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## Cytotoxic and thrombolytic activity of roots of *Musa paradisiaca* (Linn)

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**Abstract**

The study was aimed to evaluate the cytotoxic and thrombolytic activity of methanol extract of *Musa paradisiaca*. The cytotoxic activity of the crude extract was determined by using brine shrimp lethality bioassay and LC<sub>50</sub> values of the sample were 22.336 ± 0.41 µg/ml whereas for standard vincristine sulfate was 8.50 ± 0.16 µg/ml as a positive control. In thrombolytic activity using *in vitro* clot lysis method, the plant's extract showed (46.26 ± 1.54%) clot lysis as compared to standard streptokinase (67.32 ± 0.34%).

**Keywords:** *Musa paradisiaca*, cytotoxic, thrombolytic activity, methanol extract

**1. Introduction**

Plant-based foods contain significant amounts of bioactive compounds, which provide desirable health benefits beyond basic nutrition. Epidemiological evidence suggests that consumption of a diet rich in vegetables and fruits has positive implications for human health. The World Health Organization reported that 80 % of the world populations rely chiefly on indigenous medicine and that the majority of traditional therapies involve the use of plant extracts or of their active constituents<sup>[1]</sup> and over 25% of modern medicines that are commonly used worldwide contains compounds extracted from medicinal plants<sup>[2]</sup>. In Bangladesh, there is abundant of medicinal plants and ninety percent of the medicinal plants are wild sourced<sup>[3, 4]</sup>.

During recent decades, there has been an increasing demand for finding newer and safer chemotherapeutic agents. Cancer is the third leading cause of death worldwide, only preceded by cardiovascular disease, infectious and parasitic disease<sup>[5]</sup>. Extracts of medicinal plants are believed to contain a wide spectrum of polyphenolic, flavonoids, alkaloids, terpenoids and saponin compounds, which might have therapeutic properties and hinder cancer formation<sup>[6]</sup>. Over 60% of current cytotoxic agents have been derived from natural sources including plants, marine organisms, and microorganisms, either directly or by chemical synthesis based on natural lead compounds<sup>[7, 8]</sup>. Therefore, natural products have a wide application in cancer chemotherapy<sup>[8]</sup>.

Cardiovascular disease caused by blood clot (thrombus) formation is one among the most severe diseases which are increasing at an alarming rate in the recent years<sup>[9]</sup>. Homeostasis maintains the integrity of circulatory system after damaging of the vascular channel<sup>[10]</sup>. Thrombus development is a critical event in the arterial diseases associated with myocardial infarction, anoxia, hypertension<sup>[11]</sup>, stroke, reduction of the blood supply to the liver<sup>[12]</sup> and venous thromboembolic disorders that account for a considerable number of deaths worldwide<sup>[13]</sup>. Remarkable efforts have been made towards the discovery and development of natural constituents from various plant and animal sources which have antiplatelet<sup>[14, 15]</sup> anticoagulant<sup>[16, 17]</sup>, antithrombotic<sup>[18]</sup> and thrombolytic activity<sup>[19-21]</sup>.

Thrombolytic agents are used to dissolving clot and in the management of thrombosis in patients<sup>[22]</sup>. Thrombolytic agents such as tissue plasminogen activator (t-PA), Urokinase (UK), streptokinase (SK)<sup>[23]</sup> etc. are used all over the world for the treatment<sup>[24]</sup> but their use is associated with hyper risk of haemorrhage<sup>[25]</sup>, anaphylactic reaction and lacks specificity<sup>[10, 25]</sup>. Because of the shortcomings in the existing thrombolytic agents, a number of researches are underway to improve the variants of these drugs for their better effective nature<sup>[26]</sup>.

*Musa paradisiaca* (Linn) commonly known as Banana belonging to the family Musaceae [27] is an indigenous plant to Bangladesh. Traditionally the plant was used for different purposes such as abscess, alopecia (female), anasarca, burns, cancer, cataplasm, diabetes, diarrhea, dog bites, dysentery, dyspepsia, corruptions, fractures, gangrene, headache, hematuria, hemiplegia, hemoptysis, hemorrhage, hypertension, lizard bites, mange, marasmus, migraine, nausea, otalgia, psoriasis, ringworm, scorpion sting, septicemia, shingles, smallpox, snake bite, sore, strain, syphilis, tuberculosis, warts, and wound [28, 29]. Pharmacological investigations revealed that banana fruits, Stem juice, flowers are screened for analgesics activity [30], hair growth promoting activity [31], anticonvulsant activity [32], antimicrobial activity [33-38]. The present study was undertaken to investigate the cytotoxic and thrombolytic activity of methanol extract of the roots of this plant.

## 2. Materials and methods

### 2.1 Chemicals

Lyophilized streptokinase vial (1500000 I.U.) was purchased from Square Pharmaceuticals Ltd, Bangladesh. Methanol was purchased from Merck, Germany. Normal saline solution was purchased from Beximco Infusion Ltd. All chemicals used were of analytical reagent grade.

### 2.2 Plant materials

Fresh roots of *Musa paradisiaca* for this study were collected from the local area of Chittagong, Bangladesh and were authenticated by Dr. Sheikh Bokhtear Uddin, Associate Professor, Department of Botany, University of Chittagong, Chittagong-4331, Bangladesh.

### 2.3 Preparation of crude extract

The collected roots were dried for a period of 2 weeks under shade and ground. The ground roots (750 gm) were soaked in sufficient amount of methanol for one week at room temperature with occasional shaking and stirring. The sediments were filtered and the filtrates were dried at 40 °C in a water bath. The solvent was completely removed by filtering with whatman number-1 filter paper. The solvent was evaporated under reduced pressure at room temperature to yield semisolid. The extract was then preserved in a refrigerator till further use [39].

### 2.5 Thrombolytic test

This test was performed according to the method described by Prasad *et al.*, 2006 [40]. In the commercially available lyophilized streptokinase vial (1500000 I.U.) 5 mL sterile distilled water was added and mixed properly. This suspension was used as a stock solution from which appropriate dilution was made. Five milliliters of venous blood was drawn from the healthy volunteers (n=10) without the history of oral contraceptive or anticoagulant therapy and was distributed (0.5 mL/tube) to each ten previously weighed sterile microcentrifuge tube and incubated at 37 °C for 45 min to form the clot. After the clot formation, serum was completely removed without disturbing the clot and each tube having clot was again weighed to determine the clot weight. A volume of 100 µL of methanol extract (10 mg/ mL) was added to each microcentrifuge tube containing pre-weighed clot. As a positive control, 100 µL of streptokinase and as a negative control 100 µL of distilled water were separately added to the control tube numbered. All the tubes were then incubated at 37 °C for 90 min and observed for clot lysis. After incubation,

fluid released was removed and tubes were again weighed to observe the difference in weight after clot disruption. The difference obtained in weight taken before and after clot lysis was expressed as a percentage of clot lysis [41].

### 2.6 Brine shrimp lethality assay

The assay was carried out according to the principle and protocol previously described by [42-44], with slight modifications. Here simple zoological organism (*Artemia salina*) was used as a convenient monitor for the screening. Dried cysts of *Artemia salina* were collected from an aquarium shop (Chittagong, Bangladesh) and hatched in artificial seawater (3.8% NaCl solution) for 48 h to mature shrimp called nauplii. After hatching, active nauplii free from egg shells were collected from a brighter portion of the hatching chamber and used for the assay.

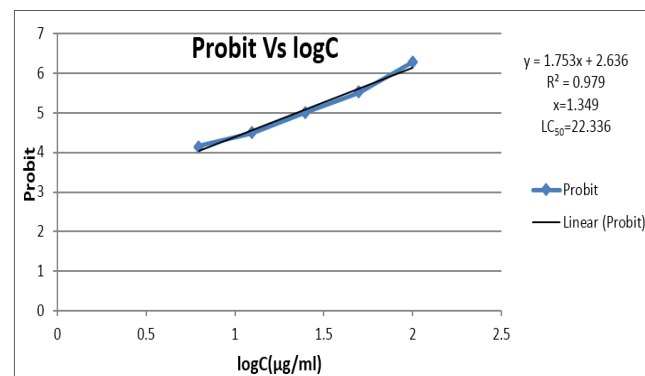
The test sample (extract) were prepared by dissolving them in DMSO (not more than 50 µL of 5 mL solutions) plus sea water (3.8% NaCl in water) to attain concentrations of 6.25, 12.5, 25, 50, 100, 300 and 500 µg/ml. A vial containing 50 µL DMSO diluted to 5 mL was used as a control. Vincristine sulfate [45] was used as positive control. After 24 hours the number of survival of nauplii was counted and percentage of mortality was determined using the equation:

$$\% \text{ mortality} = (\text{no. of dead nauplii} / \text{initial no. of live nauplii}) \times 100.$$

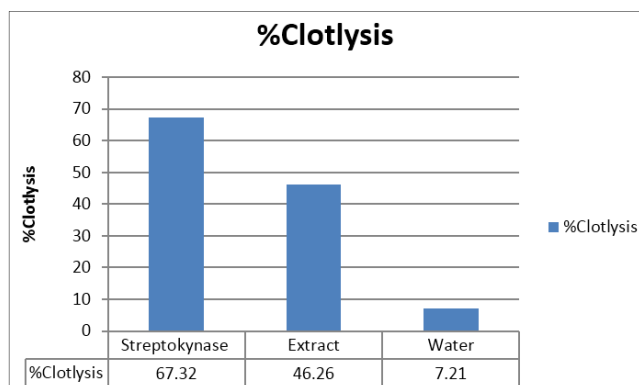
The statistical method of probit analysis (Finney's table) [46] was used to calculate LC<sub>50</sub>. The criterion of toxicity for fractions was established according to (Déciga-Campos *et al.*, 2007) [47]. LC<sub>50</sub> values > 1000 µg/mL (non-toxic), ≥ 500 ≤ 1000 µg/mL (weak toxicity) and < 500 µg/mL (toxic).

## 3. Results

The lethality of the crude extract of *Musa paradisiaca* root to brine shrimp was determined on *Artemia salina* after 24 h of exposure the samples, the negative control DMSO and sea water and vincristine sulfate used as a standard. This technique was applied for the determination of a general toxic property of the plant extract. The LC<sub>50</sub> value (Figure 1) of the extract was 22.336 µg/mL and that for standard vincristine sulfate was 8.50 µg/mL. No mortality was found in the control group, using DMSO and sea water. The plant extract showed moderate clot lysis activity (46.26%) as compared to standard streptokinase clot lysis (67.32%) activity (Figure 2).



**Fig 1:** Toxicity assay of *Musa paradisiaca* on brine shrimp. The results are expressed as Mean±SEM of three measurements.



**Fig 2:** The clot lysis activity of *Musa paradisiaca* extract and streptokinase. All results are expressed as Mean $\pm$ SEM of three consecutive experiments.

#### Statistical analysis

All the results obtained by *in vitro* experiment were expressed as Mean $\pm$ SEM of three measurements followed by Dunnet's test where  $P < 0.01$  was considered as statistically significant.

#### 4. Discussion

Most thrombolytic agents work by activating the enzyme plasminogen, which clears the cross-linked fibrin mesh. This makes the clot soluble and subject to further proteolysis by other enzymes and restores blood flow over occluded blood vessels. Thus thrombolytic agents are useful for the treatment of myocardial infarction, thromboembolic strokes, deep vein thrombosis and pulmonary embolism to clear a blocked artery and avoid permanent damage to the perfused tissue (e.g. myocardium, brain, and leg).

Ideally, any agent useful in the treatment of cancer should not be toxic to the normal cell. However, in reality, anticancer agents are often toxic to normal cells, particularly towards rapidly growing cells<sup>[48]</sup>. It is necessary to test this extract in low concentration to evaluate its potency and also against various cancer cell lines as well as normal cell line so justify the potential to further investigate this plant for anticancer activity.

Hence this study was conducted by crude root extract of *Musa paradisiaca*, further advanced studies should be carried out for compound isolation and it is necessary to observe which compounds are actually responsible for the specific effect.

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#### 6. Reference

- World Health Organization. Summary of WHO guidelines for the assessment of herbal medicines. *Herbal Gram*. 1993; 28:13-14.
- Robbers JE, Speedle MK, Tyler VE. *Pharmacognosy and Pharmacobiotechnology*. Williams and Wilkins, Baltimore, USA, 1996.
- Ghani A. *Medicinal Plants of Bangladesh: Chemical Constituents and Uses*. Asiatic Society of Bangladesh, Dhaka, 1998.
- South Asia Enterprise Development Facility (SEDF) & Intercooperation (IC). *Medicinal Plants Marketing in*

Bangladesh. A market study report. SEDF-Intercooperation, Dhaka, 2003.

- Mia MMK. *Traditional Medicines of Bangladesh*. In *Traditional medicines* (ed. Ghani A). Jahangirnagar University, Dhaka, 1990.
- Rojas A, Hernandez L, Rogeho PM, Mata R. Screening for antimicrobial activity of crude drug extracts and pure natural products from Mexican medicinal plants. *J. Ethnopharmacol*. 1992; 35:127-149.
- Zima TS, Fialova L, Mestek O, Janebova M, Crkovska J, Malbohan I *et al*. Oxidative stress, metabolism of ethanol and alcohol-related diseases. *Journal of Biomedical Science*. 2001; 8:59-70.
- Halliwell B, Gutteridge JMC. *Free radicals in biology and medicine*. Ed 3, Oxford University Press, Oxford, 1999, 415-421.
- Stadtman ER. Role of oxidant species in aging. *Current Medicinal Chemistry*. 2004; 11:1105-1112.
- Ames BN, Shigenaga MK, Hagen TM. Oxidants, antioxidants and the degenerative diseases of aging. *Proceedings of the National Academy of Sciences of the United States of America*. 1993; 90(17):7915-7922.
- Halliwell B, Gutteridge JMC. *Free Radicals in Biology and Medicine*, 2nd Edition, Clarendon Press, Oxford, 1989.
- Halliwell B. How to Characterize a Biological Antioxidant, *Free Radical Research Communications*. 1990; 9(1):1-32. doi:10.3109/10715769009148569
- Valko M, Leibfritz D, Moncola J, Cronin MTD, Mazura M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *International Journal Biochemistry Cell Biology*. 2007; 39:44-84.
- Wolfe KWX, Liu RH. Antioxidant activity of apple peels. *Journal of Agricultural and Food Chemistry*. 2003; 51(3):609-614.
- Suhaj M. Spice antioxidants isolation and their antiradical activity: A review. *Journal of Food Composition and Analysis*. 2006; 19:531-537.
- Sun T, Ho CT. Antioxidant activities of buckwheat extracts. *Food Chemistry*. 2005; 90:743-749.
- Hinneburg I, Damien Dorman HJ, Hiltunen R. Antioxidant activities of extracts from selected culinary herbs and spices. *Food Chemistry*. 2006; 97:122-129.
- Kumar GS, Nayaka H, Dharmesh SM, Salimath PV. Free and bound phenolic antioxidants in amla (*Embllica officinalis*) and turmeric (*Curcuma longa*). *Journal of Food Composition*. 2006; 19:446-452.
- Cousins M, Adelberg J, Chen F, Rieck J. Antioxidant capacity of fresh and dried rhizomes from four clones of turmeric (*Curcuma longa* L.) grown *in vitro*. *Industrial Crops and Products*. 2007; 25:129-135.
- Mathers CD, Boschi-Pinto C, Lopez AD, Murray CJL. Cancer incidence, mortality and survival by site for 14 regions of the world. World Health Organization, 2001, 3.
- Dia J, Mumper RJ. Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*. 2010; 15:7313-7352.
- Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981-2002. *J Nat Prod*. 2003; 66:1022-1037.
- Cragg GM, Kingston DGI, Newman DJ. *Anticancer Agents from Natural Products*. Boca Raton FL. CRC Press, 2005.
- Sai Sandeep Y, Mamatapanigrahi, Divya GC Beena DB.

- Evaluation of *in vitro* thrombolytic activity of phytochemicals in *Bacopa monnieri* Linn. Journal of Pharmacy Research. 2012; 5(1):100-101.
25. Islam MA, Mahmud ZA, Rahman SMA, Md. Monirujjaman, Saha SK *et al.* Evaluation of Thrombolytic activity and Brine Shrimp Lethality Bioassay of Methanol extract of stems of *Tinospora crispa*. International Journal of Pharmaceutical Sciences and Research. 2013; 4(3):1148-1153.
  26. Sultana I, Noor MA, Barua J, Mahmood A, Das MC, Ibrahim MM, *et al.* In-vitro antiatherothrombosis activity of four Bangladeshi plants. International. Journal of Green Pharmacy. 2012; 6(1):5-8.
  27. Randy C, Ploetz, Angela Kay Kepler, Jeff Daniells, Scot C, Nelson Banana. Plantain-an overview with emphasis on Pacific island cultivars-Musaceae (banana family); Species Profiles for Pacific Island Agro forestry. 2007; 1:1-27.
  28. Khare CP. Indian Medicinal Plants. Berlin, Springer, 2007, 426.
  29. Kirtikar KR, Basu BD. Indian Medicinal Plant. Edn 3-4, Periodical Experts Book Agency, Delhi, 1991, 2452-2456.
  30. Gupta S, Garg VK, Sharma PK, Singh A. Analgesic activity of aqueous extract of *Musa paradisiaca*. Der Pharmacia Sinica. 2011; 2(4):74-77.
  31. Savali AS, Bhinge SD, Chitapurkar HR. Evaluation of hair growth promoting activity of *Musa paradisiaca* unripe fruit extract. Journal of Natural Pharmaceuticals. 2011; 3:120-124.
  32. Hallikeri CS, Suresh HM, Chandur VK, Bhoomannavar VS, Shivakumar SI, Hatapakki BC *et al.* Anticonvulsant effect of the unripe fruits of *Musa paradisiaca* in albino rats. Phytopharmacology and therapeutic values 2008, 433-438.
  33. Richter ER, Vore LA. Antimicrobial activity of banana puree. Food Microbiol. 1989; 6:179-187.
  34. Ahmad I, Beg AZ. Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drug resistant human pathogens. J. Ethnopharmacol. 2001; 74:113-123.
  35. Mokbel MS, Hashinaga F. Antibacterial and Antioxidant Activities of Banana (*Musa*, AAA cv. Cavendish) Fruits Peel. Am J. Biochem Biotechnol. 2005; 1(3):125-131.
  36. Alisi CS, Nwyanwu CE, Akujobi CO, Ibegbulem CO. Inhibition of dehydrogenase activity in pathogenic bacteria isolates by aqueous extracts of *Musa paradisiaca* (var. *sapientum*). Afr J. Biotechnology. 2008; 7(12):1821-1825.
  37. Fagbemi JF, Ugoji E, Adenipekun T, Adelowotan O. Evaluation of the antimicrobial properties of unripe banana (*Musa sapientum* L.), lemon grass (*Cymbopogon citratus* S.) and turmeric (*Curcuma longa* L.) on pathogens. Afr J. Biotechnol. 2009; 8(7):1176-1182.
  38. Jahan M, Warsi MK, Khatoon F. Concentration influence on antimicrobial activity of banana blossom extract-incorporated chitosan-polyethylene glycol (CS-PEG) blended film. J Chem Pharm Res. 2010; 2(5):373-378.
  39. Rahman MM, Hossain MA, Siddique SA, Biplob KP, Uddin MH. Antihyperglycemic, antioxidant, and cytotoxic activities of *Alocasia macrorrhizos* (L.) rhizome extract. Turk J Biol. 2012; 36:574-579
  40. Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Daginawala HF. Development of an *in vitro* model to study clot lysis activity of thrombolytic drugs. Thromb J. 2006; 4(14):1-4.
  41. Handin RI, Kasper DL, Braunwald E, Fauci AS. Harrison's Principles of Internal Medicine: bleeding and thrombosis. 16th ed., 2005.
  42. Krishnaraju AV, Rao TVN, Sundararaju D, Vanisree M, Tsay HS, Subbaraju GV. Assessment of bioactivity of Indian medicinal plants using brine shrimp (*Artemia salina*) lethality assay. Int. J Appl Sci Eng. 2005; 3(2):125-134.