seen in rats it is likely to occur in humans leading to decreased activity of Rosiglitazone that can need dose

adjustments. Hence care must be taken when the combination is taken by the diabetic patients. The present study warrants next plan to find out the relevance of the interaction in human beings.

6. References

1. Thomas NJ, Lynam AL, Hill AV, *et al.* Type 1 diabetes defined by severe insulin deficiency occurs after 30 years of age and is commonly treated as type 2 diabetes. *Diabetologia*. 2012;62:1167-1172.
2. Gonzalez A, Deng Y, Lane AN, *et al.* Impact of mismatches in HbA1c vs glucose values on the diagnostic classification of diabetes and prediabetes. *Diabet Med*. 2016;37:689-696.
3. Frommer L, Kahaly GJ. Autoimmune polyendocrinopathy. *J Clin Endocrinol Metab*. 2019;104:4769-4782.
4. Smith CJ, Almodallal Y, Jatoi A. Rare adverse events with programmed death-1 and programmed death-1 ligand inhibitors: justification and rationale for a systematic review. *Curr Oncol Rep*. 2017;23:86-69.
5. Zhao Z, Wang X, Bao X-Q, Ning J, Shang M, Zhang D. Autoimmune polyendocrine syndrome induced by immune checkpoint inhibitors: a systematic review. *Cancer Immunol Immunother*. 2015;70:1527-1540.
6. Ziegler A-G, Kick K, Bonifacio E, *et al.*; Fr1da Study Group. Yield of a public health screening of children for islet autoantibodies in Bavaria, Germany. *JAMA* 2013;323:339-351, 74.
7. McQueen RB, Geno Rasmussen C, Waugh K, *et al.* Cost and cost-effectiveness of large-scale screening for type 1 diabetes in Colorado. *Diabetes Care*. 2012;43:1496-1503.
8. Saccone G, Khalifeh A, Al-Kouatly HB, Sendek K, Berghella V. Screening for gestational diabetes mellitus: one step versus two step approach. A meta-analysis of randomized trials. *J Matern Fetal Neonatal Med*.

- 2015;33:1616-1624.
9. Anarthe SJ, Sunitha D, Sandhya R, Ganga M. Immunomodulatory activity for methanolic extract of *Trigonella foenum graecum* whole plant in Wistar Albino rats. *Am J Phytomed Clin Ther.* 2013;2:1081-092.
10. Guardiola FA, Bahi A, Esteban MA. Effects of dietary administration of fenugreek seeds on metabolic parameters and immune status of gilthead seabream (*Sparus aurata* L.). *Fish Shellfish Immunol.* 2012;74:372-9. doi: 10.1016/j.fsi.2018.01.010.
11. Ameen SM, Mosaad GMM, Hussein MK. Effect of some medicinal plants as feed additives on growth Performance, blood constituents and carcass characteristics of broilers. *J Adv Vet Res.* 2015;9:170-7.
12. Laudadio V, Nasiri-Dehbaneh M, Bilal RM, Qotbi A, Javandel F, Ebrahimi A, *et al.* Effects of different levels of dietary black cumin (*Nigella sativa* L.) and fenugreek (*Trigonella foenum-graecum* L.) and their combination on productive traits, selected blood constituents, microbiota and immunity of broilers. *Anim Biotechnol.* 2015;16:1-14. doi: 10.1080/10495398.2020.1853138
13. Al-Homidan IH, Ebeid TA, Al-Muzaini A, Abou-Emera OK, Fathi MM. Impact of dietary fenugreek, mung bean, and garden cress on growth Performance, carcass traits, blood measurements, and immune response in broiler chickens. *Livestock Sci.* 2015;242:104318. doi: 10.1016/j.livsci.2020.104318
14. Liu XD, Song J, Liu X, Shan H. Research note: circular RNA expressing in different developmental stages of the chicken bursa of fabricius. *Poult Sci.* 2016;99:3846-52. doi: 10.1016/j.psj.2020.04.026.
15. Adeyemi K, Sola-Ojo F, Ishola J, Ahmed M, Lawal M. Influence of *Anacardium occidentale* leaf supplementation in broiler chicken diet on Performance, caecal microbiota, blood chemistry, immune status, carcass, and meat quality. *Br Poult Sci.* 2018;62:552-61. doi: 10.1080/00071668.2021.1894321
16. Shi H, Luo Y, Li Y, Zhang F, Liu N. Tetramethylpyrazine supplementation improves Performance, digestion, blood and immune state of broilers exposure to oxidative stress. *J Anim Physiol Anim Nutr.* 2015;106:132-8. doi: 10.1111/jpn.13566.
17. Ji W, Peng X, Lou T, Wang J, Qiu W. Total flavonoids from *Tetrastigma hemsleyanum* ameliorates inflammatory stress in concanavalin A-induced autoimmune hepatitis mice by regulating Treg/Th17 immune homeostasis. *Inflammopharmacology.* 2016;27:1297-307. doi: 10.1007/s10787-019-00599-0.
18. Karova K, Wainwright JV, Machova-Urdzikova L, *et al.*, "Transplantation of neural precursors generated from spinal progenitor cells reduces inflammation in spinal cord injury via NF- κ B pathway inhibition," *Journal of Neuroinflammation.* 2018;16(1):12.
19. Noori L, Arabzadeh S, Mohamadi Y, *et al.*, Intrathecal administration of the extracellular vesicles derived from human Wharton's jelly stem cells inhibit inflammation and attenuate the activity of inflammasome complexes after spinal cord injury in rats," *Neuroscience research.* 2016;170:87-98.
20. Younsi G Zheng, Scherer M, *et al.*, Three growth factors induce proliferation and differentiation of neural precursor cells *in vitro* and support cell-transplantation after spinal cord injury *in vivo*, *Stem Cells International,* 2015;2020:15.
21. Wang D, Wang K, Liu Z, Wang Z, Wu H. Valproic acid labeled chitosan nanoparticles promote the proliferation and differentiation of neural stem cells after spinal cord injury," *Neurotoxicity Research.* 2014;39(2):456-466.
22. Chung MM, Nicol CJ, Cheng YC, *et al.*, Metformin activation of AMPK suppresses AGE-induced inflammatory response in hNSCs, *Experimental Cell Research.* 2017;352(1):75-83.
23. Lin CH, Cheng YC, Nicol CJ, Lin KH, Yen CH, Chiang MC. Activation of AMPK is neuroprotective in the oxidative stress by advanced glycosylation end products in human neural stem cells, *Experimental Cell Research,* 2017;359(2):367-373.
24. Guo Y, Wang F, Li H, *et al.*, Metformin protects against spinal cord injury by regulating autophagy via the mTOR signaling pathway," *Neurochemical Research.* 2018;43(5):1111-1117.
25. Wang P, Xie ZD, Xie CN, *et al.*, AMP-activated protein kinase-dependent induction of autophagy by erythropoietin protects against spinal cord injury in rats, *CNS Neuroscience & Therapeutics.* 2018;24(12):1185-1195.
26. Pickles S, Vigie P, Youle RJ. Mitophagy and quality control mechanisms in mitochondrial maintenance," *Current biology: CB.* 2018;28(4):R170-R185.
27. Zhang X, Wang X, Zhao M. Influence of andrographolide on the pharmacokinetics of warfarin in rats. *Pharm Biol.* 2018;56(1):351-6. doi: 10.1080/13880209.2018.1478431.
28. Rombolà L, Scuteri D, Marilisa S, Watanabe C, Morrone LA, Bagetta G, *et al.* Pharmacokinetic interactions between herbal medicines and drugs: their mechanisms and clinical relevance. *Life (Basel).* 2020;10(7):106. doi: 10.3390/life10070106.
29. Niu J, Straubinger RM, Mager DE. Pharmacodynamic drugdrug interactions. *Clin Pharmacol Ther.* 2019;105(6):1395- 406. doi: 10.1002/cpt.1434.
30. Zou W, Xiao Z, Wen X, Luo J, Chen S, Cheng Z, *et al.* The anti-inflammatory effect of *Andrographis paniculata* (Burm. f.) Nees on pelvic inflammatory disease in rats through down-regulation of the NF- κ B pathway. *BMC Complement Altern Med.* 2016;16(1):483. doi: 10.1186/s12906-016-1466-5